

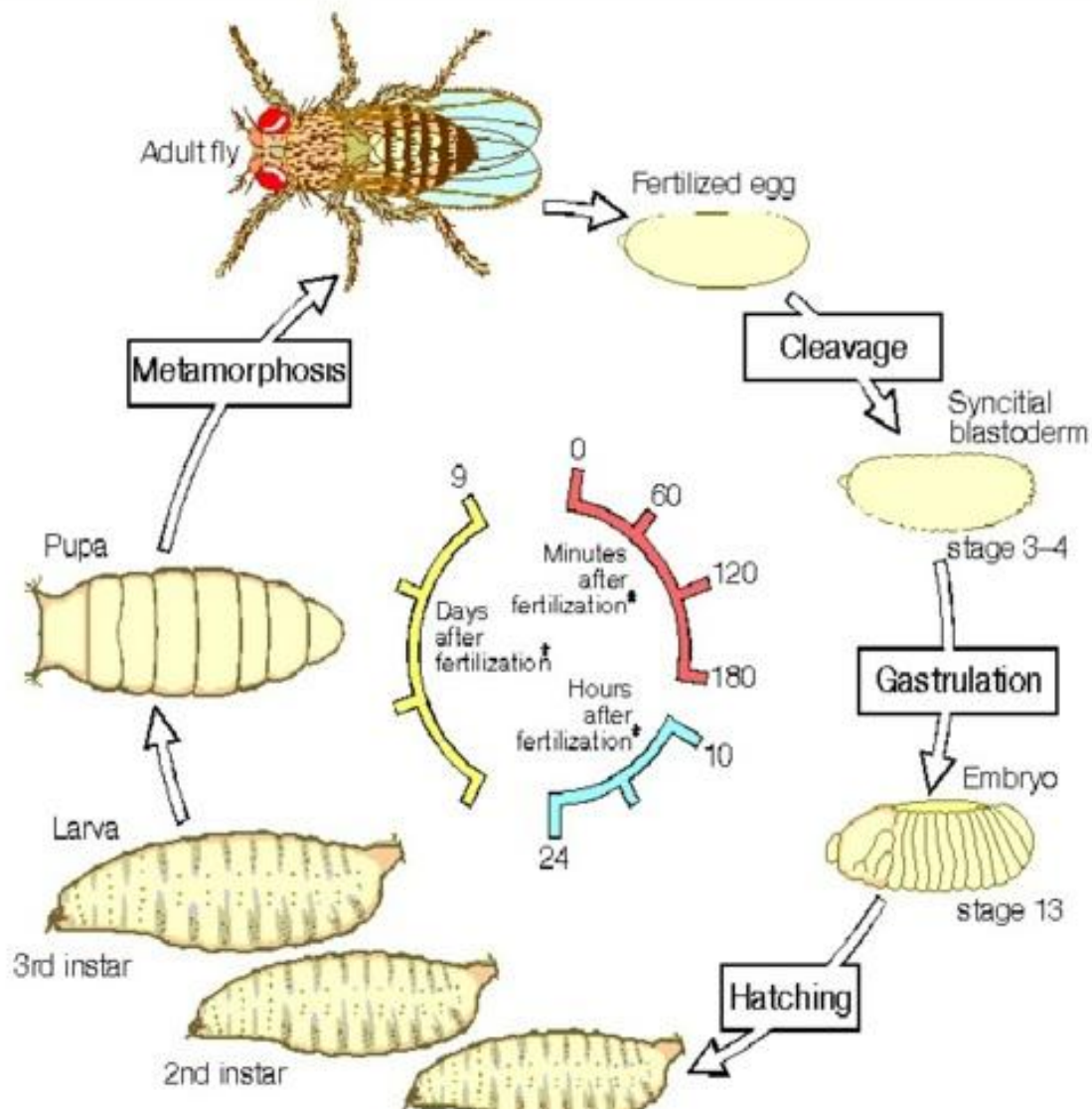
Эволюционная иммунология

Лекция 4 «Иммунитет дрософилы»

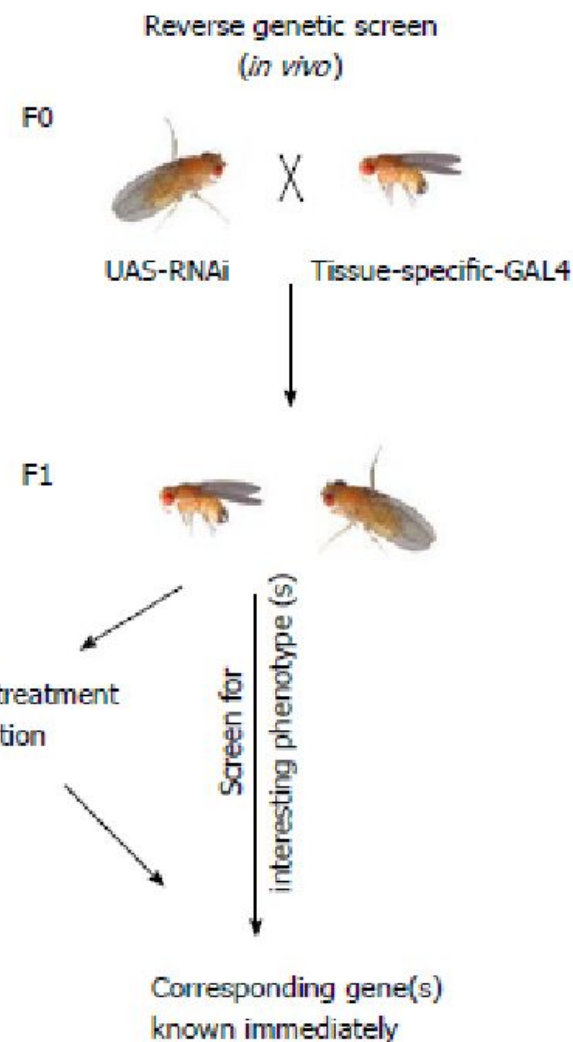
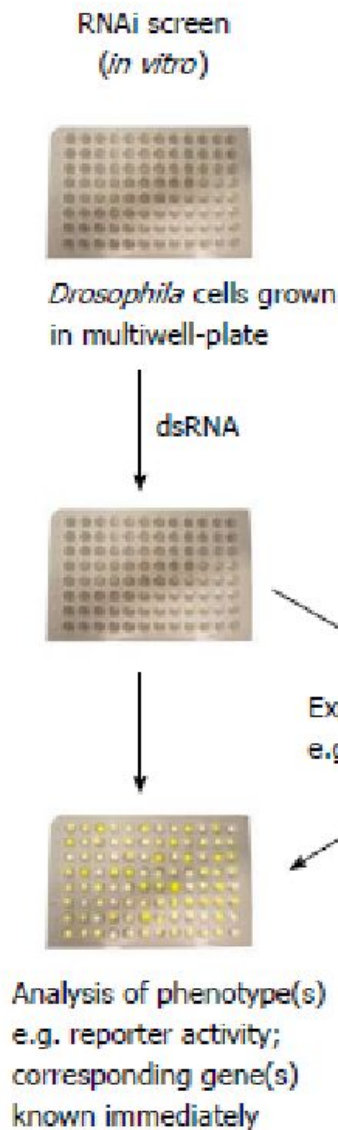
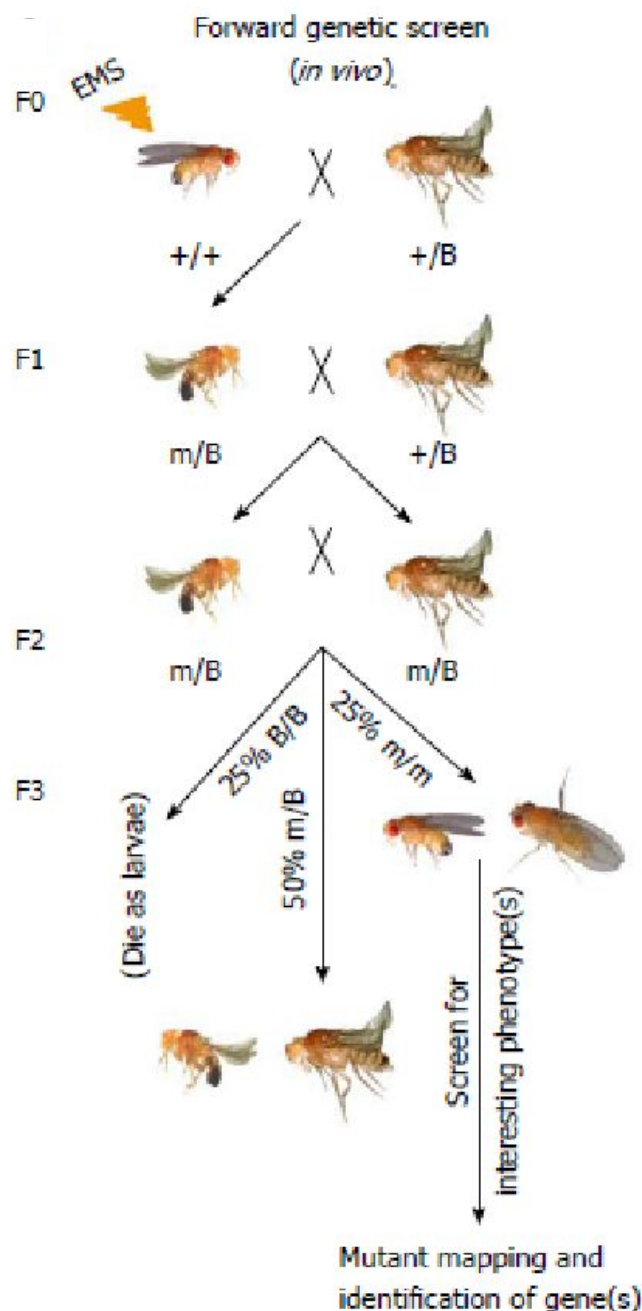


Шилов Е.С.
5 марта МГУ, 2018

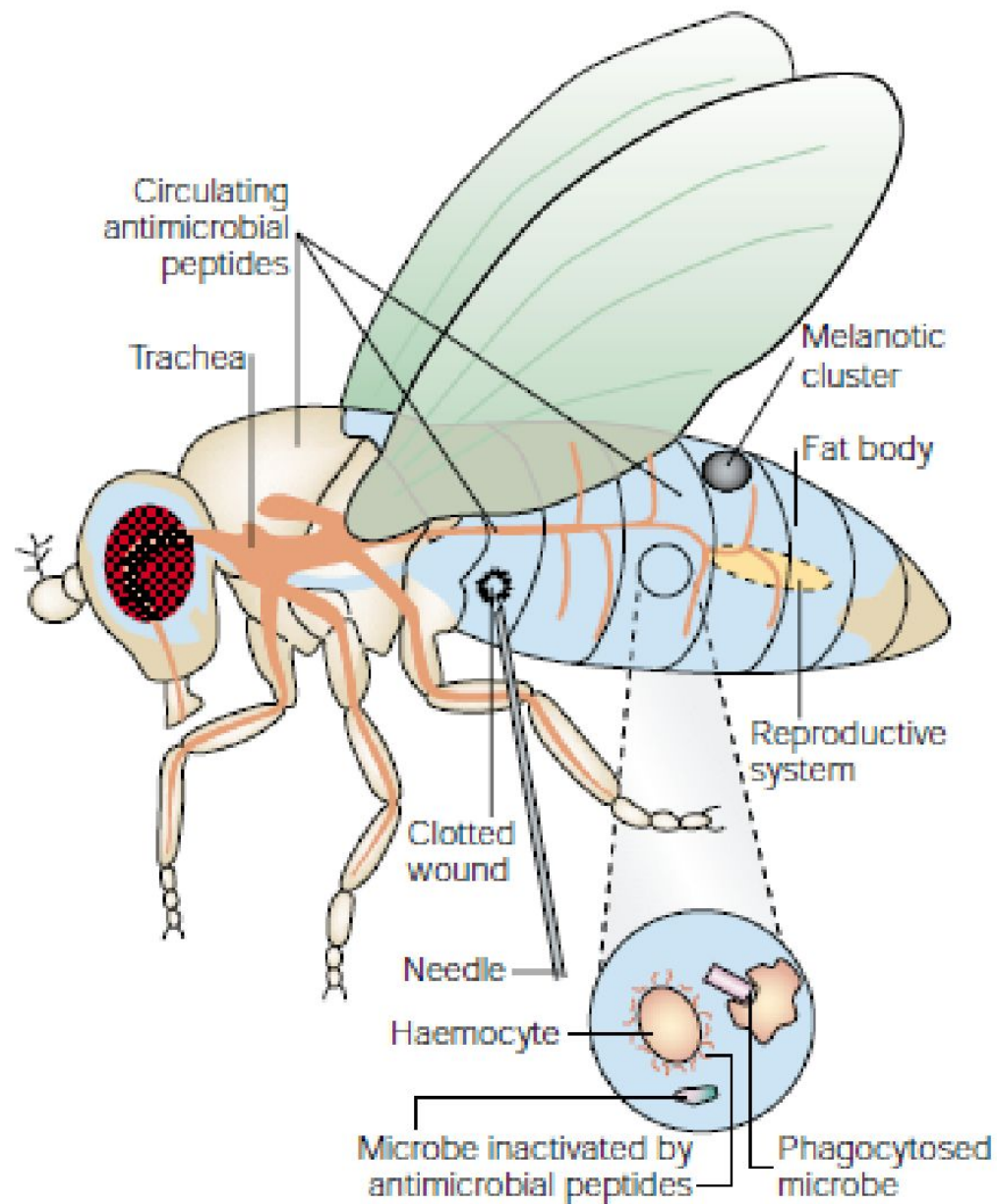
Жизненный цикл дрозофилы



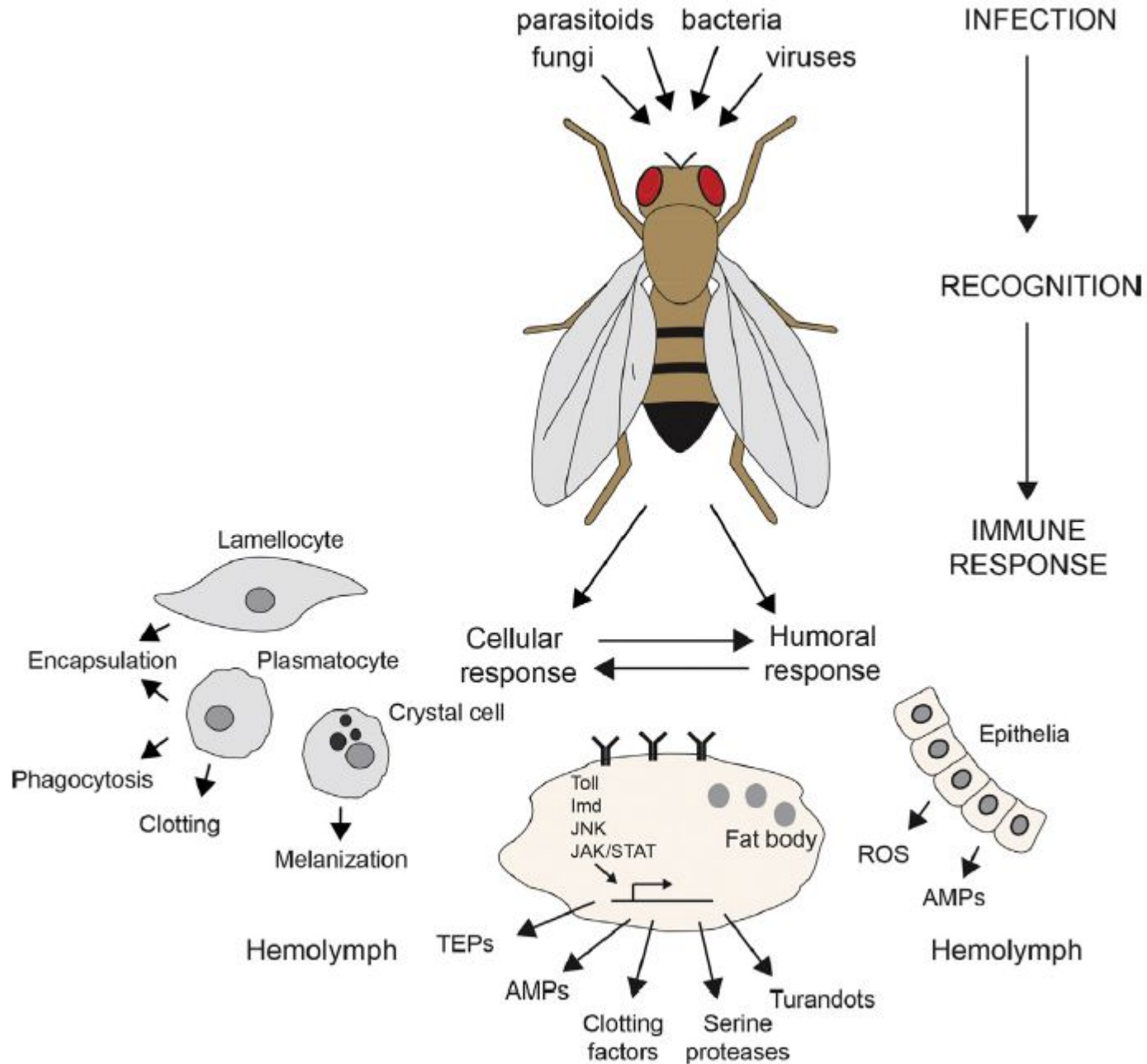
Генетические подходы, применимые к дрозофиле



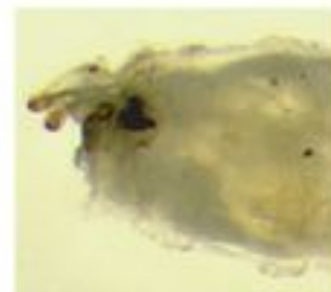
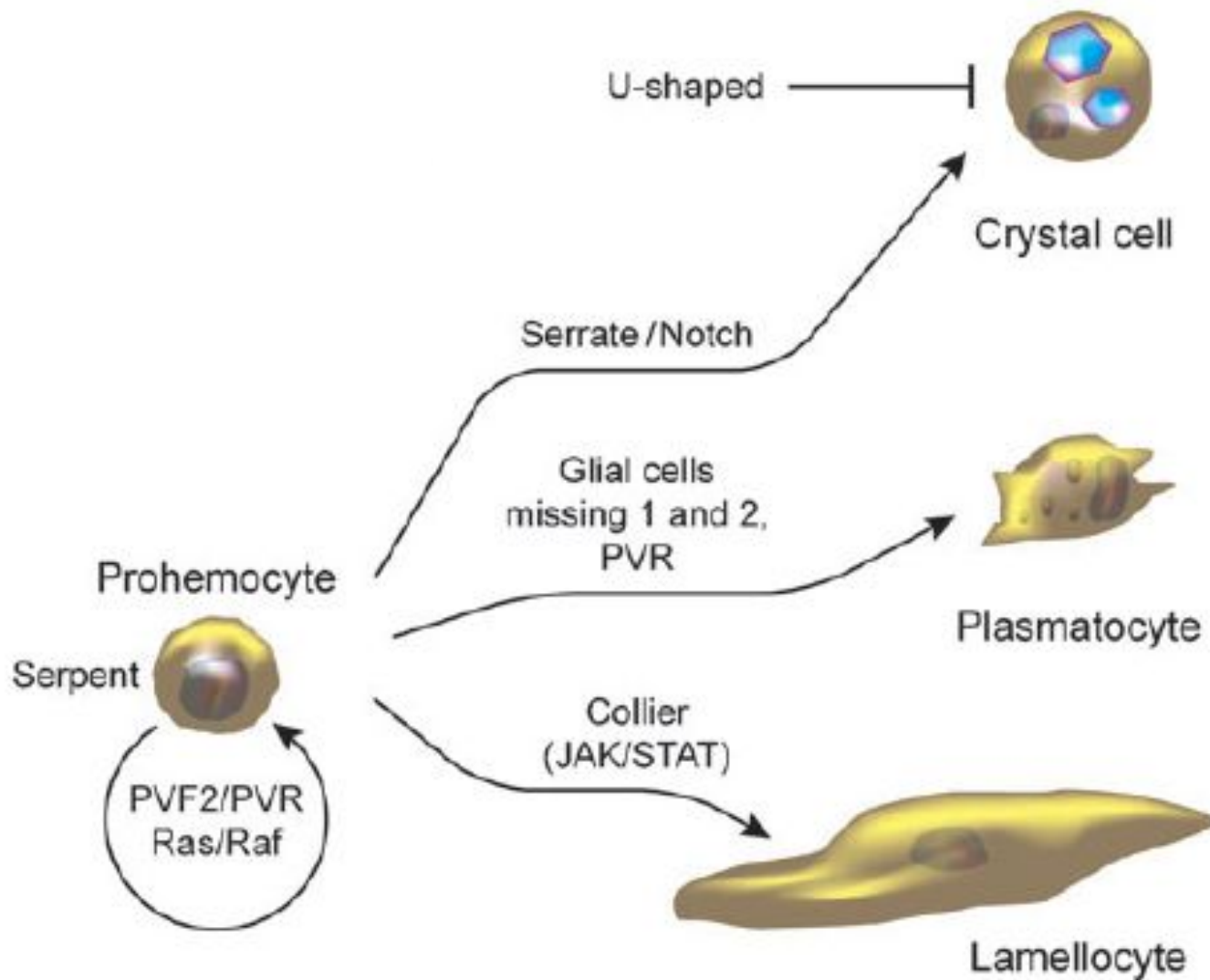
Иммунные реакции дрозодилы



Разнообразие иммунных реакций дрозифилы



Типы иммунных клеток личинки дрозофилы



Melanization



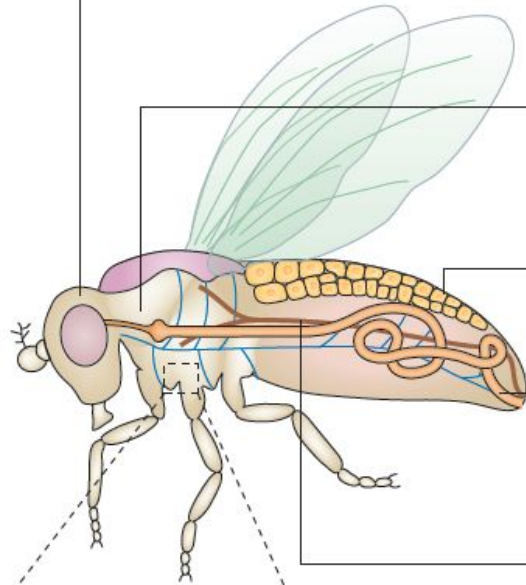
Phagocytosis



Encapsulation

Сравнение иммунитета насекомых и млекопитающих

Drosophila melanogaster



Central nervous system

- Production of AMPs and/or cytokines
- Inflammation
- Neuronal death and degeneration

Respiratory system

- (Trachea in flies and lungs in humans)
- Production of AMPs

Systemic response

- (Fat body in flies and liver in humans)
- Production of AMPs
 - Acute phase response

Digestive system

- (Gut in flies and humans)
- Production of AMPs
 - Local ROS production via Duox and Nox

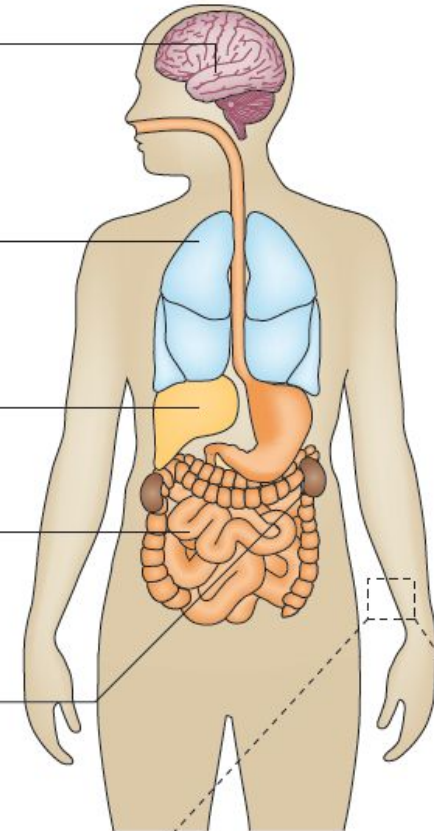
Excretory system

- (Malpighian tubules in flies and kidneys in humans)
- Production of AMPs
 - Hormonal regulation

Cellular response

- (Haemolymph in flies, and blood and lymph in humans)
- Phagocytosis
 - Clotting and coagulation
 - Cytokine secretion

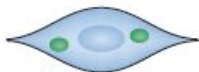
Human



Plasmatocyte (phagocytosis)



Lamellocyte (encapsulation)



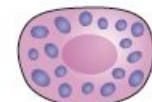
Crystal cell (melanization and clotting)



Macrophage

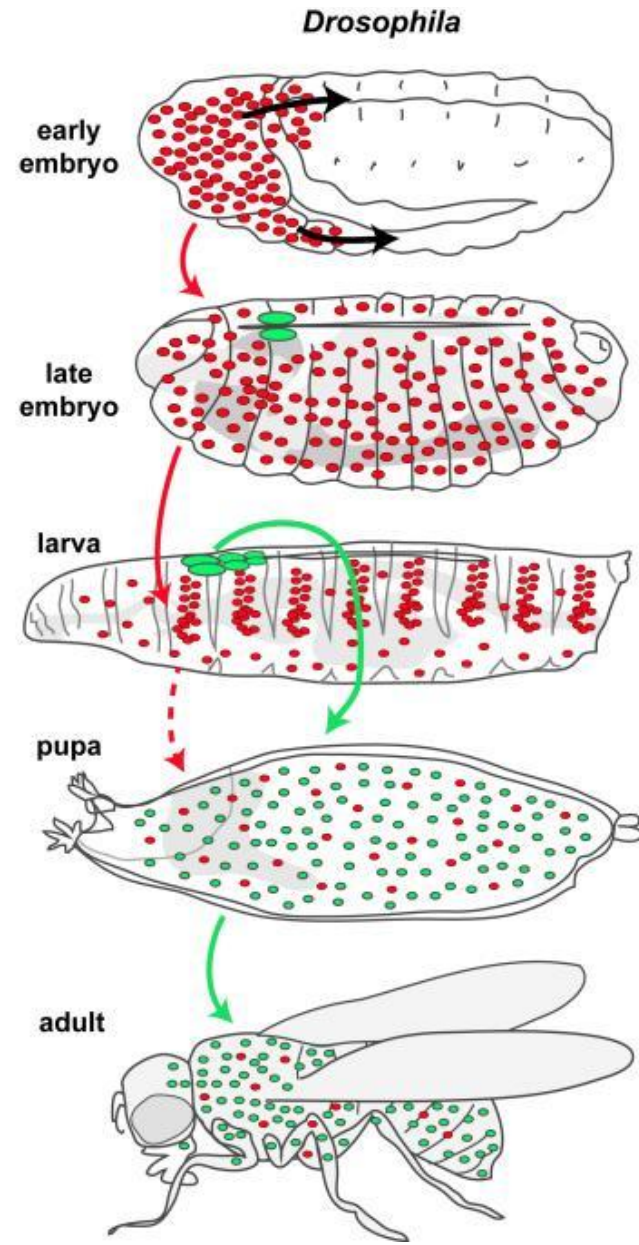
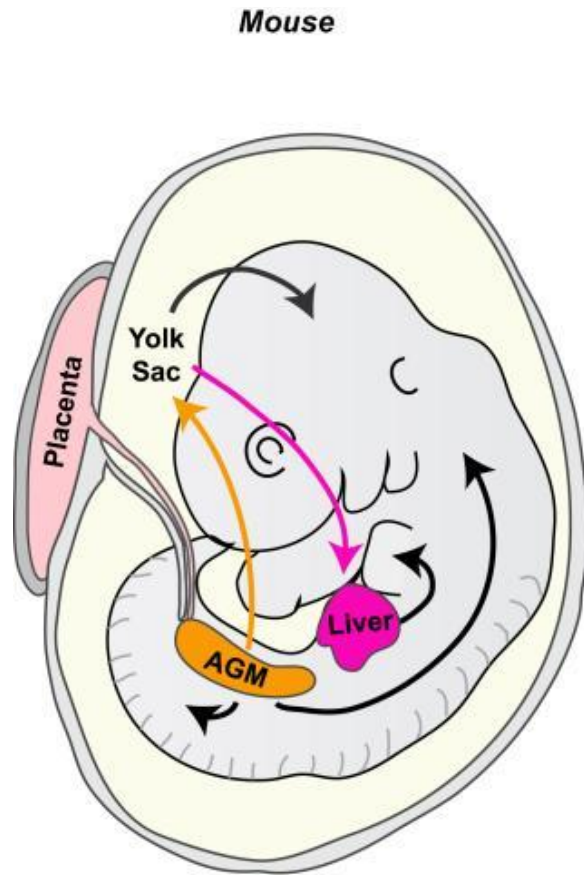


Natural killer cell

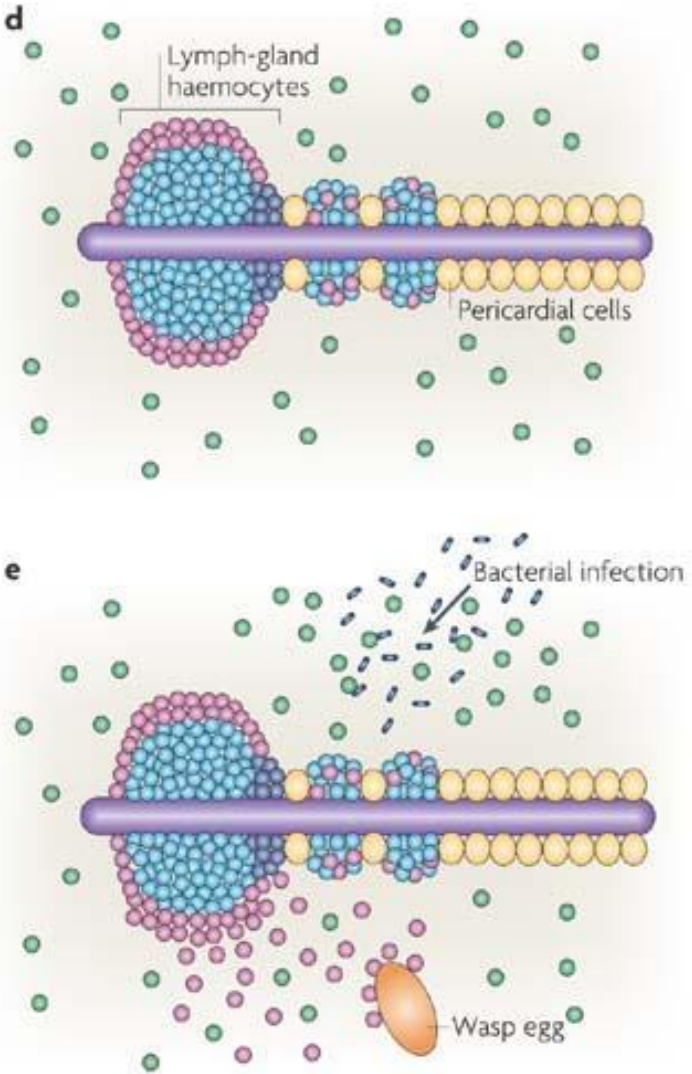
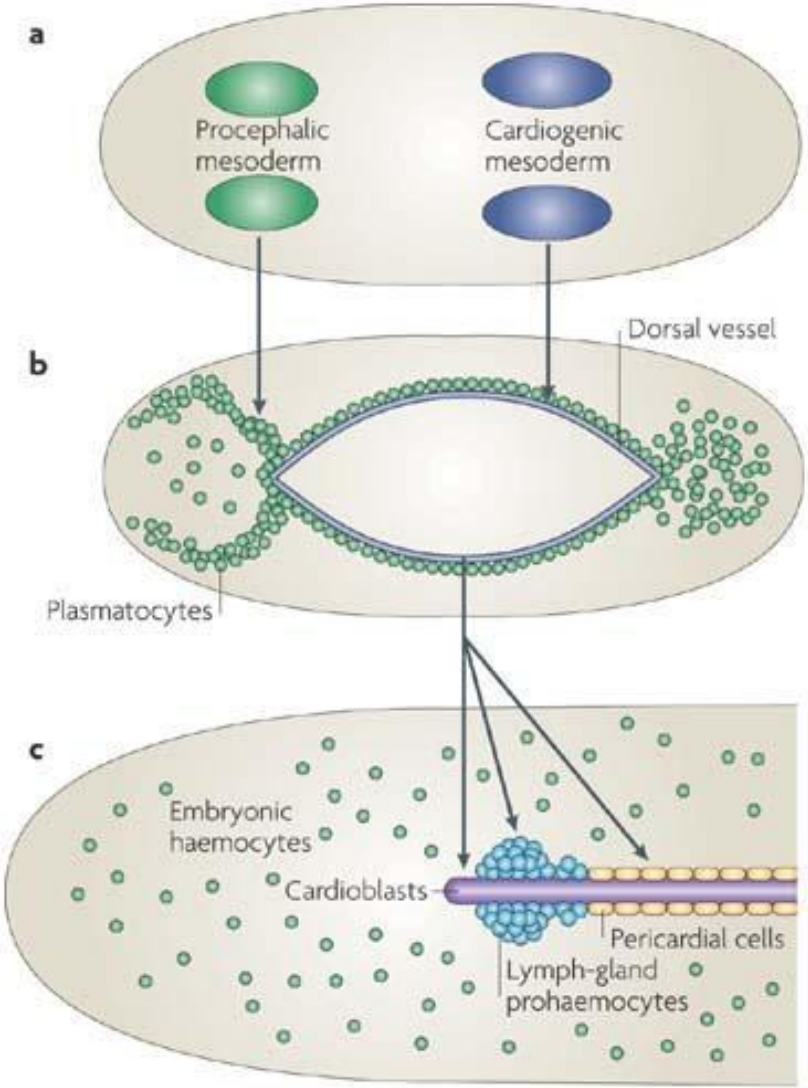


Granulocyte

Сравнительный гематопозез

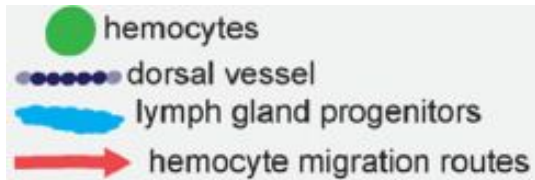
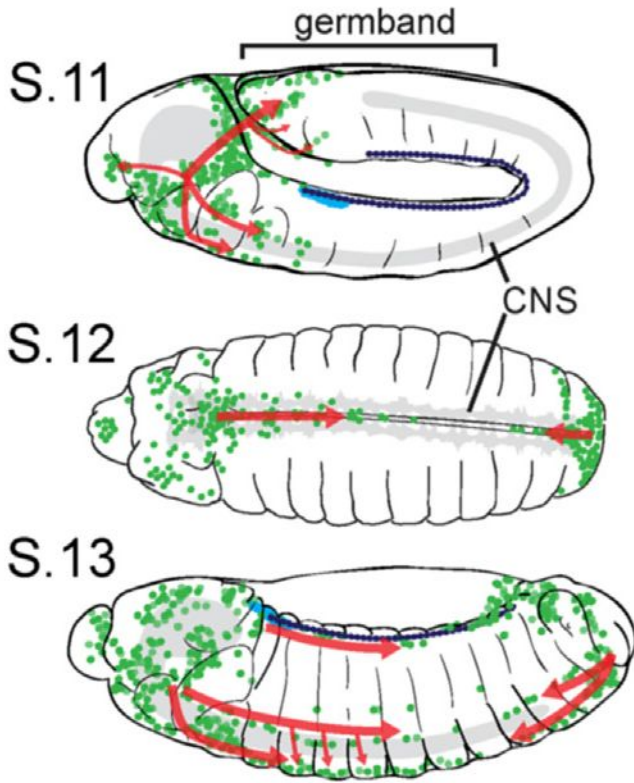


Происхождение гемоцитов дрозofilы

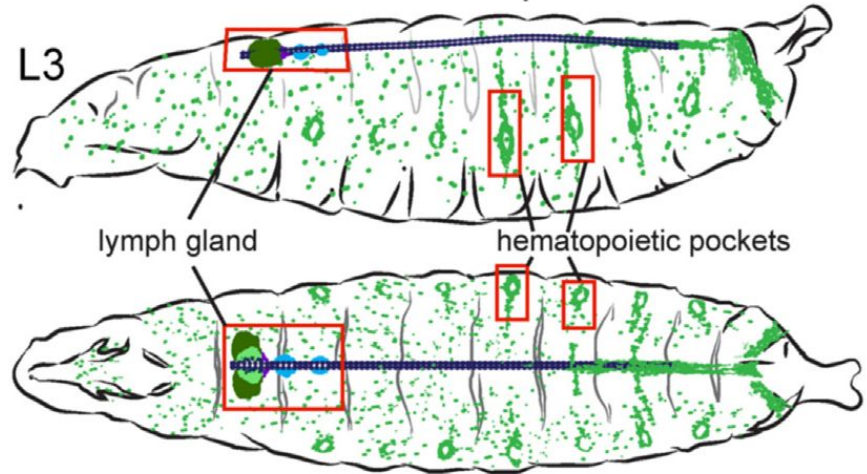


Гематопоз у имаго дрозофилы

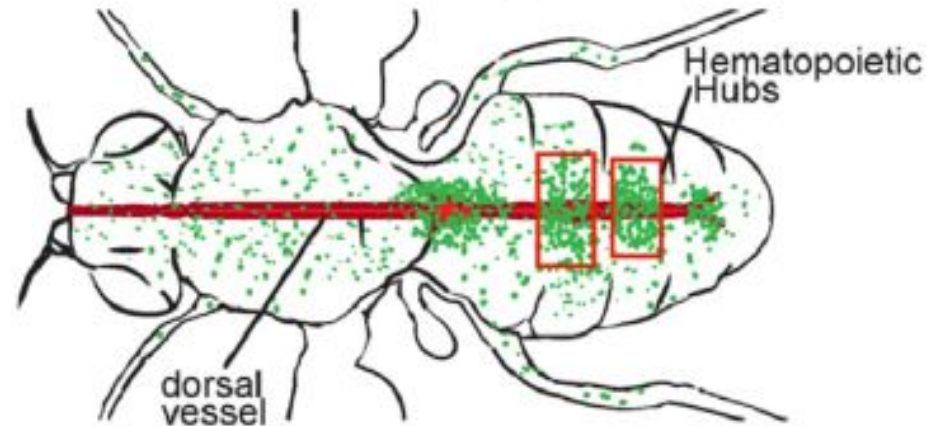
Embryonic Hematopoiesis



Larval Hematopoiesis

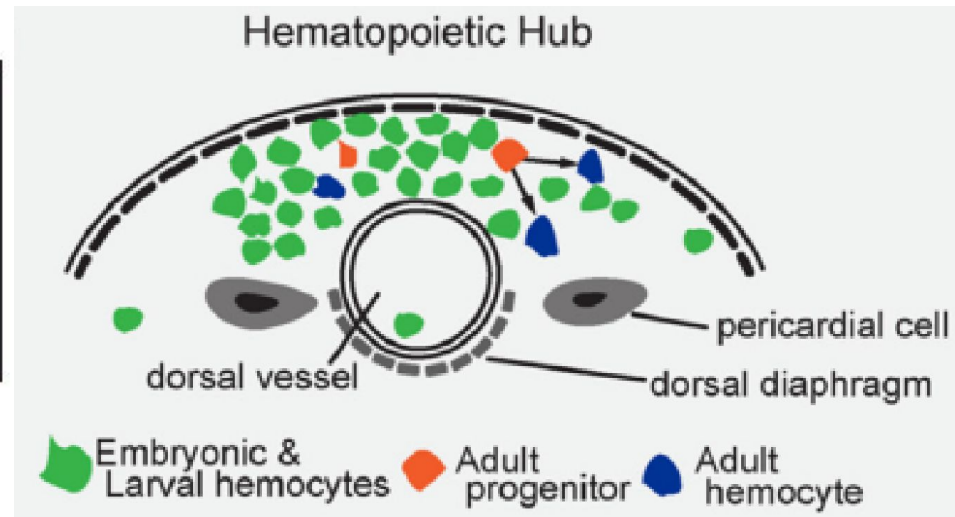
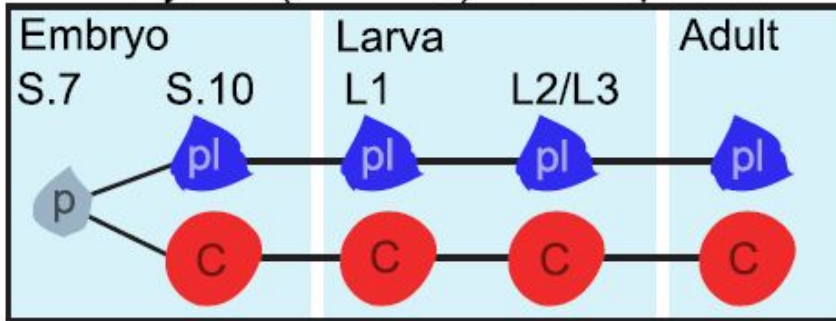


Adult Hematopoiesis

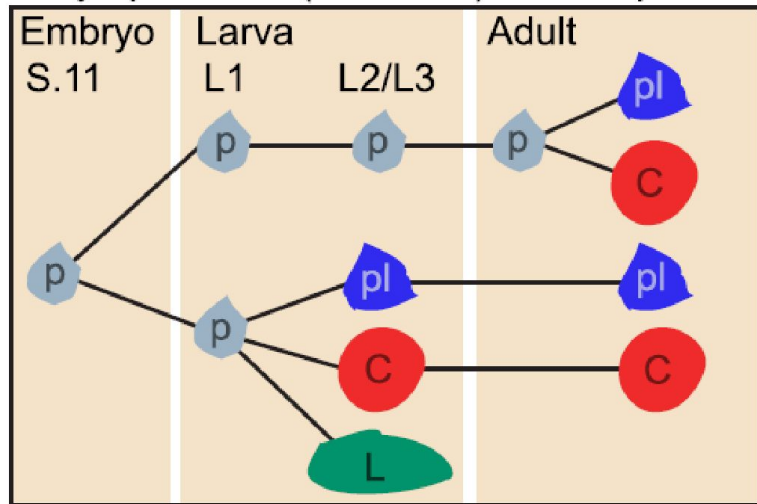


Гематопоз дрозодилы

A Embryonic (Primitive) Hematopoiesis

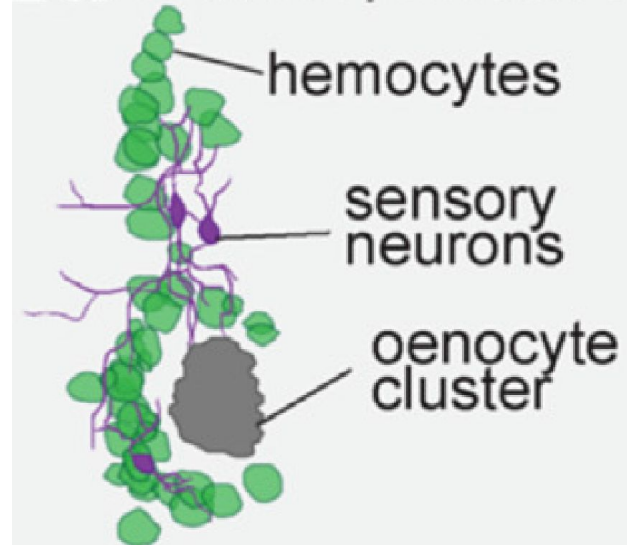


Lymph Gland (Definitive) Hematopoiesis



p prohemocyte pl plasmatocyte
 C crystal cell L lamellocyte

Bii Hematopoietic Pocket



Коагуляция гемолимфы у дрозофилы

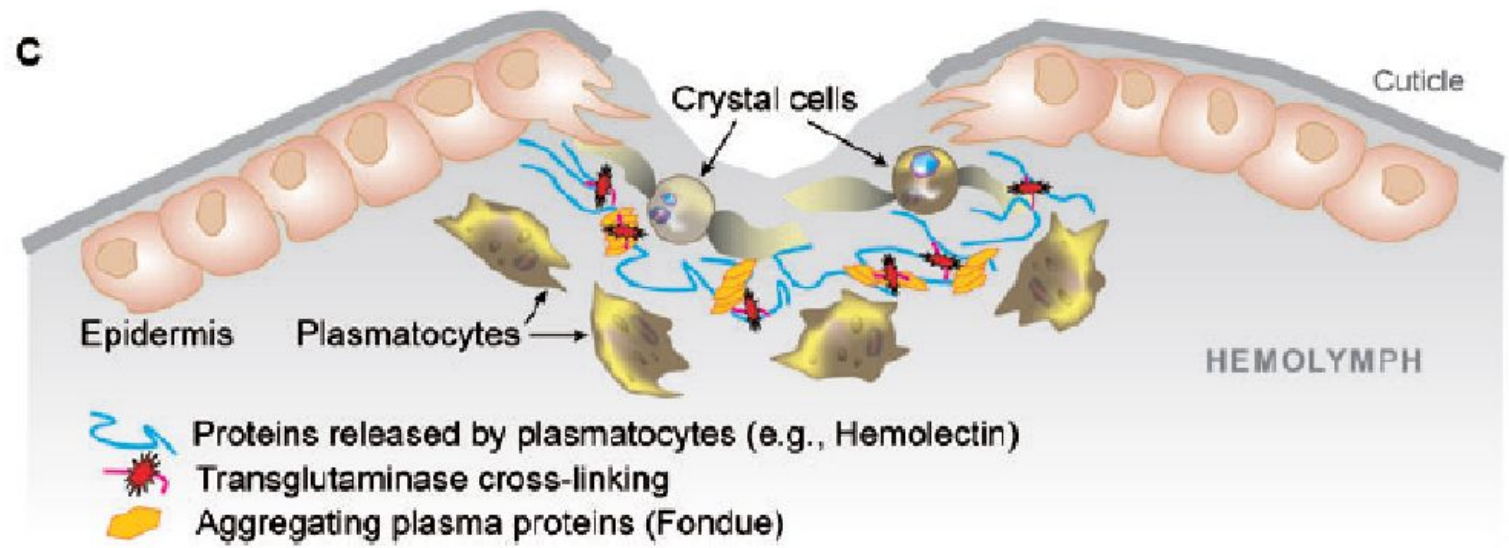
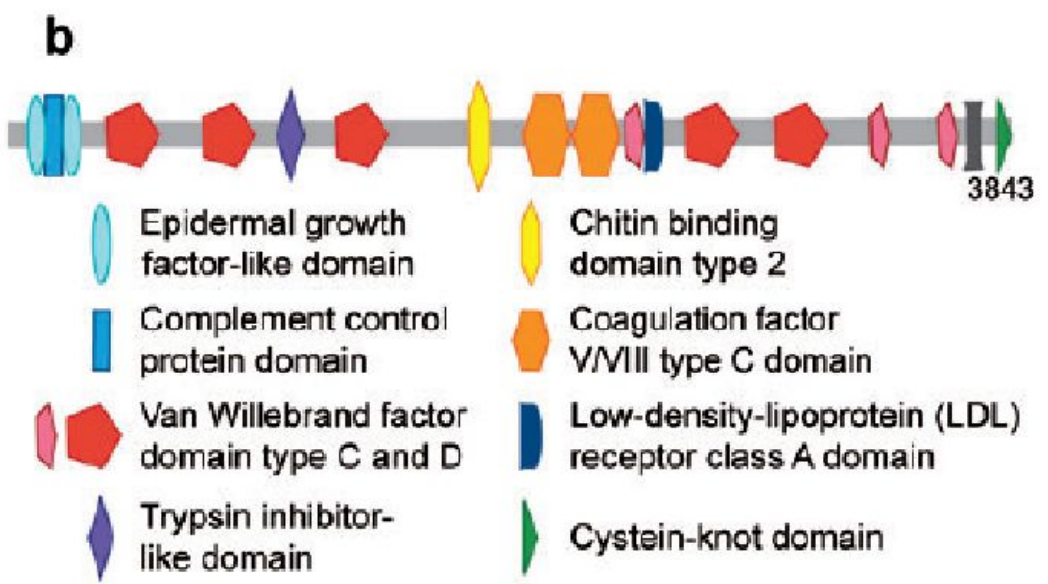
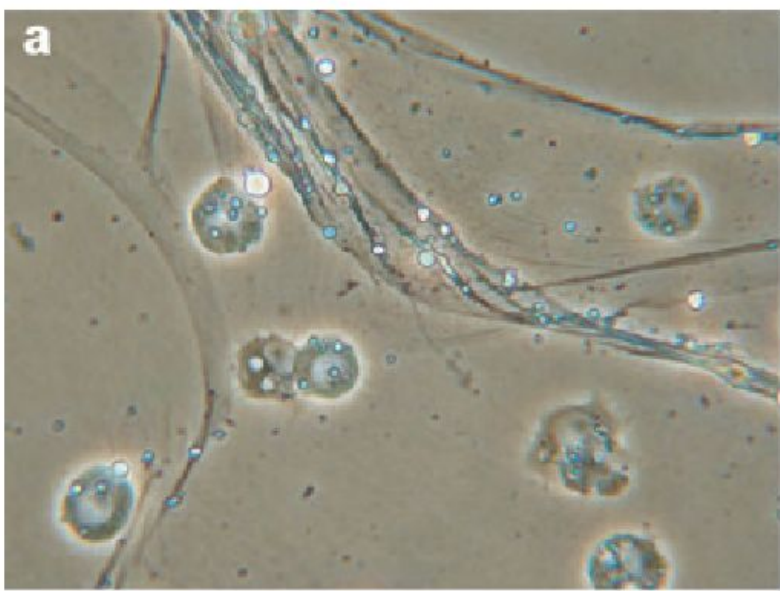
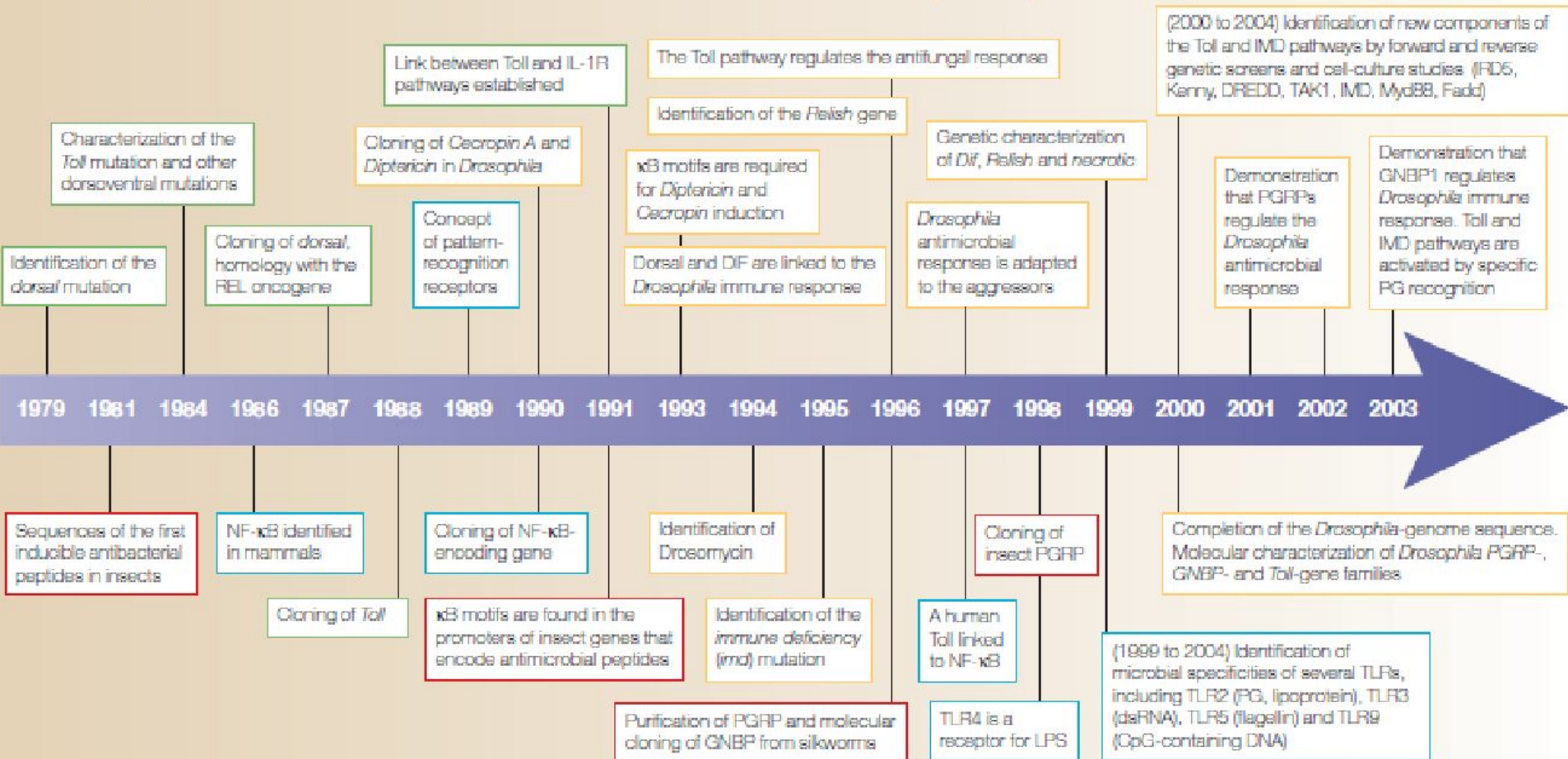


Table 1 | Immune-related genes in *Drosophila* and mammals: homologies and analogies

| | <i>Drosophila</i> | Mammals* | References [§] |
|--|--|---|-----------------------------|
| Haematopoietic determinants | | | |
| Transcription factors | <i>lozenge</i> <i>serpent</i> <i>gcm</i> | <i>AML1</i> Gata family Gcm family | 98 98 98 |
| Receptors | | | |
| Scavenger receptors (SR) | <i>epithelial membrane protein</i> <i>SR-C1</i> | <i>CD36</i> Domains of <i>SRC1</i> have homology to various mammalian proteins; see also SR family | 99 5,100 |
| Gram-negative-binding proteins (GNBP) | GNBP family | Not identified | 11 |
| Peptidoglycan-recognition proteins (PGRP) | PGRP family | Pglyrp family | 10 |
| Lectins | <i>galactin</i> | Lgals family | 55 |
| Toll family | See text | See text | See text |
| Toll signalling components | | | |
| | <i>dorsal</i> , <i>Dif</i> , <i>Rel</i> <i>cactus</i> <i>spätzle</i> | NF- κ B family I κ B family <i>Bdnf</i> ; homology is to cysteine knot nerve growth factors | 17–19,101 102 103 |
| | <i>Spn43Ac</i> of <i>necrotic</i> locus | <i>Spi17</i> and others that encode serine protease inhibitors of the serpin family | 24 |
| | <i>gastrulation defective</i> | <i>F2/prothrombin</i> and others that encode clotting factors | 104,105 |
| | <i>pelle</i> <i>snake</i> and <i>easter</i> <i>IKK</i> and <i>kenny</i> | <i>IRAK1</i> <i>Prss7</i> and <i>KIK/kallikrein</i> family IKK family | 106 105 21,22 |
| Signalling pathways other than Toll | | | |
| | <i>hopscotch</i> <i>Stat92E/mareille</i> <i>raf</i> <i>basket</i> <i>p38b</i> | JAK family STAT family <i>Raf1</i> JNK family <i>p38</i> | 64 107 108 109 |
| Effectors | | | |
| Immunoglobulin motif | Various in <i>Drosophila</i> , but only reported insect immune related is <i>hemolin</i> of silkworm | Immunoglobulin superfamily, including some <i>TIR</i> -related genes | 110 |
| Antimicrobial proteins | Antimicrobial protein gene families | Antimicrobial protein gene families | 54,56,57,59 |
| Metal ion accessibility and metalloproteinase regulation | <i>malvolio</i> <i>Transferrin</i> Other matrix metalloproteinase members, for example, <i>kuzbarian</i> | <i>Nramp1/Scf11</i> <i>Transferrin</i> <i>Mmp7/matrixlysin</i> | 111 112 113 |
| Translation related | <i>Thor</i> <i>lethal(1)aberrant immune response</i> [§] | 4E-binding protein family <i>ribosomal protein S6</i> | 114 66 |
| Cytotoxic molecules | Reactive intermediates of oxygen and nitrogen | Reactive intermediates of oxygen and nitrogen | 115 |
| Apoptosis and immunity | | | |
| | <i>croquemort</i> <i>dredd</i> | <i>CD36</i> CASP family | 6 90–92 |
| Additional components | | | |
| | Tep family Various from genome annotation | C3 and A2M For example, genes that encode C1 and fibrinogen domains | 65,72 13,84 |

Хронология изучения иммунитета мушки

Timeline | Discoveries relevant to the function of the Toll and Imd pathways in *Drosophila* immunity



The discovery of the role of Toll in the *Drosophila* immune response (yellow) was influenced mainly by research in three fields: insect immunity (red), vertebrate immunology and signalling (blue) and developmental genetics (green). DIF, Dorsal-related immunity factor; DREDD, Death-related ced-3/Nedd2-like protein; ds, double-stranded; GNBPs, Gram-negative-bacteria-binding protein; I κ B, inhibitor of NF- κ B; IKK, I κ B kinase; IL-1R, interleukin-1 receptor; IMD, Immune deficiency; IRD5, Immune-response deficient 5; LPS, lipopolysaccharide; NF- κ B, nuclear factor- κ B; PG, peptidoglycan; PGRP, PG-recognition protein; serpin, serine-protease inhibitor; TAK1, Transforming growth factor- β -activated kinase 1; TLR, Toll-like receptor.



Christiane Nüsslein-Volhard



Kathryn V. Anderson

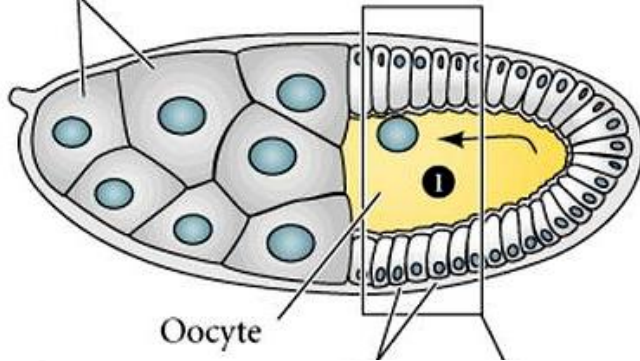


Lemaitre Bruno

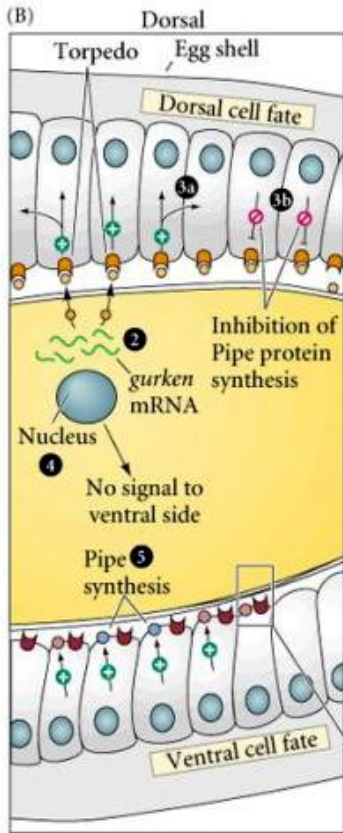


Jules A. Hoffmann

Ovarian nurse cells



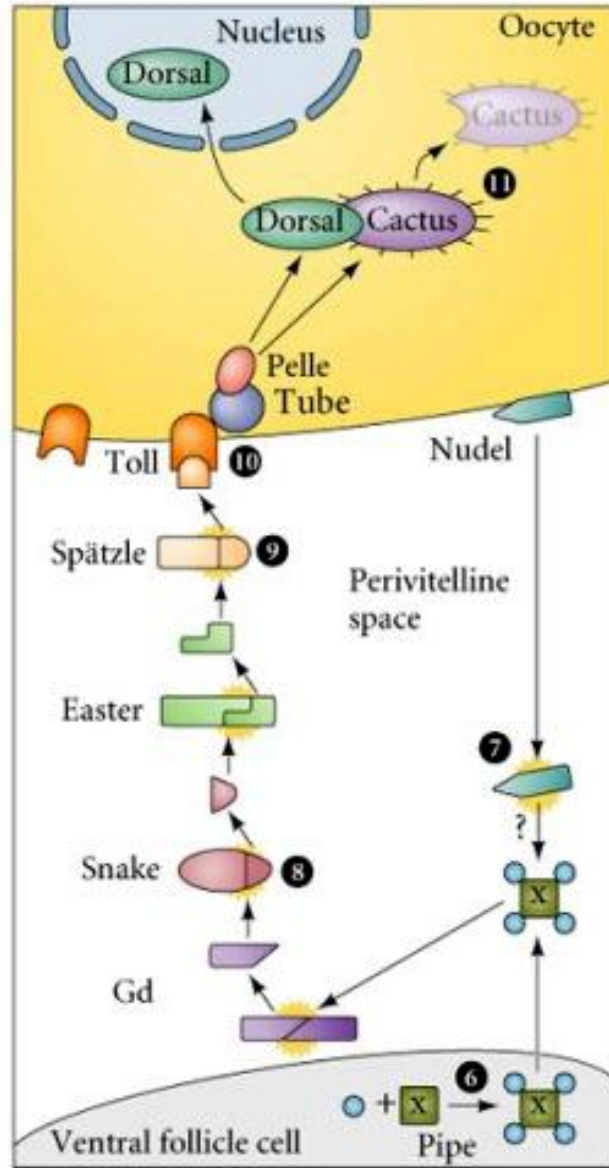
Oocyte



- 2 Gurken protein made by oocyte and received by Torpedo protein
- 3a Torpedo causes dorsal morphology
- 3b Pipe synthesis inhibited
- 4 Gurken does not diffuse
- 5 Ventral follicle cells make Pipe

See Part 3

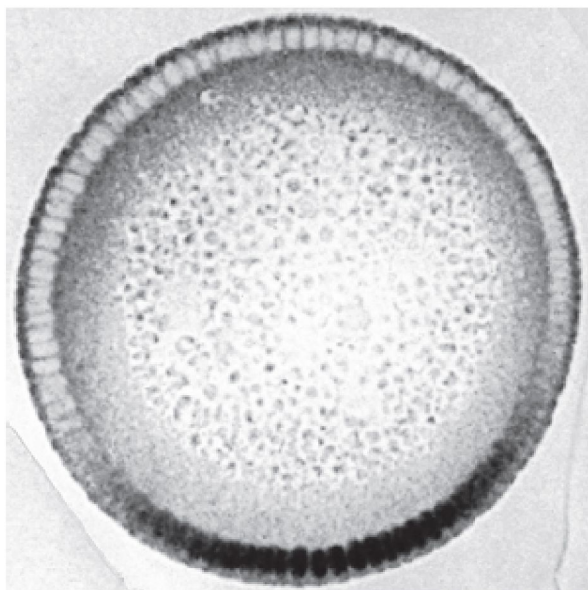
Ventral



(C)

- 6 Pipe modifies (x)
- 7 Nudel and (x) activate Gd
- 8 Gd activates Snake; Snake activates Easter
- 9 Easter activates Spätzle, which binds to Toll
- 10 Toll activates Tube and Pelle, to phosphorylate Cactus
- 11 Cactus is degraded; Dorsal protein enters nucleus

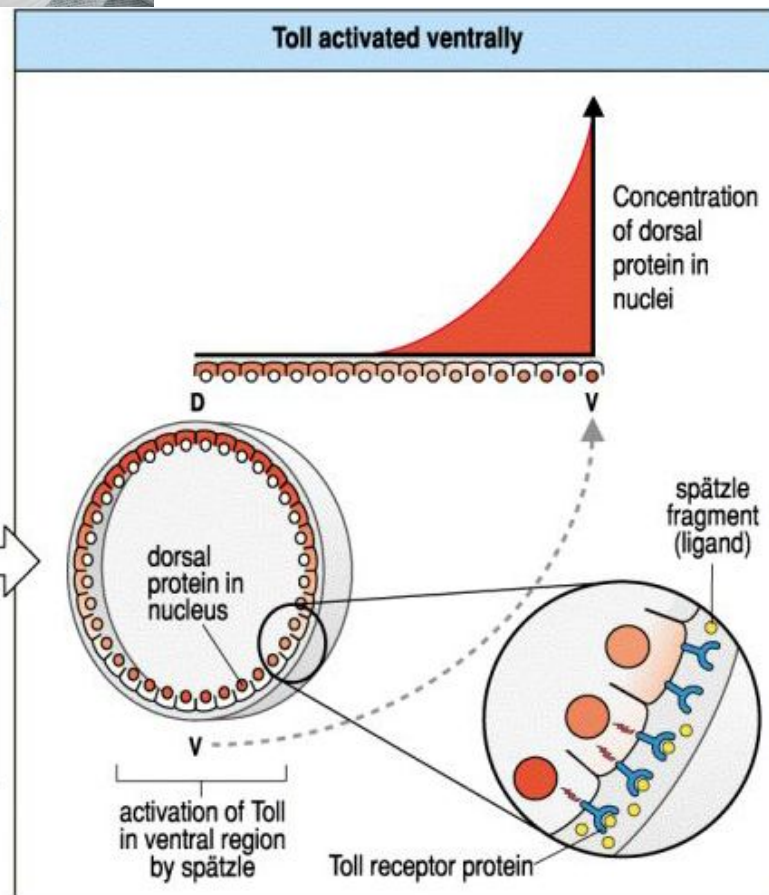
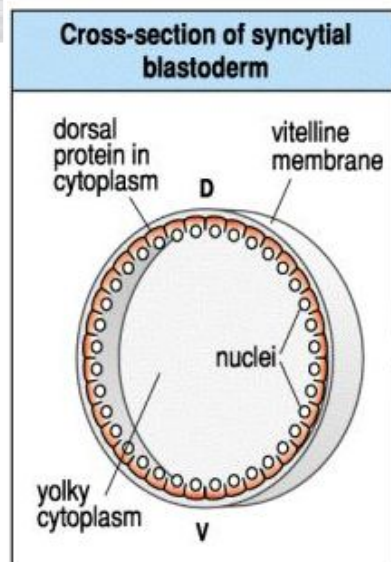
Белок Dorsal в верхней части зародыша мухи попал в ядра, в нижней части – остался в цитоплазме



100 мкм



Spaetzle



NF-κB у дрозофилы: Dorsal, DIF, Relish

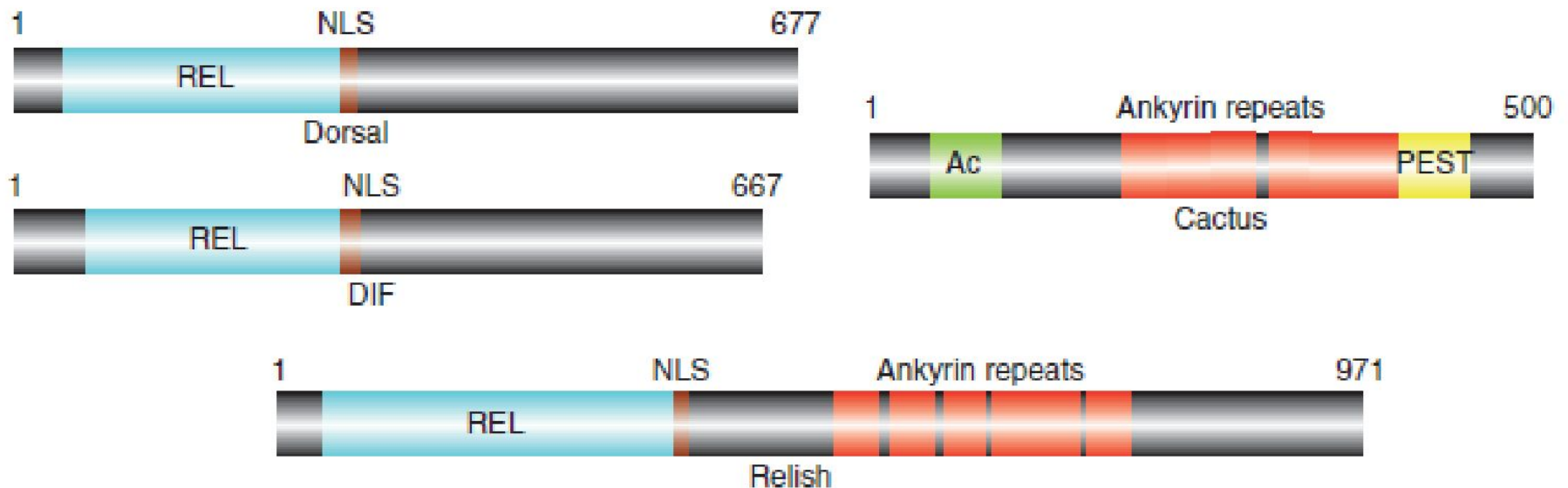
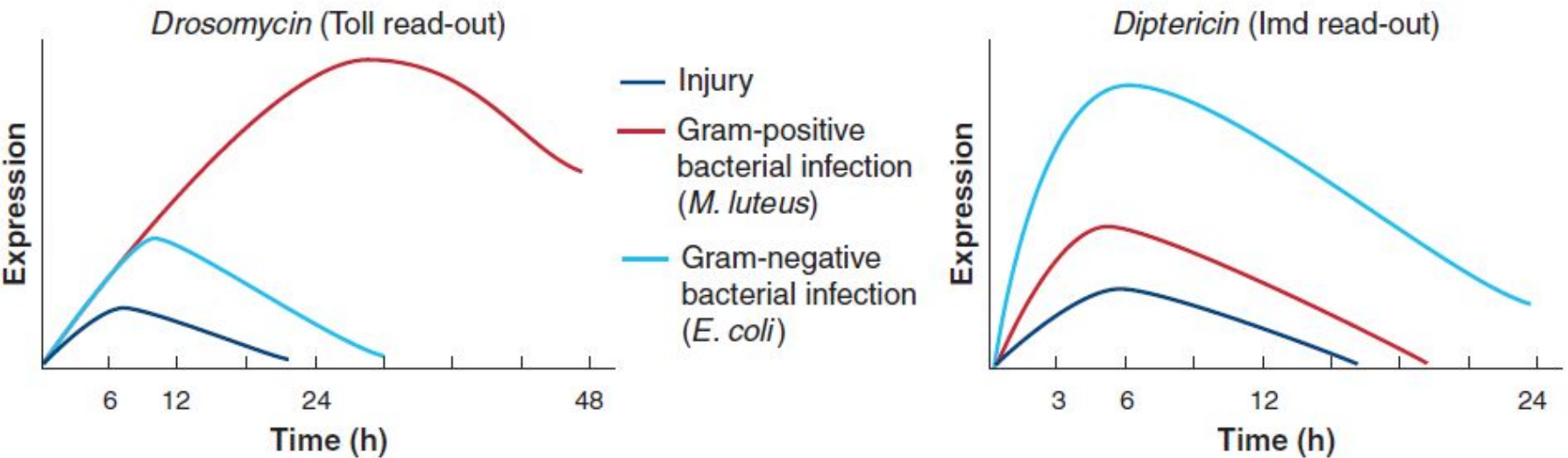
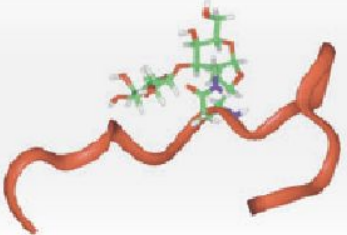
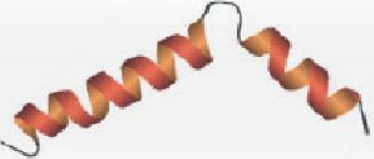
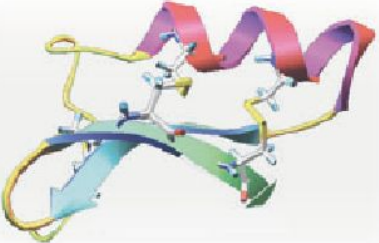
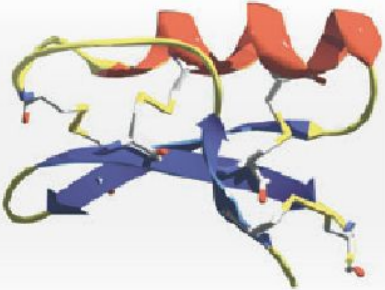


Figure 1. The NF-κB and IκB proteins in *Drosophila*. The length in amino acids is indicated by numbers. REL, Rel-homology domain; NLS, nuclear localization sequence; PEST, proline, glutamic acid, serine, and threonine-rich segment; Ac, acidic domain.

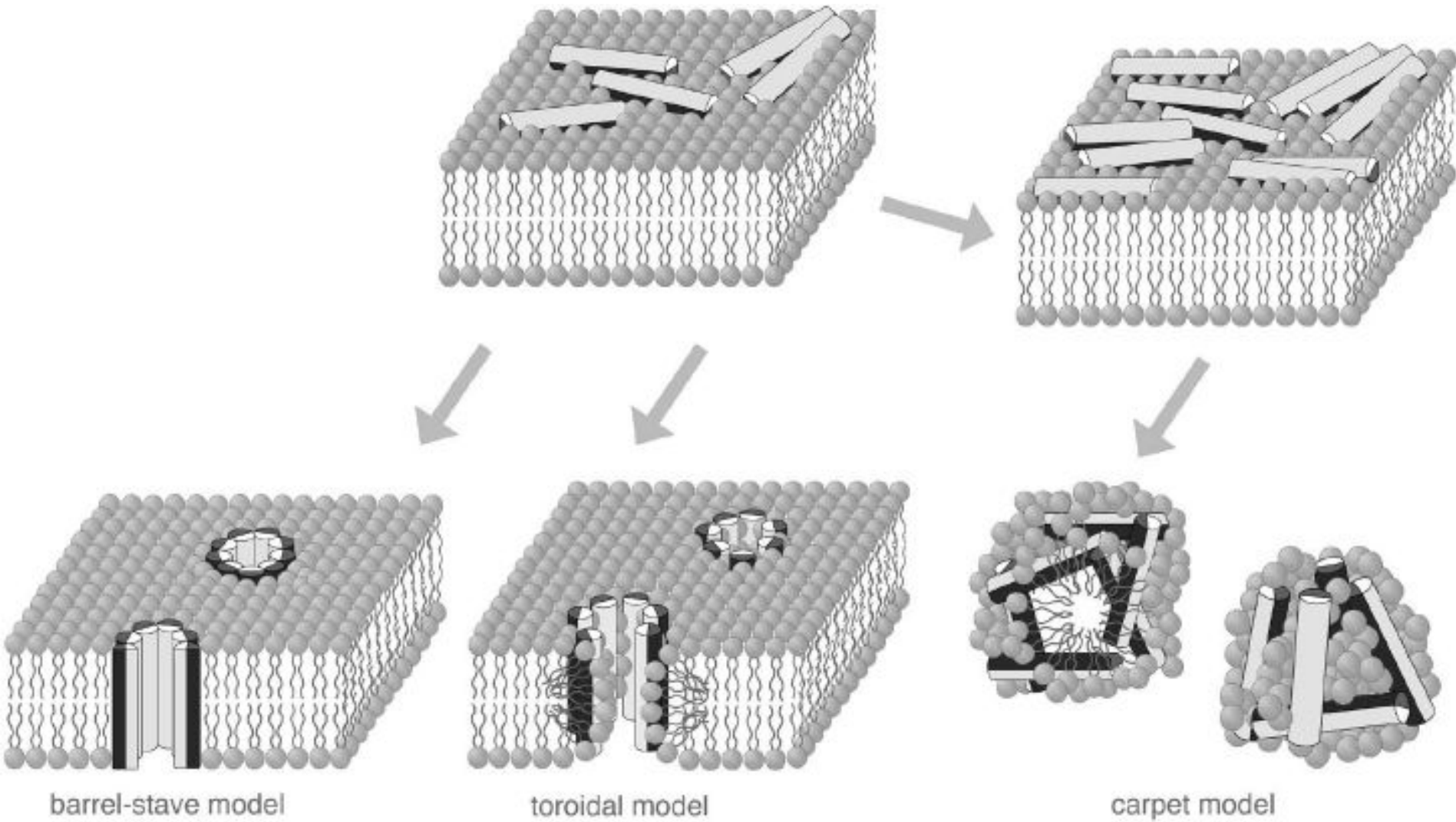
Грамотрицательные и грамположительные бактерии запускают соответственно Imd- и TLR- пути, которые различаются по спектру активируемых антибактериальных пептидов



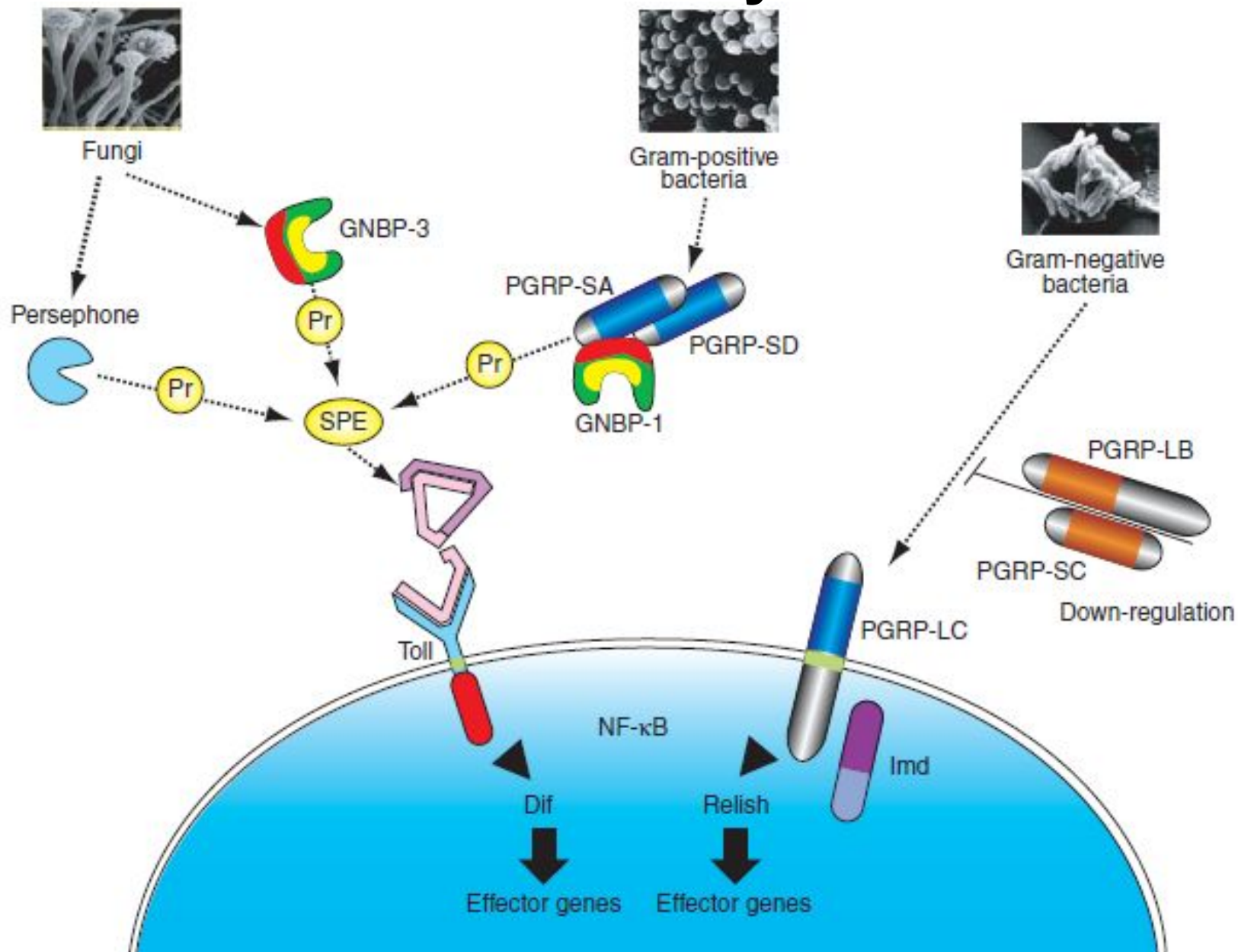
Семь семейств антимикробных пептидов дрозофилы

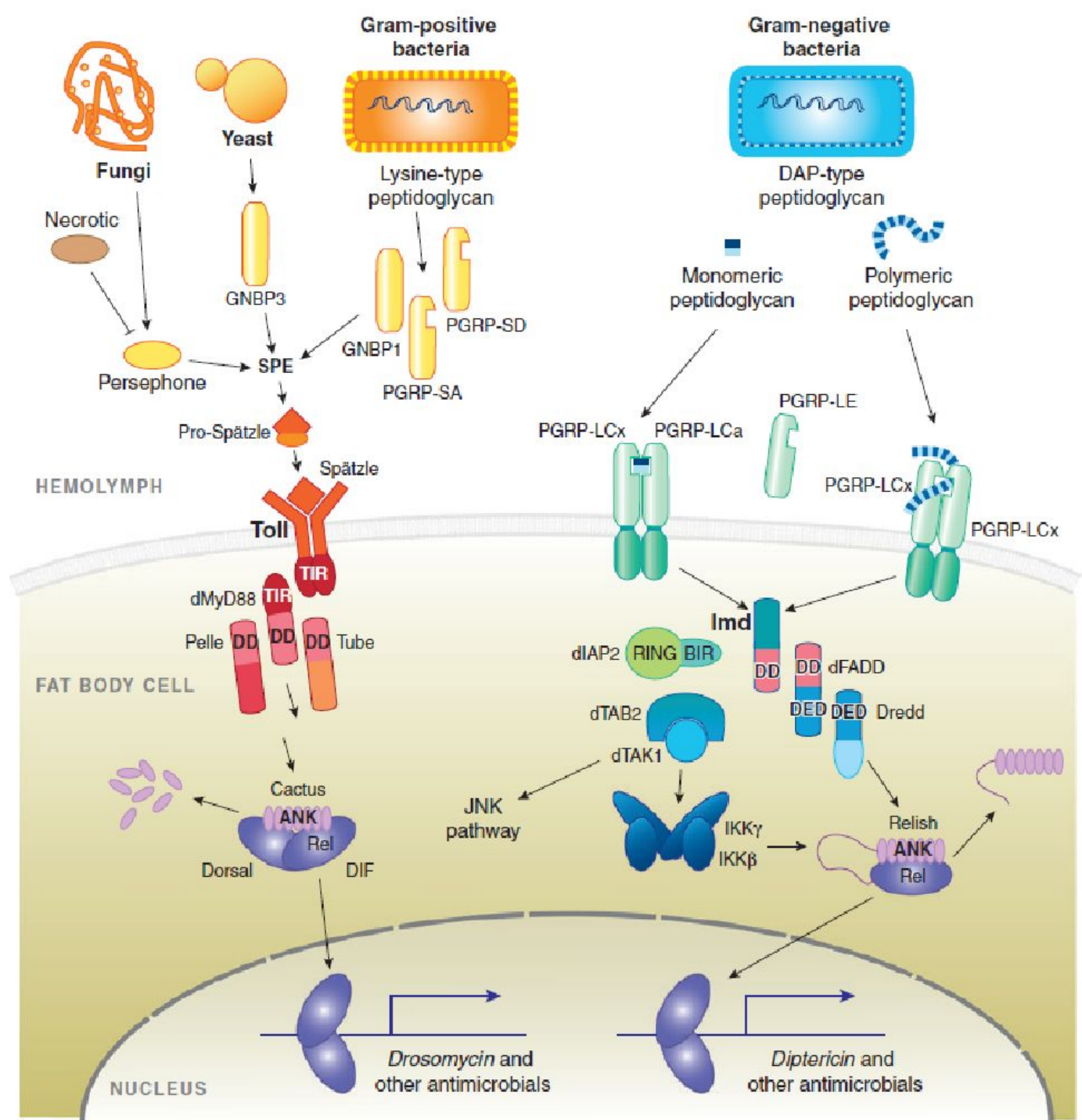
| Peptides | # of genes | Main activity | Concentration | 3-D structure |
|--------------|------------|------------------------|-------------------|---|
| Diptericin | 2 | Gram-negative bacteria | 0.5 μM | nd |
| Attacin | 4 | Gram-negative bacteria | nd | nd |
| Drosocin | 1 | Gram-negative bacteria | 40 μM |  |
| Cecropin | 4 | Gram-negative bacteria | 20 μM |  |
| Defensin | 1 | Gram-positive bacteria | 1 μM |  |
| Drosomycin | 7 | Fungi | 100 μM |  |
| Metchnikowin | 1 | Fungi | 10 μM | nd |

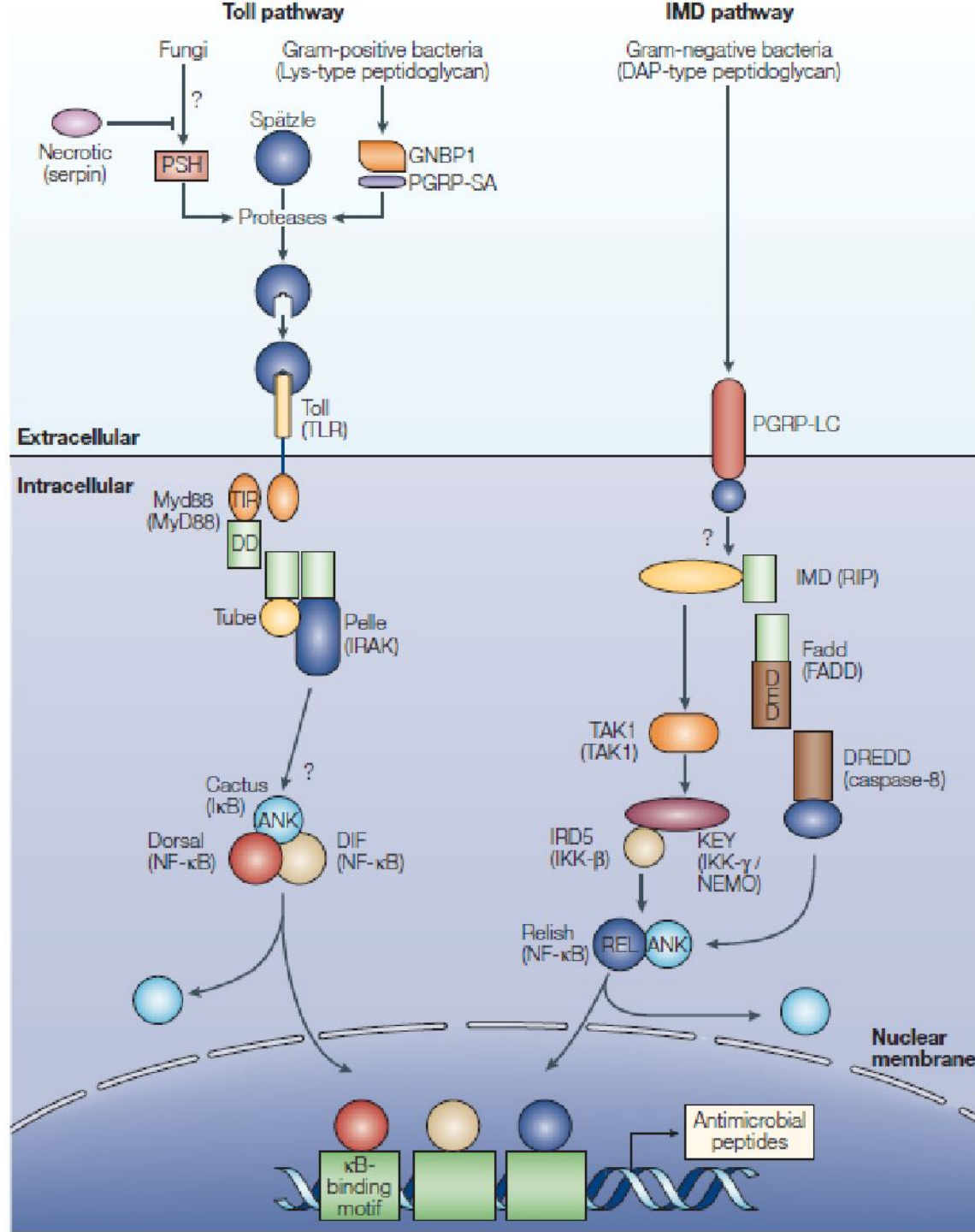
Механизмы встраивания АМП в мембрану патогена



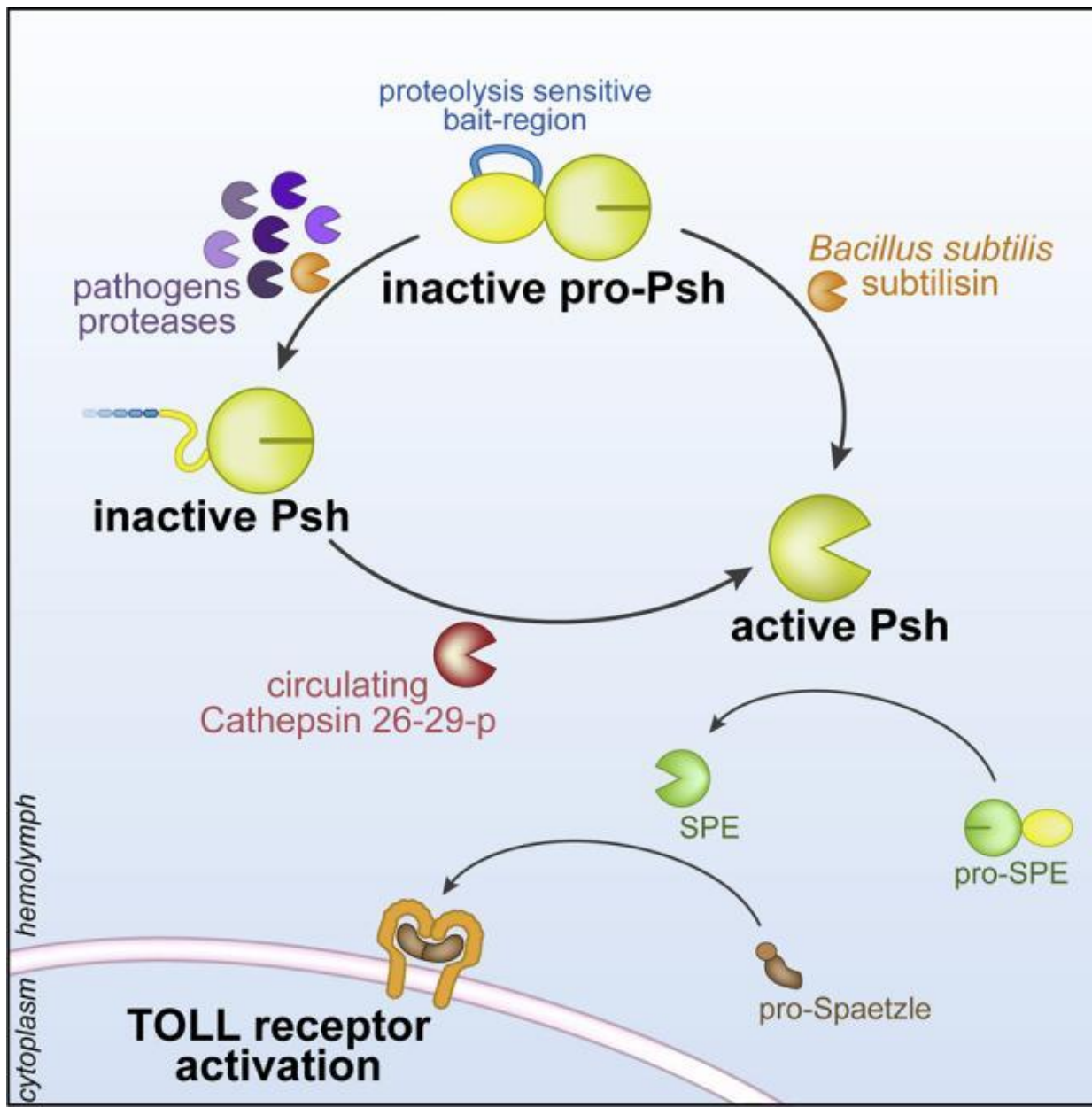
Toll и Imd пути



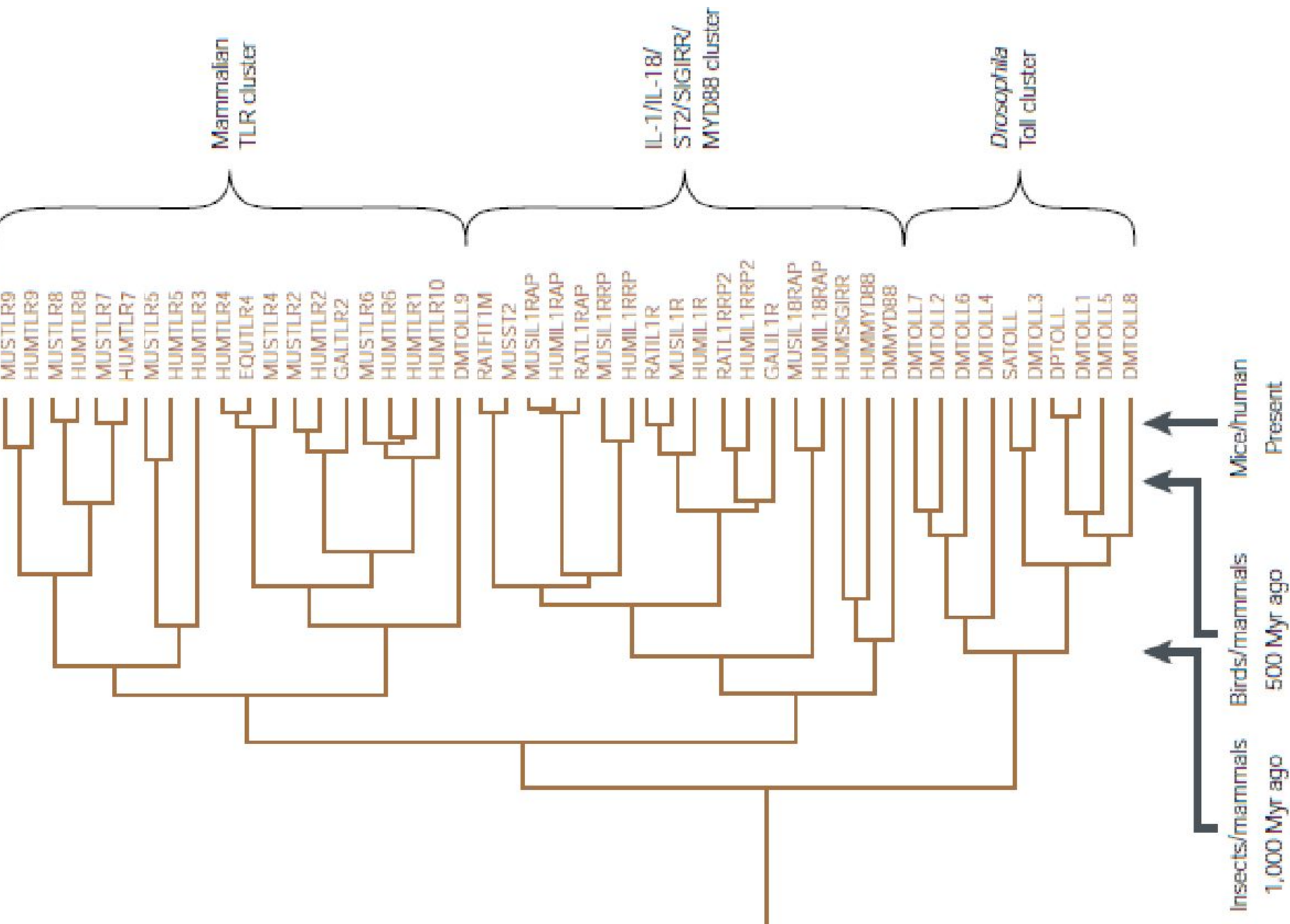


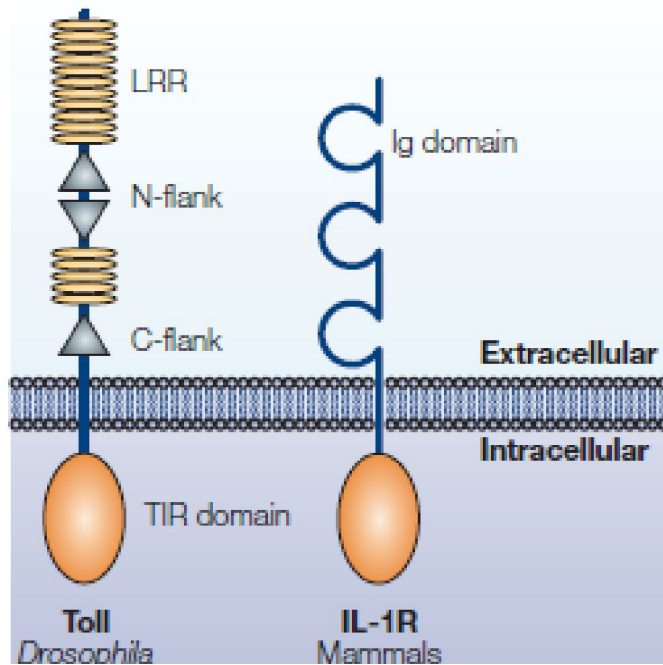


Персефона активируется бактериальными протеазами

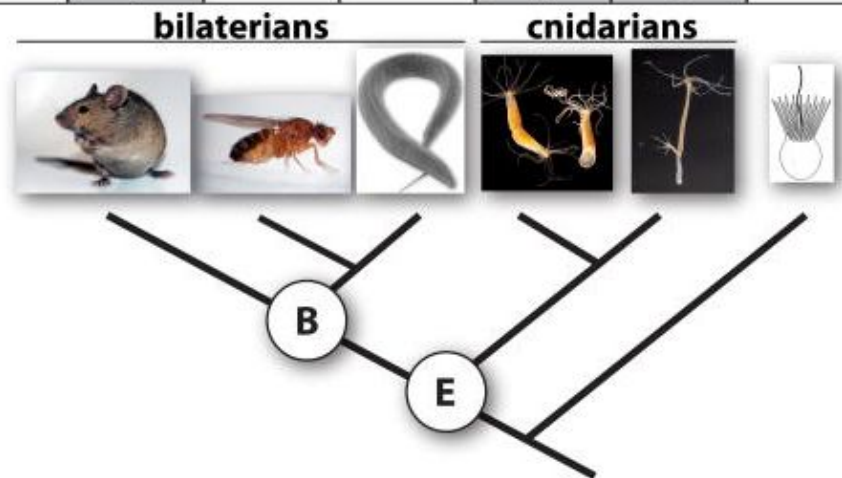


Эволюция Toll-подобных рецепторов

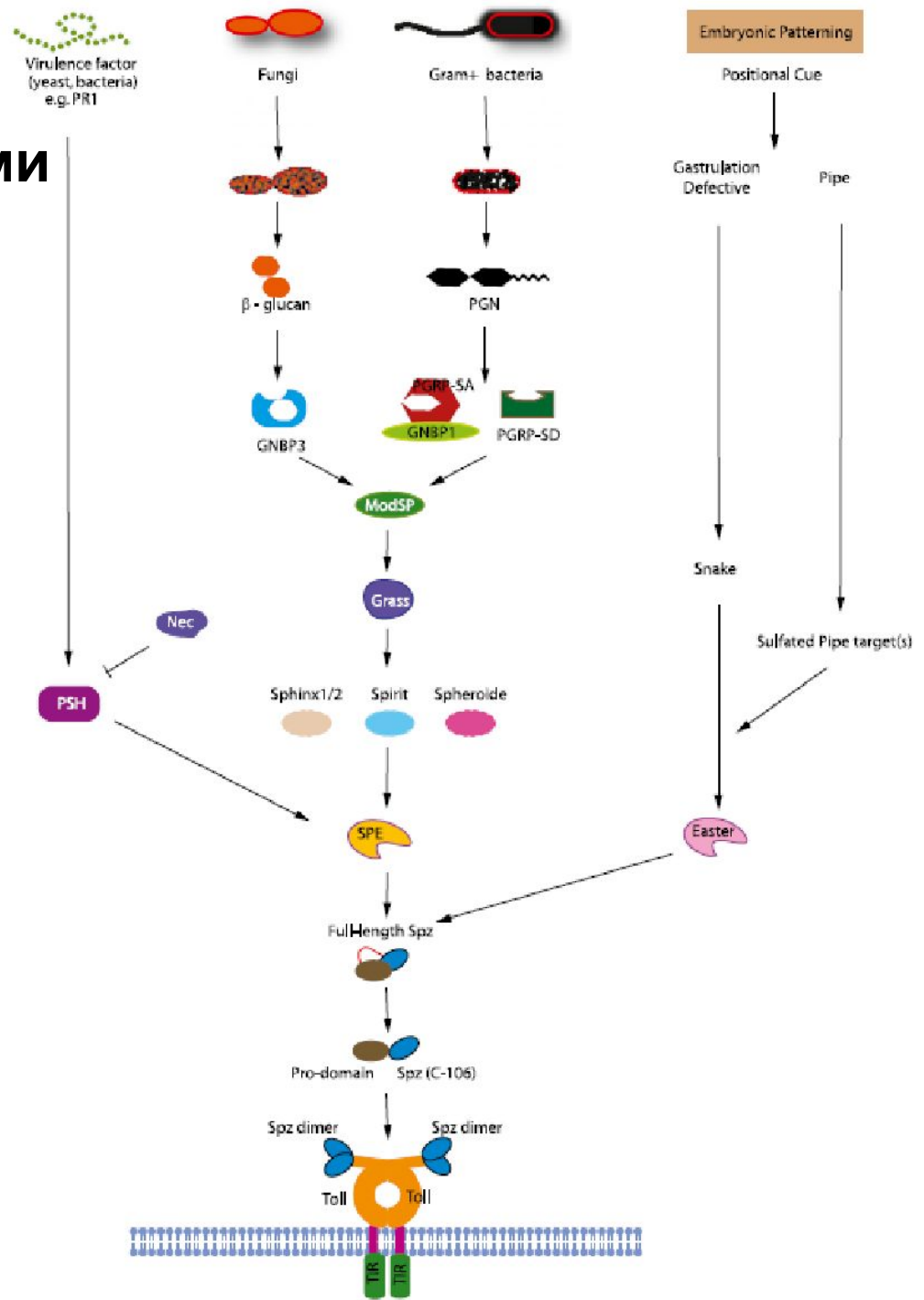




| | <i>Mus musculus</i> | <i>Drosophila melanogaster</i> | <i>Caenorhabditis elegans</i> | <i>Nematostella vectensis</i> | <i>Hydra magnipapillata</i> | <i>Monosiga brevicollis</i> |
|-----------------|---------------------|--------------------------------|-------------------------------|-------------------------------|-----------------------------|-----------------------------|
| TLR | + | + | + | + | ? | |
| SARM | + | + | + | | | |
| MyD88 | + | + | | + | + | |
| IκB | + | + | + | + | + | |
| NF-κB | + | + | | + | | |
| p38 MAPK | + | + | + | + | + | + |
| NLR | + | | | + | ? | |

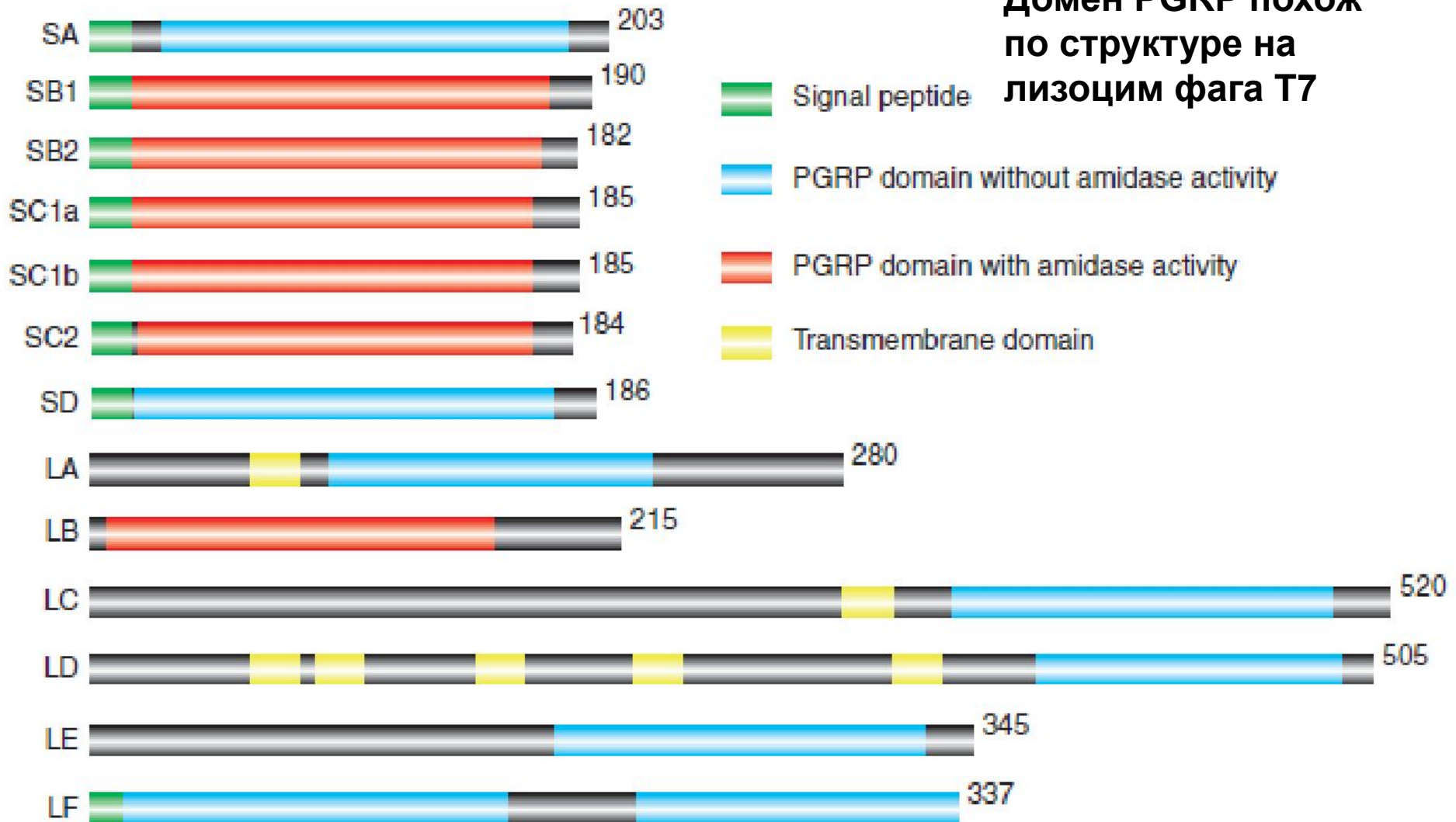


Первичными рецепторами РАМР в Toll-пути у дрозофилы является множество различных белков гемолимфы

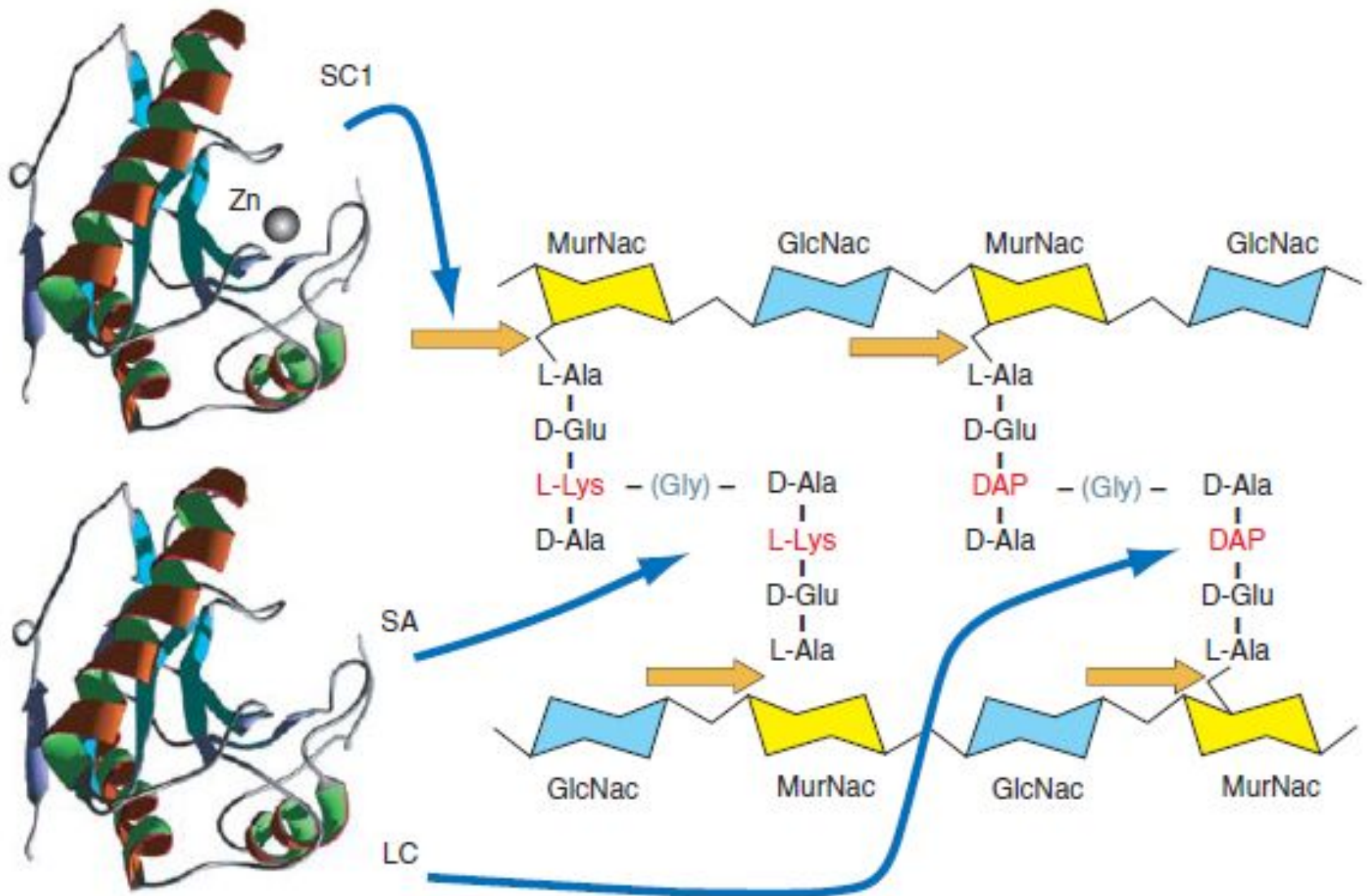


Связывающие пептидогликан белки PGRP дрозофилы

Домен PGRP похож по структуре на лизоцим фага T7



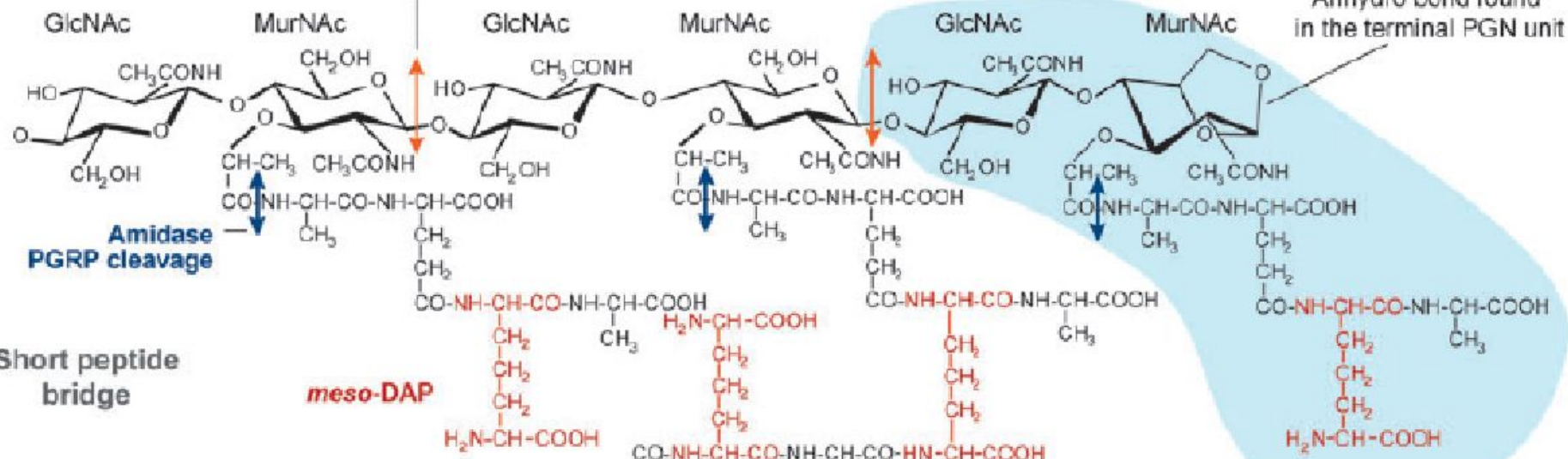
У части PGRP активный центр уже не расщепляет пептидогликан, но специфически распознает его.



Продукты расщепления пептидогликана

Glycan strand

Lysosome cleavage

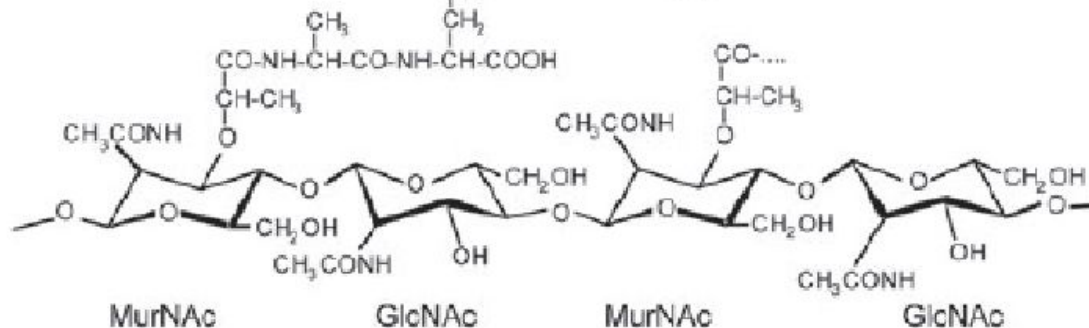


Anhydro bond found in the terminal PGN unit

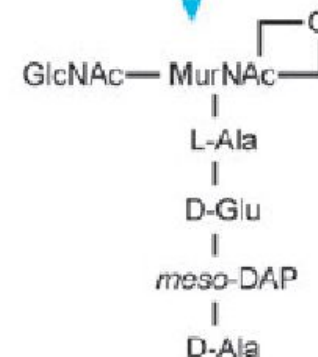
Short peptide bridge

meso-DAP

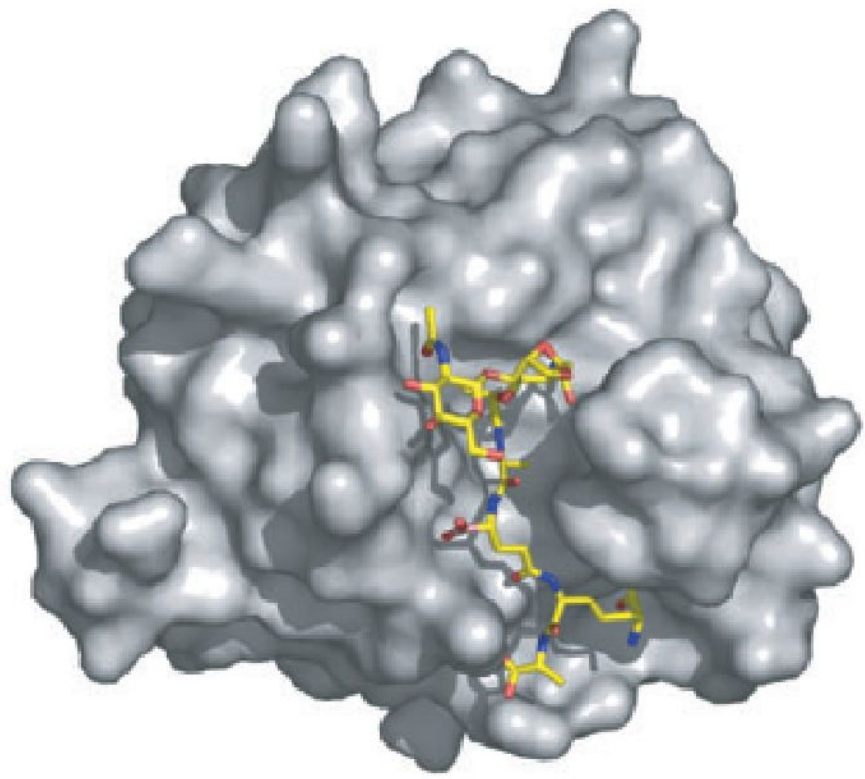
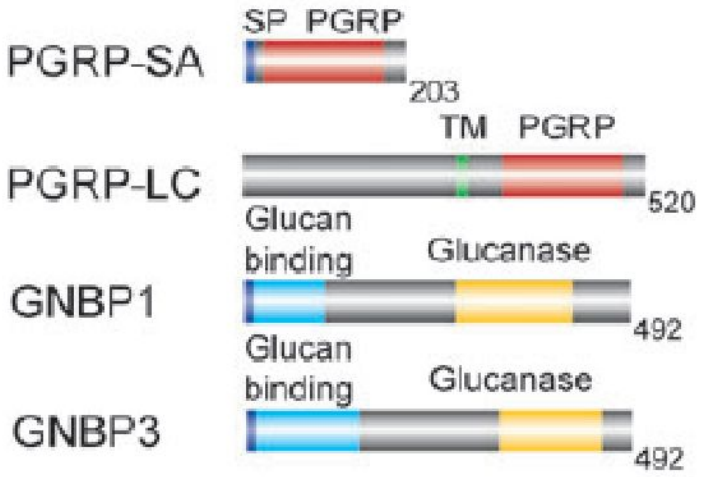
Glycan strand



TCT

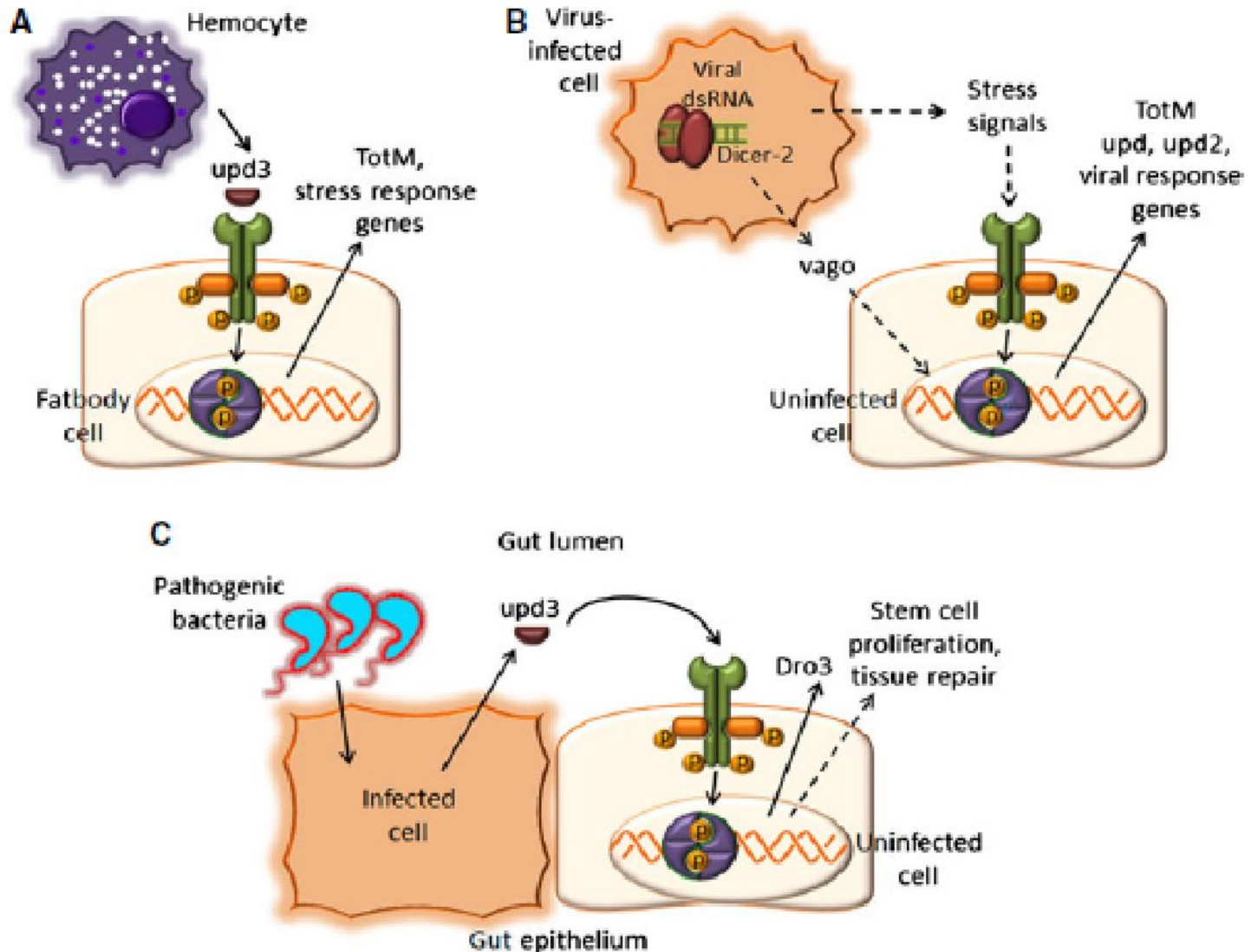


Белки PGRP и NGBP PGRP-LE связавшийся с ТСТ

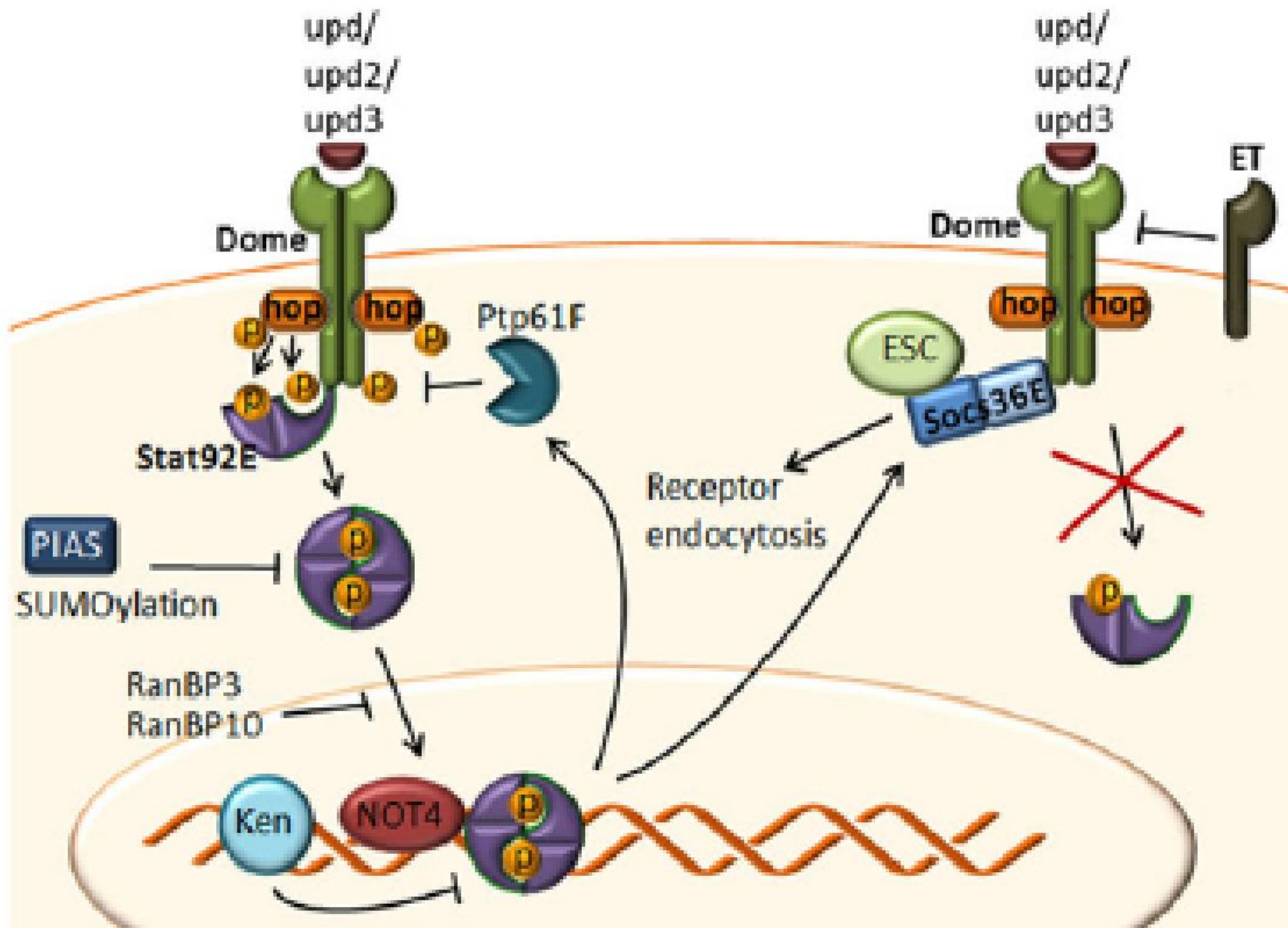


ТСТ – трахеальный цитотоксин, фрагмент пептидогликана с формулой GlcNAc-MurNAc (anhydro)-L-Ala-γ-D-Glu-meso-DAP-D-Ala

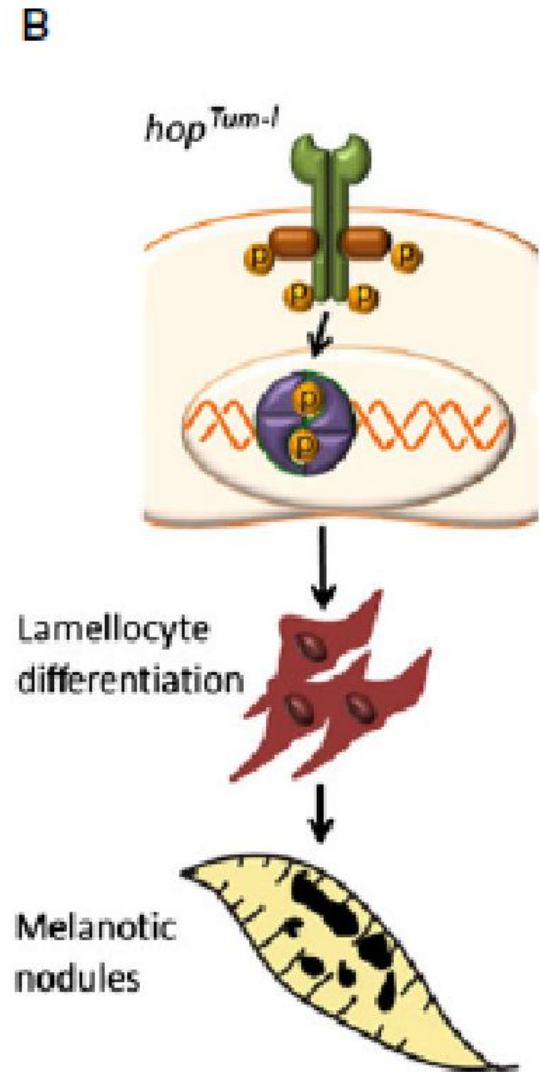
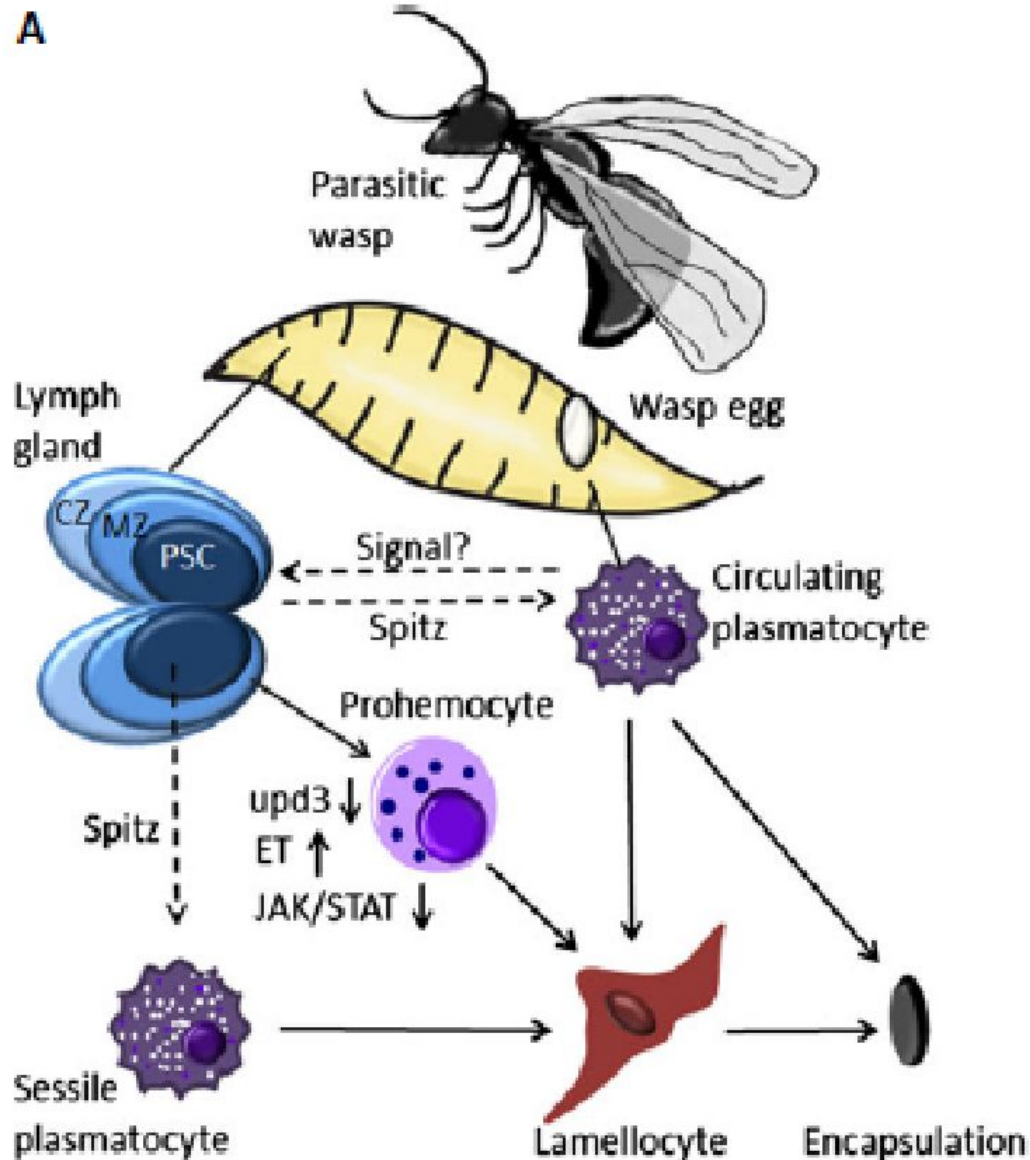
У взрослой мухи JAK/STAT нужен для запуска системного иммунного ответа жировым телом, противовирусного ответа и регуляции регенерации.

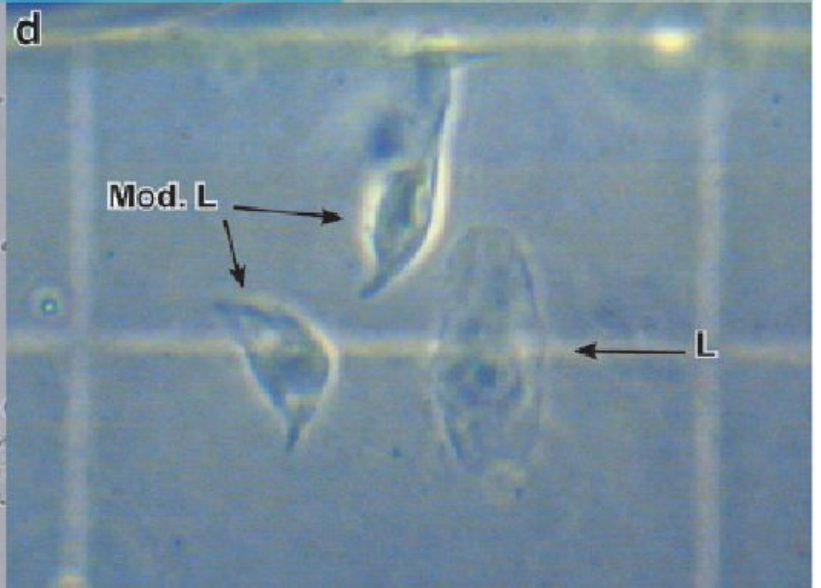
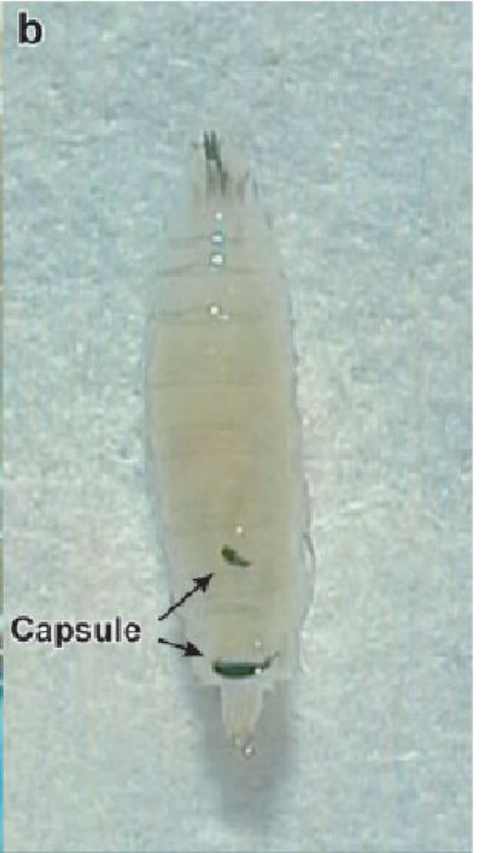


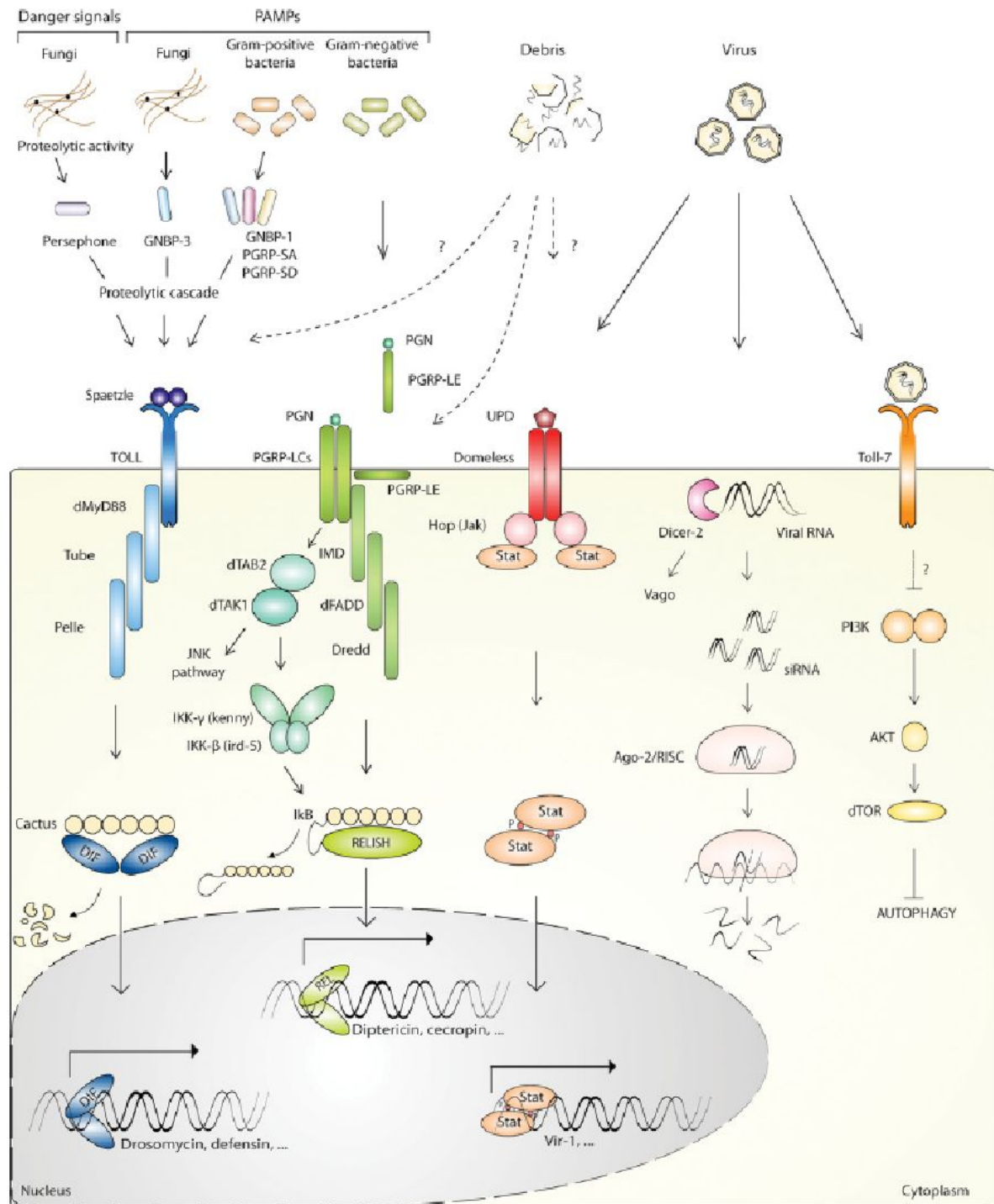
ЈАК/STAT путь



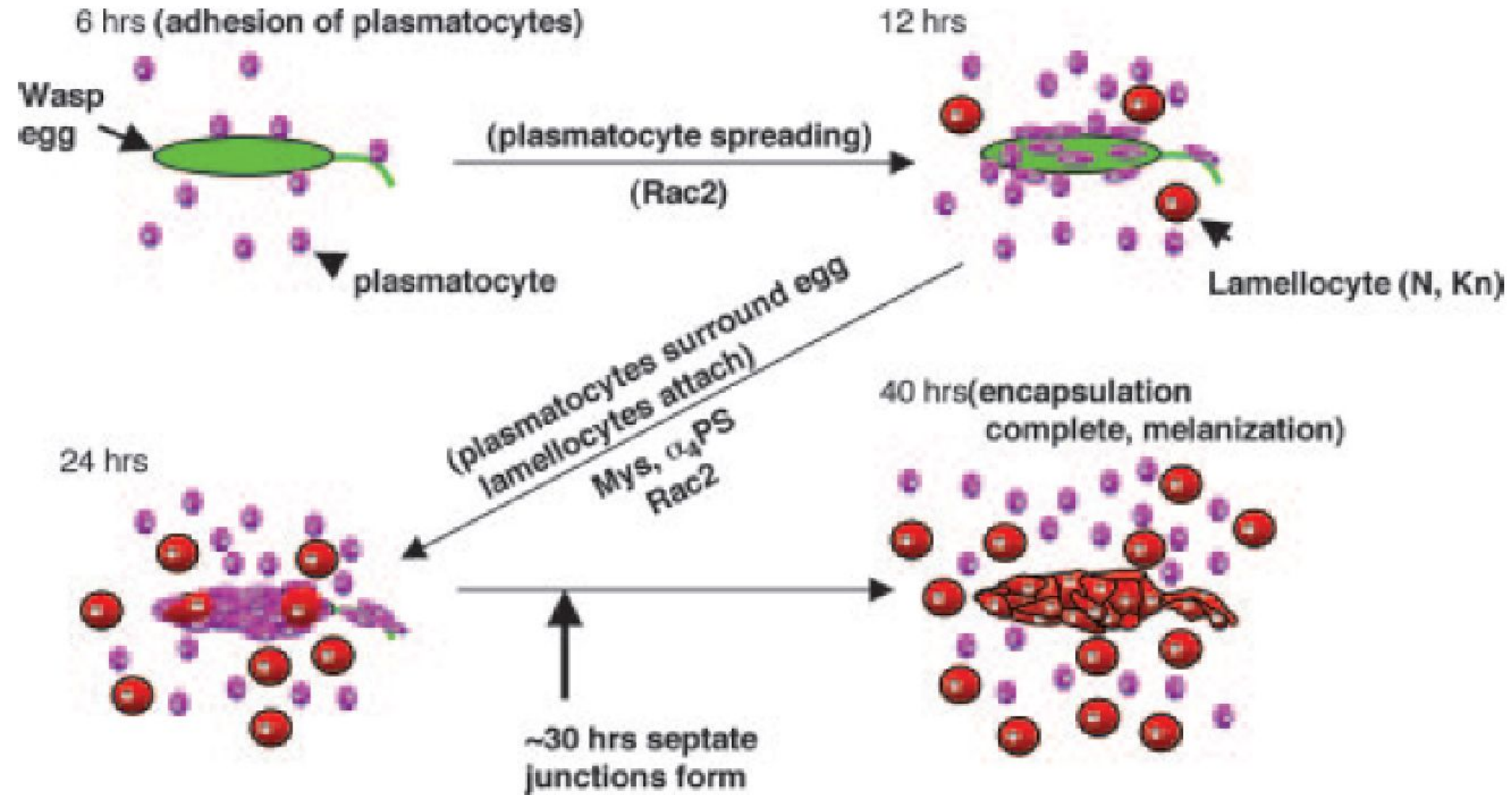
JAK/STAT путь регулирует инкапсуляцию, но может приводить к возникновению опухолей







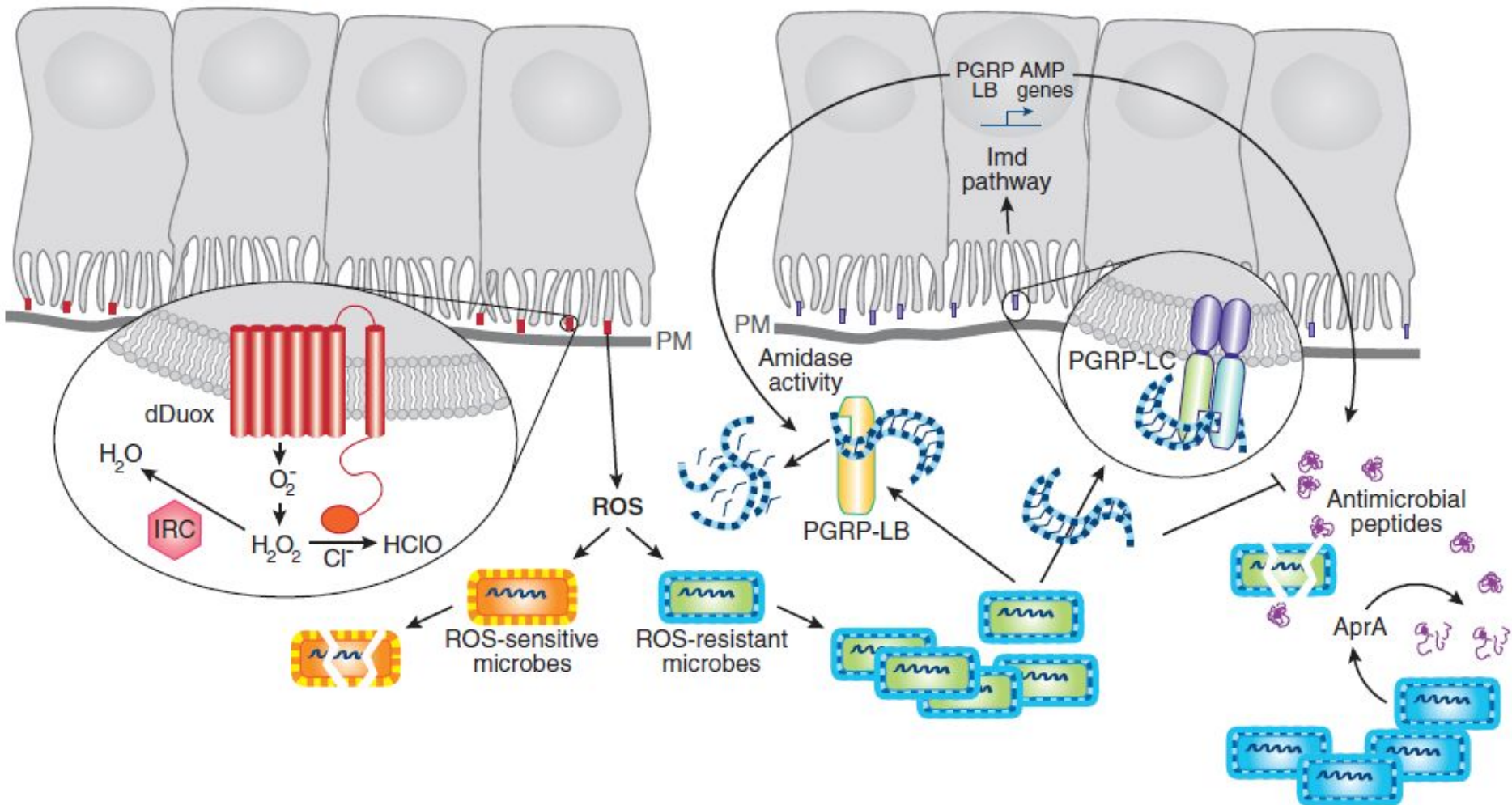
Инкапсуляция яйца паразитоида

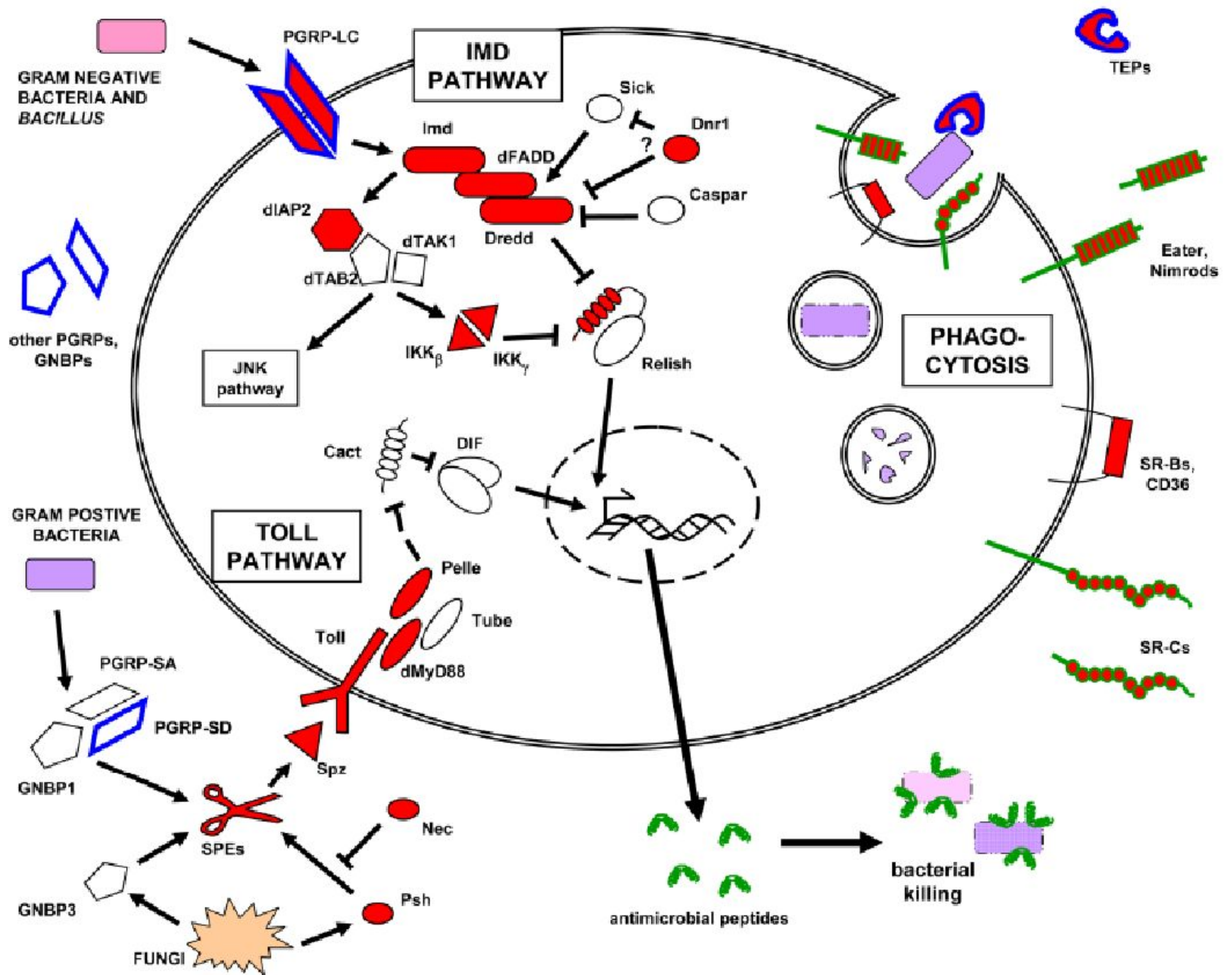


Иммунитет в кишечнике мухи

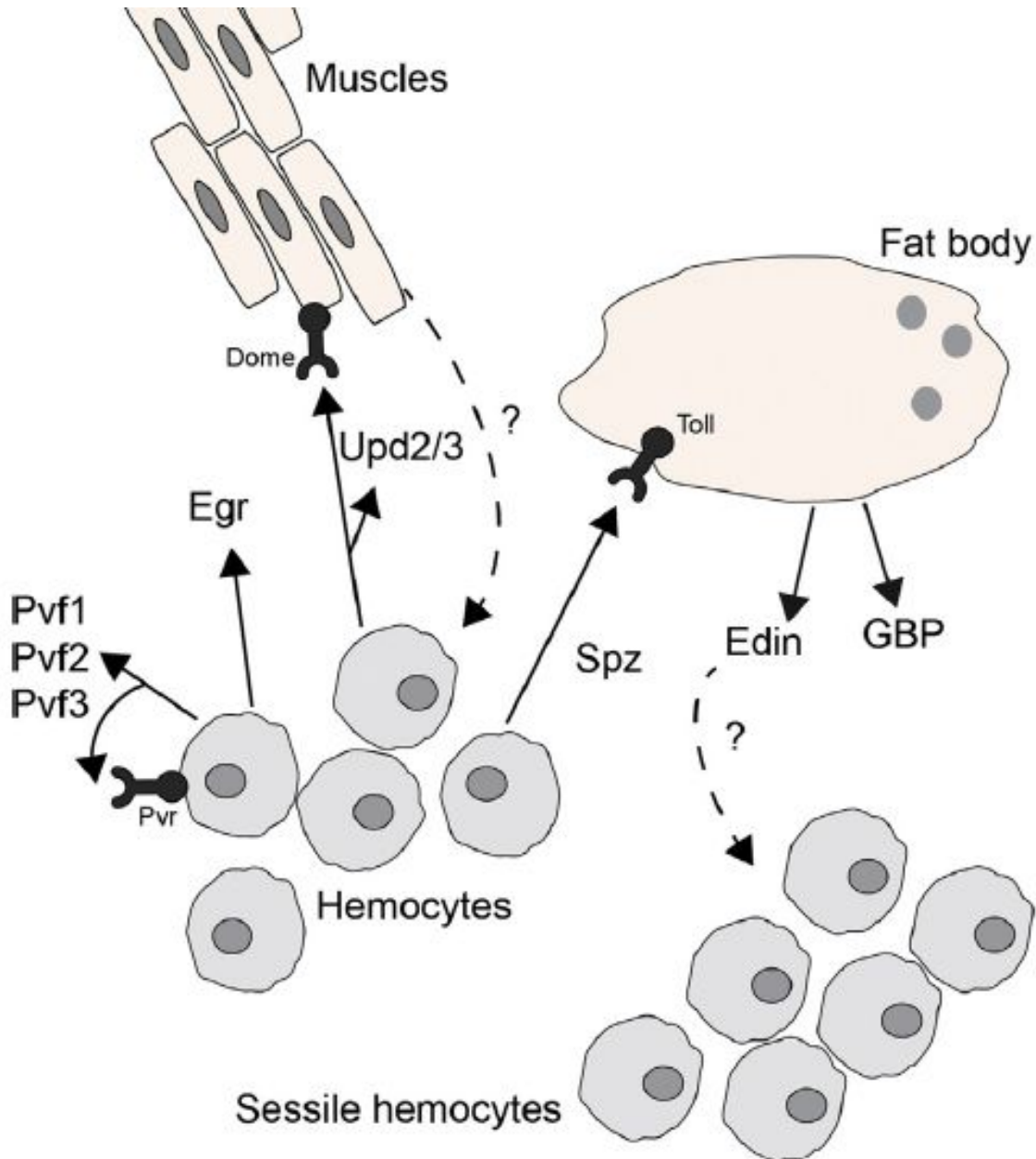
ROS production

AMP production

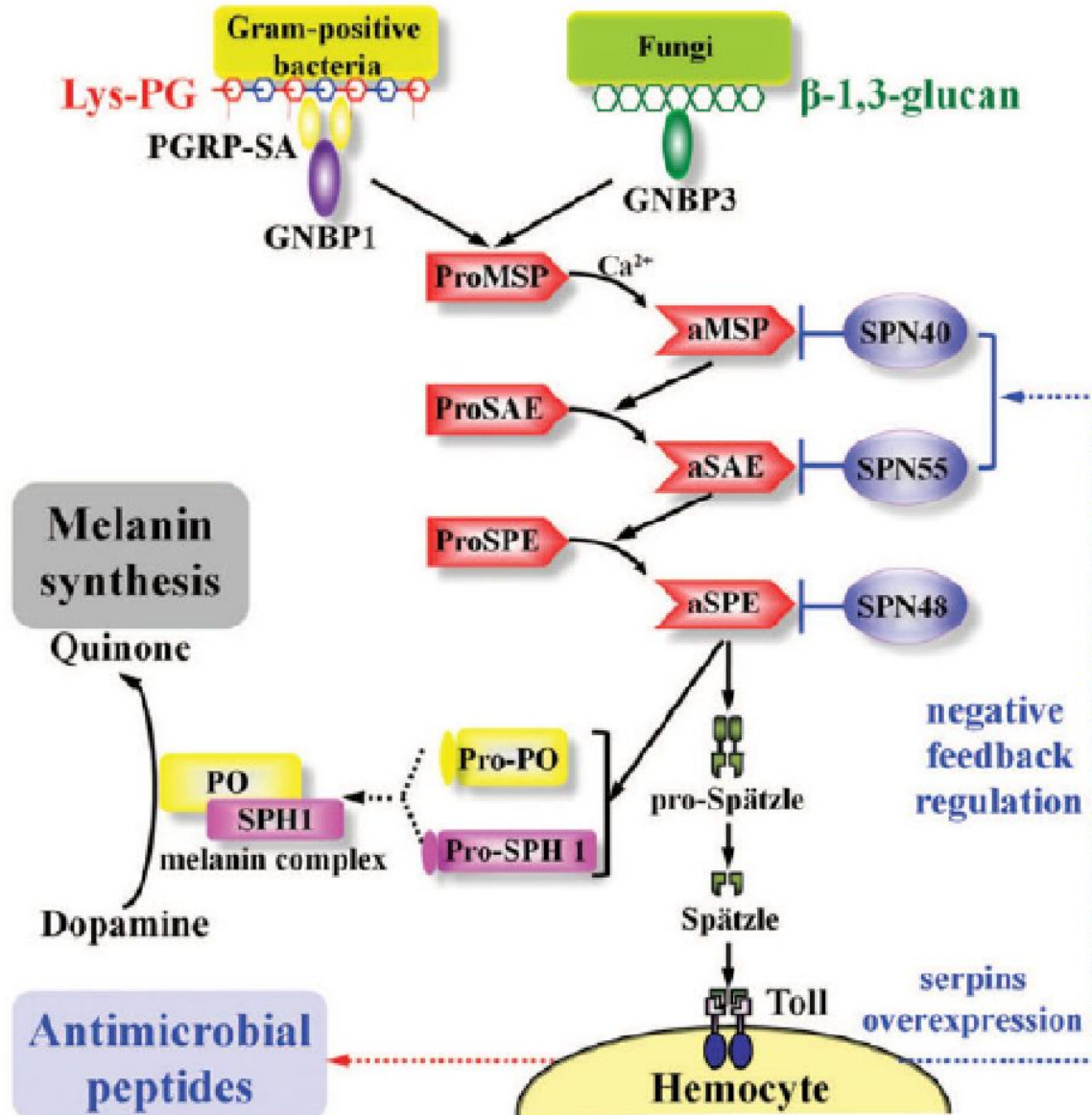




Регуляция иммунных реакций дрозодилы



Обратные связи и кросс-токи иммунного ответа идут при участии серпинов



| Gene | Type of Protein | Putative Ligand | Evidence |
|-----------------------------|--|--|---|
| <i>18 wheeler, Toll-2</i> | Toll-like Receptor | Unknown | Expression of Attacin affected in mutant flies. |
| <i>Toll-9</i> | Toll-like Receptor | Unknown | Protein activates Drosomycin in S2 cells through MyD88. |
| <i>Ird7, totem, PGRP-LC</i> | Peptidoglycan recognition protein | DAP-type peptidoglycans, G- bacteria | Activate IMD pathway in vivo. Phagocytosis of <i>E. coli</i> in S2 cell lines, affected upon RNAi. |
| <i>PGRP-LE</i> | Peptidoglycan recognition protein | DAP-type peptidoglycans, G- bacteria | Overexpression activates PPO cascade in cell lines. Help PGRP-LC recognize peptidoglycans. |
| <i>Semmelweis, PGRP-SA</i> | Peptidoglycan recognition protein | Lys-type peptidoglycans, G+ bacteria | Activation of Toll and phagocytosis of <i>S. aureus</i> pathway affected in mutants. |
| <i>PGRP-SD</i> | Peptidoglycan recognition protein | Lys-type peptidoglycans G+ bacteria | Activation of Toll pathway affected in double mutants with PGRP-SA mutants. |
| <i>Picky, PGRP-SC1a</i> | Peptidoglycan recognition protein | G+ bacterial peptidoglycans | Activation of Toll pathway and <i>S. aureus</i> phagocytosis affected in mutants. |
| <i>PGRP-SC1b</i> | Peptidoglycan recognition protein | G+ bacterial peptidoglycans | Cleaves <i>S. aureus</i> peptidoglycans. |
| <i>Osiris, GNBPI</i> | Gram-negative binding protein | Potentially G+ bacterial determinants | Hydrolyzes G+ peptidoglycan. Acts in complex with PGRP-SA to activate Toll pathway. |
| <i>TEPs</i> | Thiolester containing proteins | Possibly binding to bacterial surface | RNAi of homologous mosquito gene reduces phagocytosis of Gram-negative bacteria. Plasmodia population larger in <i>Tep1</i> mutant mosquito, causing higher vectorial capacity. RNAi of <i>Tep2</i> decreases uptake of <i>E. coli</i> ; RNAi of <i>Tep3</i> decreases uptake of <i>S. aureus</i> . |
| <i>Ma</i> | Macroglobulin complement-related | <i>Candida albicans</i> | RNAi of S2 cells reduces phagocytosis of <i>C. albicans</i> |
| <i>dSR-C1</i> | Scavenger receptor | Possibly both G+ and G- bacteria | RNAi of S2 cells reduces phagocytosis of bacteria. |
| <i>Crq (Croquemort)</i> | Scavenger receptor, CD36 like | Apoptotic cells and possibly G+ bacteria | Phagocytosis of <i>S. aureus</i> impaired in cell lines. |
| <i>Peste</i> | Scavenger receptor, CD36 like | Mycobacteria | RNAi of S2 cells reduces phagocytosis of Mycobacteria. |
| <i>Eater</i> | Scavenger receptor, epidermal growth factor like | Possibly G+ and G- bacteria | Reduction in phagocytosis of <i>S. aureus</i> and <i>E. coli</i> in vitro in cellline mutants and in vivo in deficiency. Increased susceptibility to natural infection in vivo. |

- Следующая лекция 12 марта – снова об иммунной системе дрозофилы