

# Plasmids and Plasmid Biology

By

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# Plasmids and Plasmid Biology



- Plasmid structure
- Plasmid replication and copy number control
- Plasmid transfer
- Plasmids as tools
- F plasmids

# Plasmids

- Extrachromosomal DNA, usually circular-parasite?
- Usually encode ancillary functions for in vitro growth
- Can be essential for specific environments: virulence, antibiotics resistance, use of unusual nutrients, production of bacteriocins (colicins)
- Must be a replicon - self-replicating

# Plasmids

- Plasmid DNA must replicate every time host cell divides or it will be lost
  - a. DNA replication
  - b. partitioning (making sure each progeny cells receives a plasmid)
- High copy plasmids are usually small; low copy plasmids can be large
- Partitioning is strictly controlled

# Plasmids

- Plasmid replication requires host cell functions
- Copy number is regulated by initiation of plasmid replication
- Plasmids are incompatible when they cannot be stably maintained in the same cell because they interfere with each other's replication.

# "Old School method of purifying plasmid"

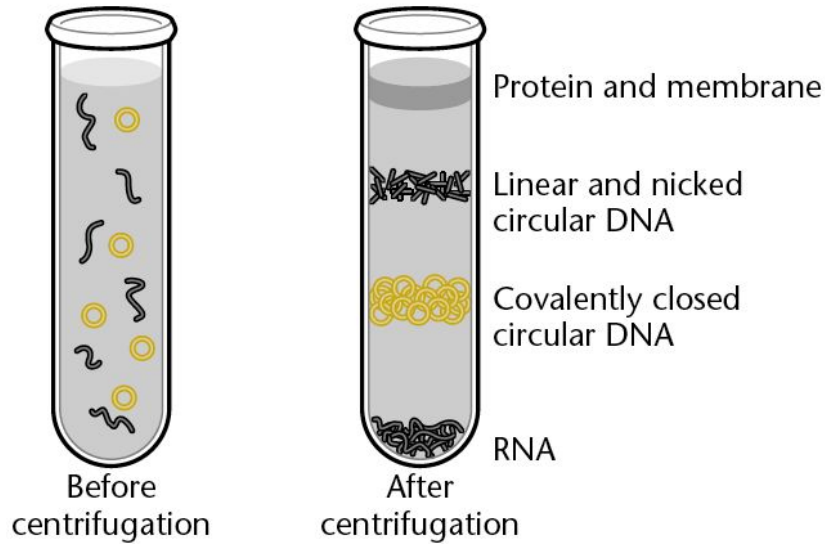
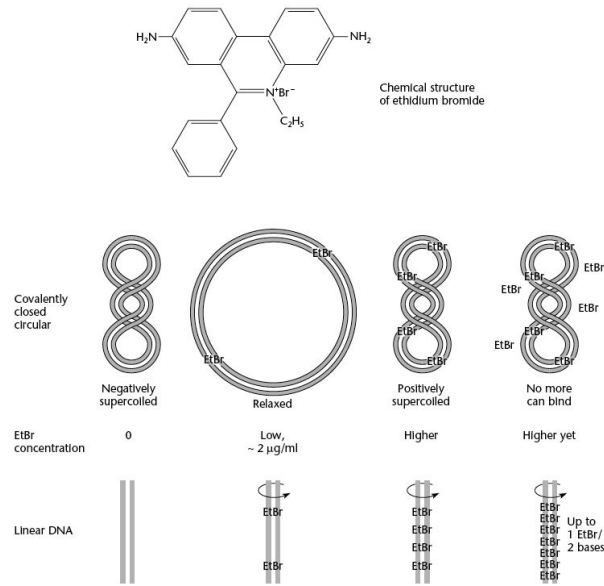


Figure 4.2



Three forms of plasmid DNA



CsCl gradient with ethidium bromide and UV light

TABLE 4.1 Some naturally occurring plasmids and the traits they carry		
Plasmid	Trait	Original source
ColE1	Bacteriocin which kills <i>E. coli</i>	<i>E. coli</i>
Tol	Degradation of toluene and benzoic acid	<i>Pseudomonas putida</i>
Ti	Tumor initiation in plants	<i>Agrobacterium tumefaciens</i>
pJP4	2,4-D (dichlorophenoxyacetic acid) degradation	<i>Alcaligenes eutrophus</i>
pSym	Nodulation on roots of legume plants	<i>Rhizobium meliloti</i>
SCP1	Antibiotic methylenomycin biosynthesis	<i>Streptomyces coelicolor</i>
RK2	Resistance to ampicillin, tetracycline, and kanamycin	<i>Klebsiella aerogenes</i>

Virulence plasmids from *Salmonella*, *Shigella*, *Yersinia*, *B. anthracis*, *E.coli*, and others.

TABLE 4.2 Copy numbers of some plasmids	
Plasmid	Approximate copy number
F	1
P1 prophage	1
RK2	4–7 (in <i>E. coli</i> )
pBR322	16
pUC18	~30–50
pIJ101	40–300

**Table 11-1** Examples of some plasmids and their properties

Plasmid	Size (Kb)	Number of copies per chromosome	Self-transmissible	Phenotypic features
<i>Col plasmids</i>				
ColE1	6.4	10–15	No	Colicin E1 disrupts energy gradient, host immunity to Colicin E1
ColE2	7.6	10–15	No	Colicin E2 is a DNase, host immunity to Colicin E2
ColE3	7.6	10–15	No	Colicin E3 is a ribosomal RNase, host immunity to Colicin E3
<i>F plasmid</i>	94.5	1–2	Yes	F-pilus, conjugation
<i>R plasmids</i>				
R100	106.7	1–2	Yes	Cam <sup>r</sup> Str <sup>r</sup> Sul <sup>r</sup> Tet <sup>r</sup>
RK2	56.0	5–8	Yes	Broad host range
pSC101	9.0	<5	No	Low copy number, compatible with ColE1-type plasmids, Tet <sup>r</sup>
<i>Phage plasmid</i>				
$\lambda$ dv	6.4	50	No	$\lambda$ genes <i>cro</i> , <i>ci</i> , <i>O</i> , <i>P</i>
<i>Recombinant plasmids</i>				
pBR322	4.4	20	No	Medium copy number, ColE1-type replication, Amp <sup>r</sup>
pUC18	2.7	200–500	No	High copy number, ColE1-type replication with a mutation that increases the copy number, Amp <sup>r</sup>
pACYC184	4.0	10–12	No	Cam <sup>r</sup> Tet <sup>r</sup>



# Plasmid replication

- Plasmid replication requires host DNA replication machinery.
- Most wild plasmids carry genes needed for transfer and copy number control.
- All self replication plasmids have a *oriV*: origin of replication
- Some plasmids carry and *oriT*: origin of transfer. These plasmids will also carry functions needed to be mobilized or *mob* genes.

# Plasmid replication

- Plasmid segregation is maintained by a *par* locus—a partition locus that ensures each daughter cells gets on plasmid. Not all plasmids have such sequences.
- There are 5 main “incompatibility” groups of plasmid replication. Not all plasmids can live with each other.
- Agents that disrupt DNA replication destabilize or cure plasmids from cells

**A**



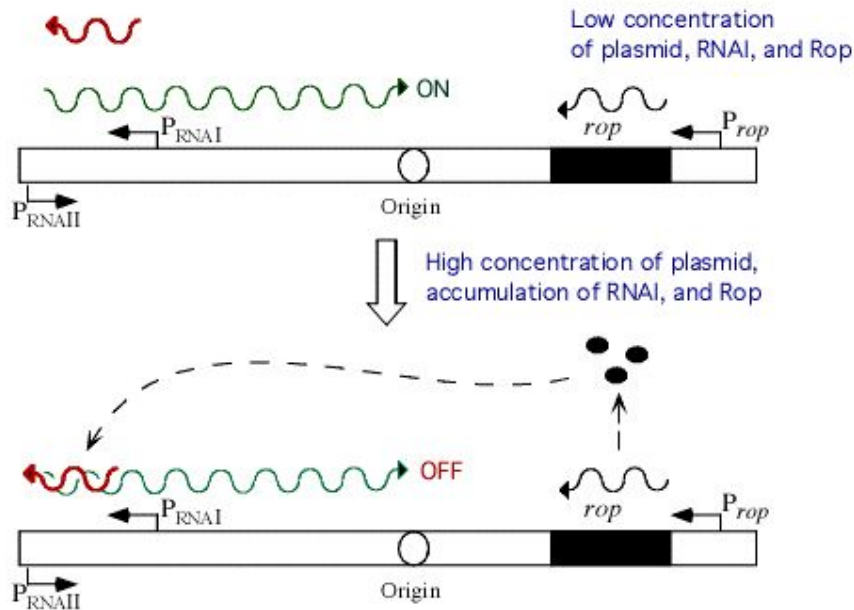
**B**



## Antisense RNA gene control.

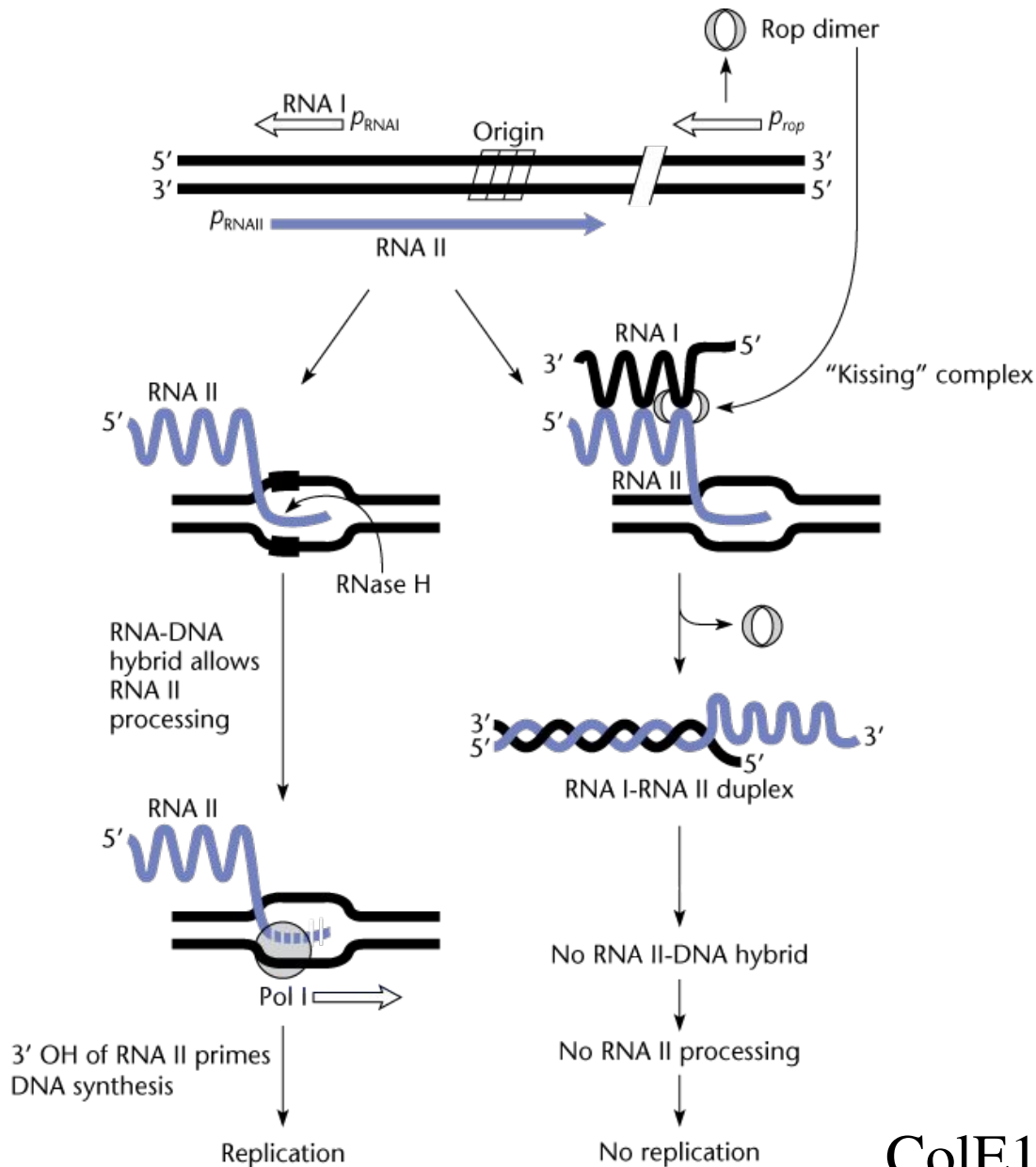
- the RNA-RNA hybrid is very stable
- blocks most translation and transcription
- requires RNAases to degrade
- common theme in bacterial gene regulation as we are learning

# Anti-sense RNA replication control



RNA I-small inhibitory RNA that binds to RNAII. RNAII will act as a primer for DNA replication  
 Rop: plasmid encoded proteins which stabilizes the RNAI-RNAII complex

Antisense RNA: RNA-RNA hybrid blocks replication  
 GGCUAAUUCC      Antisense RNA is also used in euks  
 called  
 CCGAUUAAGG      siRNA  
 Blocking RNA priming for DNA PolI prevents  
 replication



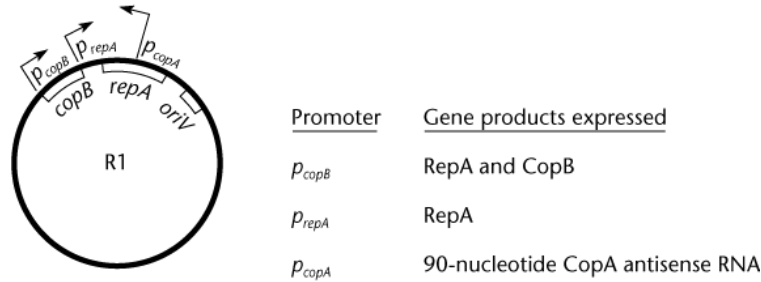
## ColE1 Replication

Control-an example of primer control of replication

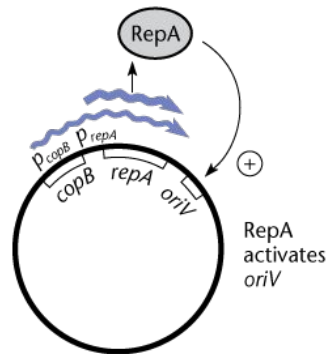
- RNAII will serve as a primer for the replication fork.
- The 3' end is processed by host RnaseH to allow efficient RNA-DNA hybrid to form
- The hybrid acts as a primer for host Pol1
- As the concentration of plasmid increases, Rop does also
- Rop stabilizes the RNA1-II complex
- No RNA for replication priming.

ColE1 replication does not need plasmid encoded rep proteins

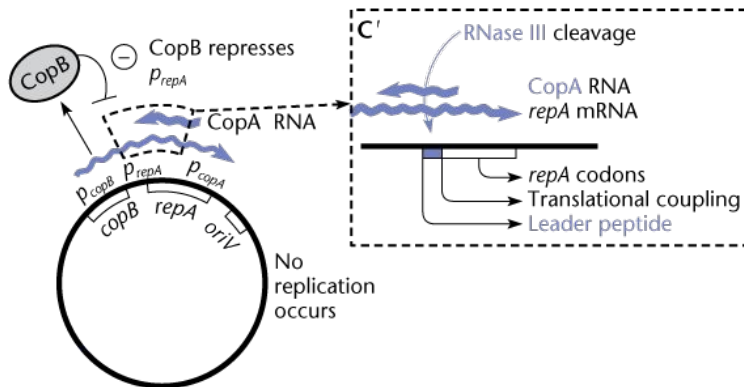
## A Plasmid genetic organization



## B Replication occurs after plasmid enters cells



## C Replication shutdown

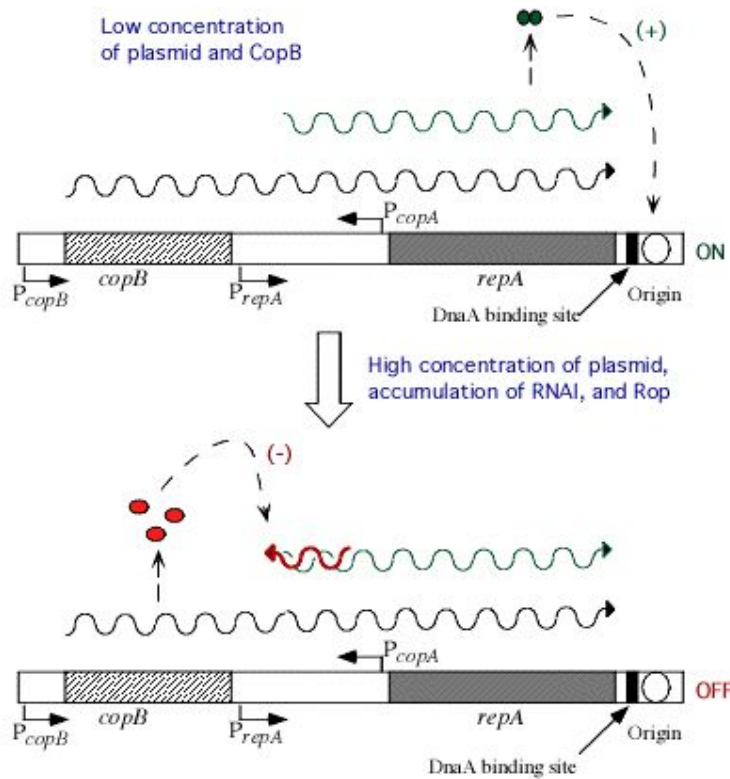


The events upon entry into a cell

- RepA mRNA is made from  $P_{repA}$  until copy number becomes high
- CopB expression increases and CopB represses RepA expression at  $P_{repA}$
- CopA now is made—a 90-base antisense RNA
- CopA binds to 5'-end of the RepA mRNA, forming dsRNA
- This is recognized by host RNAaseIII and degraded.

Thus concentration of RepA protein is maintained by rate of RNA-RNA hybrid formation.

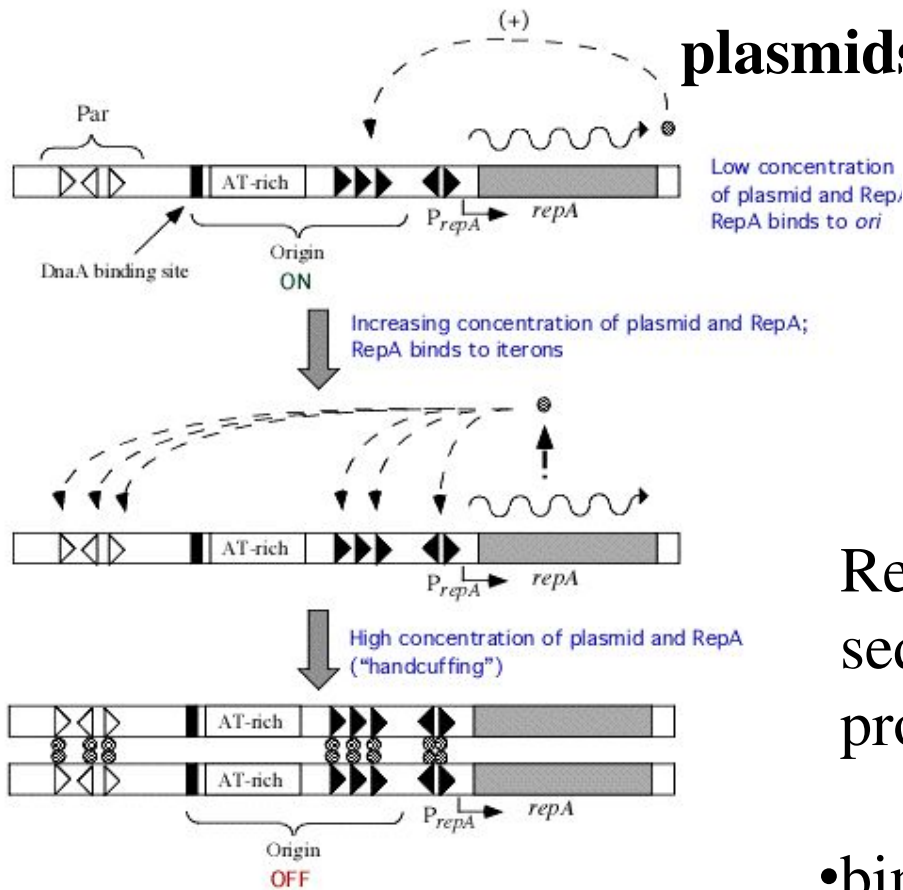
# Rep-protein control -R1 family of plasmids.



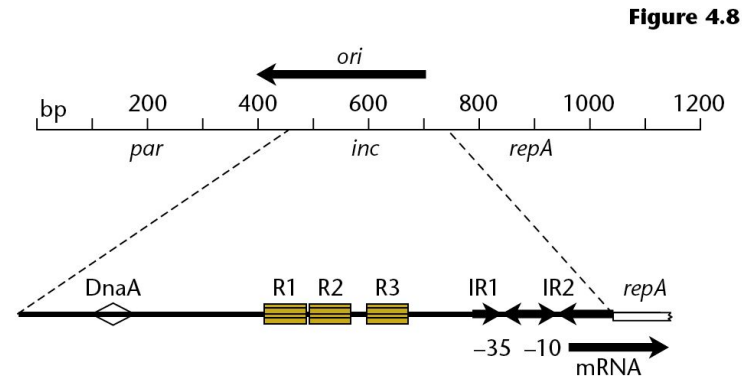
**Plasmid copy control  
balanced by host  
RNaseIII activity and  
transcription from the  
plasmid**

- Rep-protein expression controlled by antisense CopA
- $P_{copB}$ -encodes Rep and CopB
- $P_{copA}$ -encodes antisense RNA
- plasmid replicates to high level
- CopB levels rise, shutting off RepA production
- antisense RNA from  $P_{copA}$  made

# Iteron Plasmids: Handcuffing RK2 and other broad host range plasmids



**“copy up” mutants:**  
mutations in RepA that  
are less able to bind to  
each other.



RepA is able to bind the repeat sequences upstream of the promoter region for *repA*.

- binding causes two plasmid molecules to couple “handcuff”
- prevents replication.

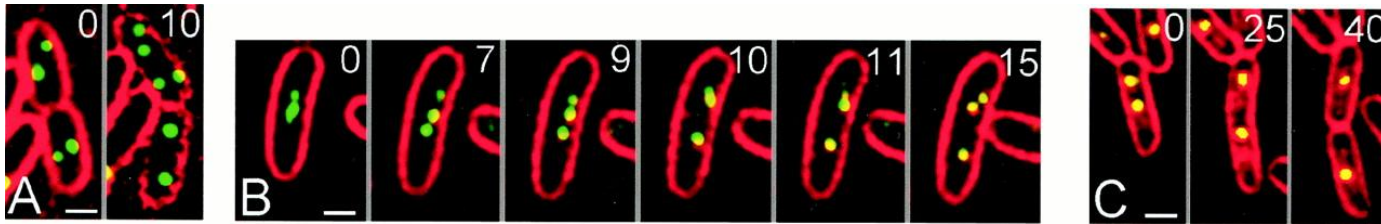


## Incompatibility Groups

- Not all plasmids can live together.
- Plasmids that are able to coexist in the same cell do not interfere with each other's replication
- A single cell can have as many Inc group plasmids as it can tolerate and replicate!

**Partion Locus:** a region on broad host range plasmids that binds to a structure on the inner membrane of the cell to ensure proper segregation.

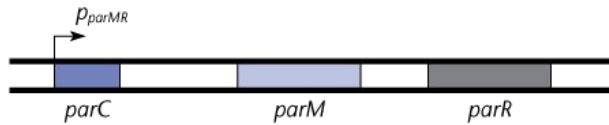
Plasmids labeled with fluorescent protein  
-move to each daughter cell during division.



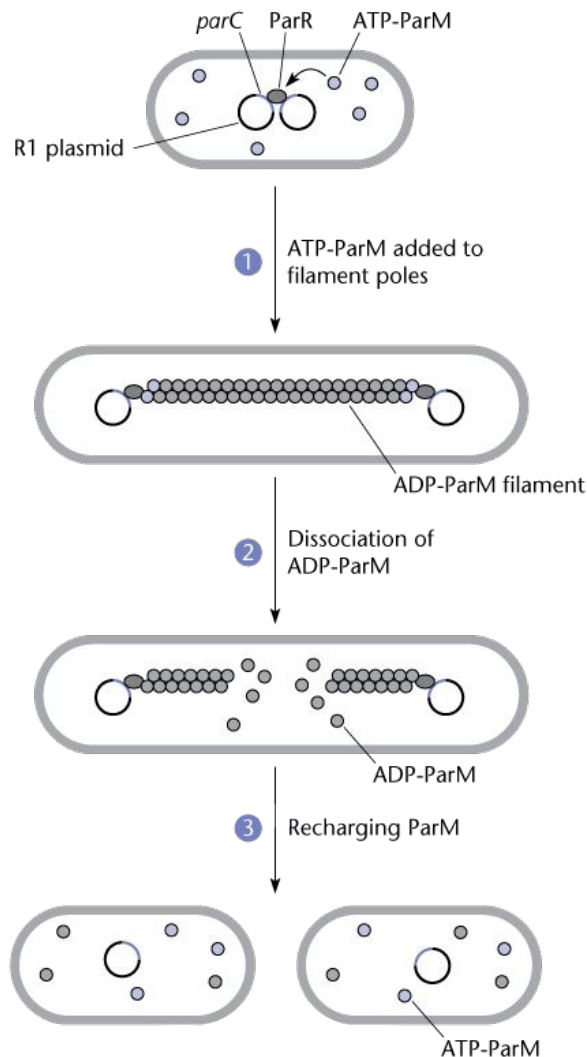
Pogliano, Joe et al. (2001) Proc. Natl. Acad. Sci. USA 98, 4486-4491

Figure 4.18

A *parCMR* locus



B Plasmid R1 partitioning



## Par locus

- think of this as a primitive centromere
- the growing filaments push the plasmids to the opposite poles of the cells

# Plasmids as genetic tools: Construction of Mutants

## Site-directed mutation: Suicide

### plasmids

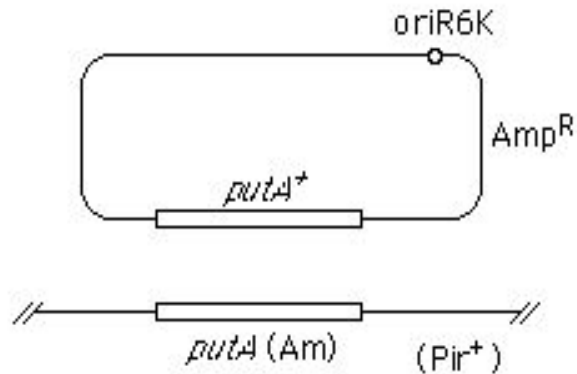
1. Plasmids must be unable to replicate without essential replication proteins provide *in trans*.
2. It helps if the plasmid can be mobilized-*oriT* required
3. Need a selectable marker
4. Large or small region of homologous DNA cloned that will integrate into the chromosomal target.
5. Need a counter selection method to kill the donor cells

### Also merodiploid reporter strains can be constructed in this manner

1. Make a *lacZ* fusion to your promoter of interest
2. Clone into a suicide plasmid
3. Mate into recipient.
4. Resulting strain will harbor a duplication of the promoter region:*lacZ* and still have a functional copy of the gene.

Why would this be important?

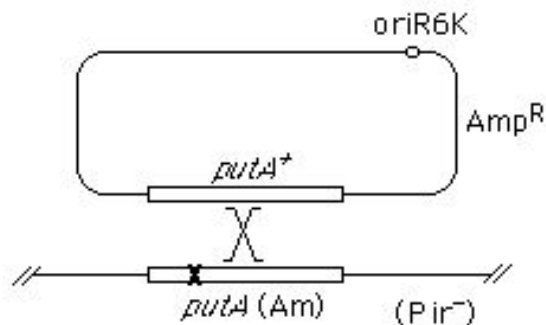
(a)



The plasmid will replicate in a  $Pir^+$  host. Hence, the cell will be  $Put^+$  due to complementation from the  $putA^+$  gene encoded by the multicopy plasmid.

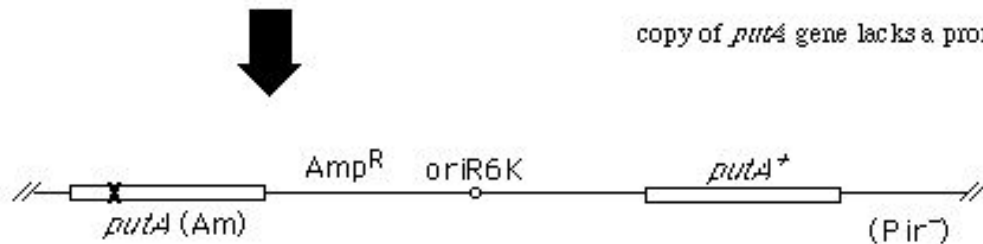
R6K: broad host plasmid.  
-Pir is the essential replication protein  
-*pir* mutants cannot replicate unless supplied *in trans*.

(b)



The plasmid cannot replicate in a  $Pir^-$  host. However, it is possible to select for integration into the chromosome by demanding  $Amp^R$ . Once integrated, the cell will be  $Put^+$  due to complementation from the copy of  $putA^+$  encoded by the integrated plasmid. (This is only true if the plasmid copy has its own promoter. What would happen if the plasmid copy of  $putA$  gene lacks a promoter?)

-integration into the chromosome is selected for by growth on ampicillin



How could you make targeted mutant using this method?

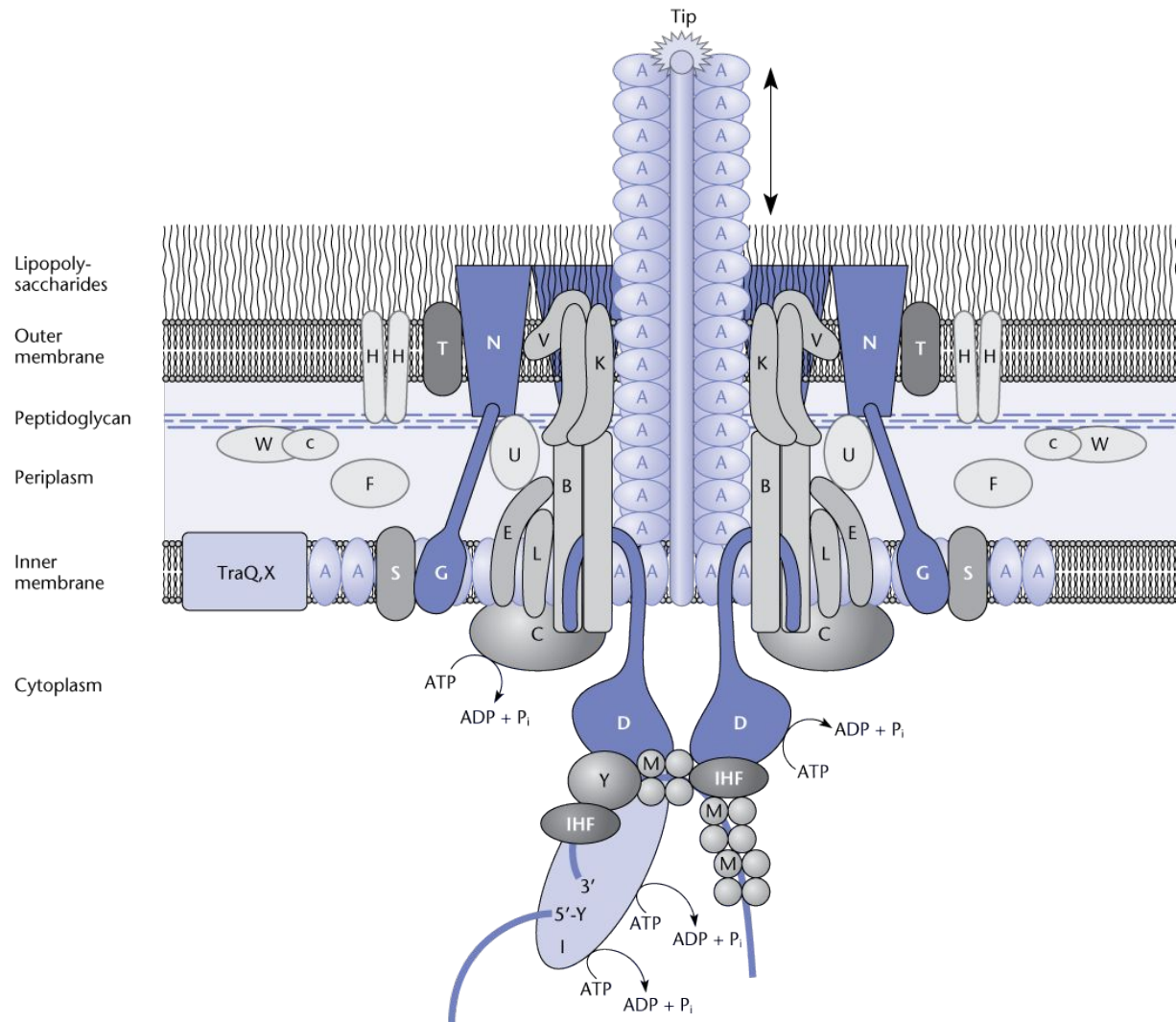
## F-plas mid

1. large (100 kb)
2. low copy (1-2 copies/cell)
3. self transmissible
4. requires protein synthesis  
(chloramphenicol-sensitive)
5. *repE* gene encodes RepE protein
6. RepE protein binds to origin of replication (*oriS*) and initiates DNA replication
7. RepE binds to the *repE* promoter and activates transcription
8. RepE binds to the *copA/incC* locus binding copies of F together via RepE – inhibiting replication (coupling)

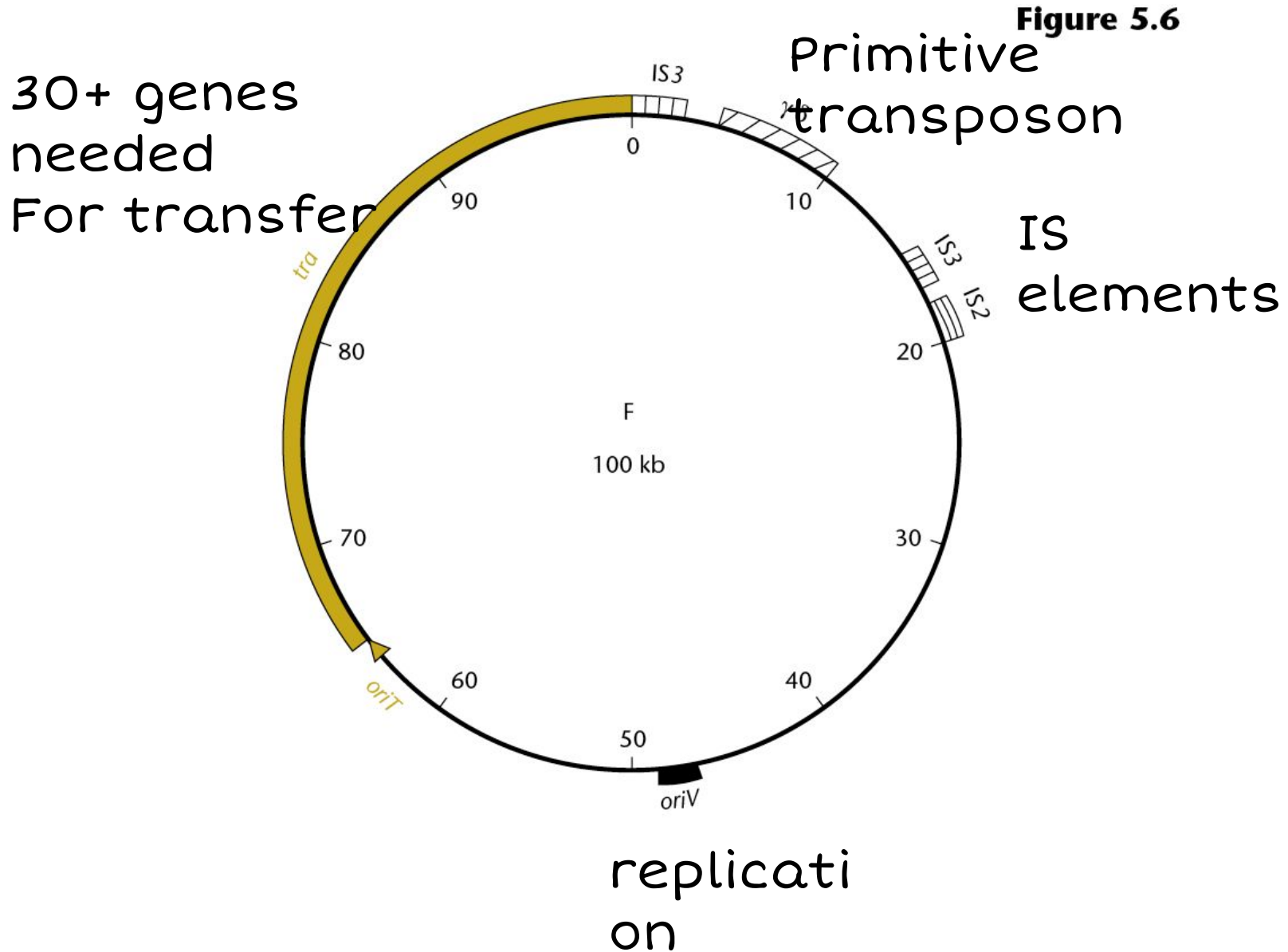
**TABLE 5.1****Some F-plasmid genes and sites**

Symbol	Function
<i>ccdAB</i>	Inhibition of host cell division
<i>incBCE</i>	Incompatibility
<i>oriT</i>	Site of initiation of conjugal DNA transfer
<i>oriV</i>	Origin of bidirectional replication
<i>sopAB</i>	Partitioning
<i>traABCEFGHKLQUVWX</i>	Pilus biosynthesis, assembly
<i>traGN</i>	Mating-pair stabilization
<i>traD</i>	Coupling protein
<i>tral</i>	Relaxase
<i>traYM</i>	Accessories for relaxosome
<i>traJ</i> , <i>finOP</i>	Regulation of transfer
<i>traST</i>	Entry exclusion

# F Pilus assembly

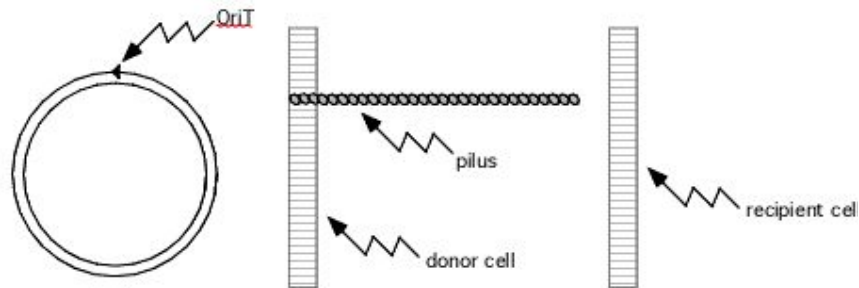



# Genetic organization of F

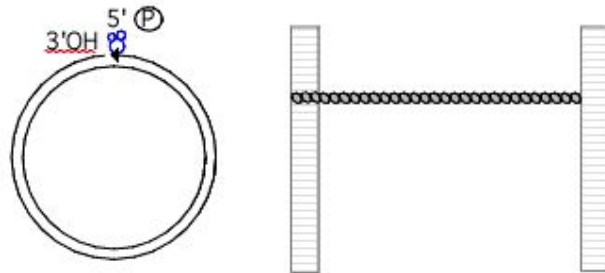




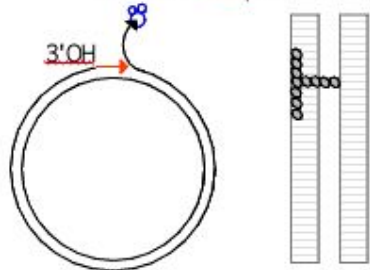
# F-transfer at fine detail



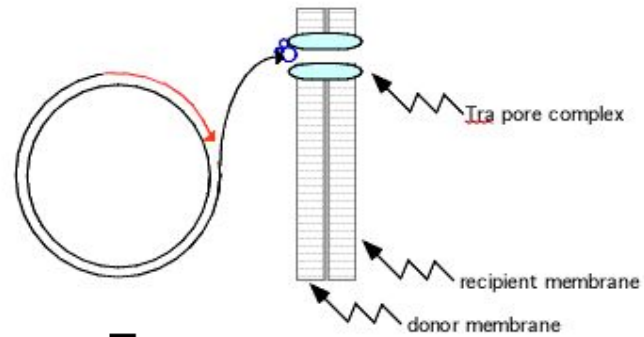
↓ Contact between donor and recipient cells.  
DNA relaxase (  ) nicks at oriT and covalently binds to 5' (P)



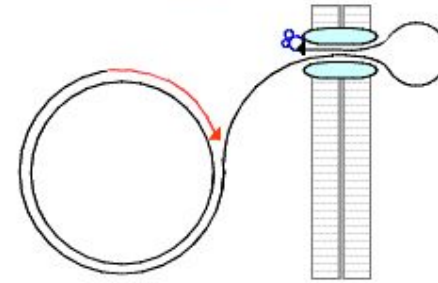
↓ Pilus retracts, bringing donor and recipient into close proximity and Tra proteins form a pore complex that spans the membranes



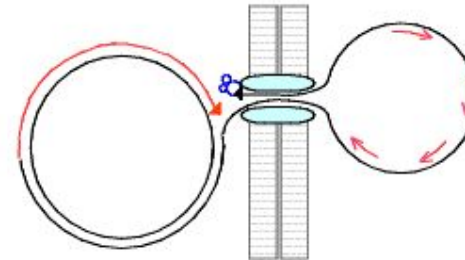
↓ Rolling circle DNA replication initiates at 3'OH and proceeds 5' to  
Membranes brought into close proximity to form mating bridge.  
Relaxase interacts with membrane Tra pore complex



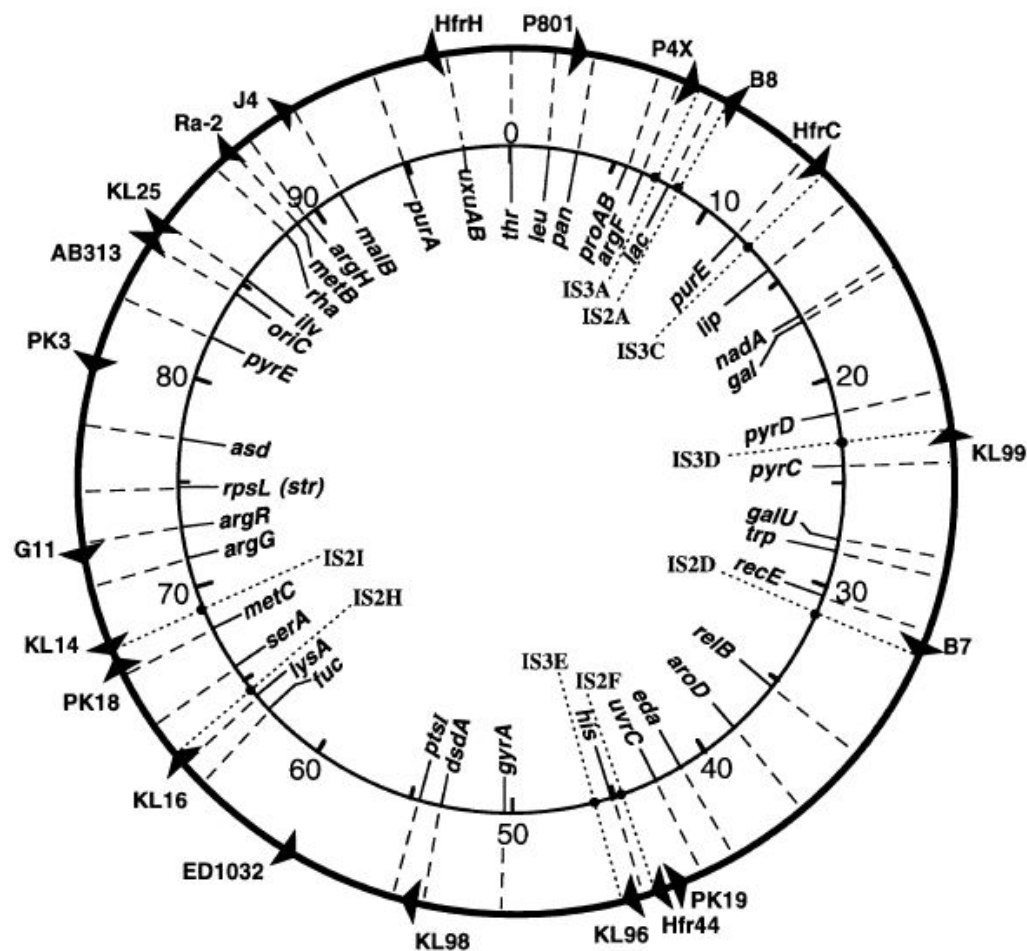
↓ DNA replication pushes the ssDNA into the recipient cell



↓ Lagging strand DNA replication in recipient cell converts ssDNA to dsDNA



↓ Upon complete replication of plasmid, the old and new oriT sites  
"collide", and nicking between oriT sites occurs



**FIGURE 1** Approximate map positions of integrated sex factors (F, F<sub>is</sub>-lac, of ColV) for some Hfr strains. See Table 1 for commonly used derivatives of these strains. The sequence of chromosomal genes transferred from a given strain begins behind the arrowhead; e.g., HfrH transfers genes in the order *uxuAB*, *thr*, *leu*, etc. The positions of the IS sequences which appear to correlate with the sites of F insertion for some of the Hfrs are indicated and can be found on the physical map in chapter 129.