

Эволюционная иммунология
Лекция 7 «Трансплантационный
иммунитет»

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26 марта 2018

Временные параметры и особенности отторжения алло(ксено)трансплантатов у животных различных типов

| Тип, класс | Вид | Алло-, ксено- трансплантат | Время выживания, дни | | Форма отторжения | |
|---|---|---|------------------------|------------------------|---|---|
| | | | Первичный трансплантат | Вторичный трансплантат | | |
| Губки | <i>Ephydatia fluviatilis</i> | Ал. | — | — | Образование коллагеноподобного слоя | |
| | <i>Axinella polypoides</i> | Ал. | — | — | Образование зоны некроза | |
| | <i>Ephydatia fluviatilis</i> , <i>Eph. mulleri</i> , <i>Spongilla lacustris</i> , <i>Eunapius fragilis</i> | Кс. | — | — | То же | |
| | <i>Hymeniacidon sinapium</i> | Ал. | — | — | » | |
| | <i>Callispongia diffusa</i> | Ал. | 11,3 | 7,0 | » | |
| | <i>Xestospongia exigus</i> | Ал. | 8,2 | 3,2 | » | |
| | <i>Hymeniacidon perleve</i> | Ал. | 10,4 | 7,0 | » | |
| | Кишечно- полостные | <i>Hydractinia echinata</i> | Ал. | — | — | Гиперпластический рост |
| | | <i>Anthopleura elegantissima</i> , <i>Anth. krebsi</i> , <i>Leptogorgia vitgulata</i> | Ал., Кс. | — | — | Гиперпластический рост или образование зоны некроза |
| <i>Eunicella stricta</i> , <i>Lophogorgia sarmentosa</i> | | Ал., Кс. | — | — | Образование зоны некроза | |
| <i>Montipora verrucosa</i> | | Ал. | 22,0 | 11,6 | То же | |
| Немертины | | <i>Lineus sanguineus</i> → <i>L. ruber</i> | Кс. | 15,4 | 8,6 | Образование зоны некроза, депигментация ткани |
| | | <i>L. ruber</i> → <i>L. sanguineus</i> | Кс. | 16,4 | 8,6 | То же |
| Членистоногие | <i>Blamberus giganteus</i> → <i>Periplaneta americana</i> | Кс. | >3 | — | Инкапсуляция | |
| Иглокожие | <i>Cucumaria tricolor</i> | Ал. | 129—185 | ≈50 | Образование зоны некроза, депигментация ткани | |
| | <i>Dermasterias imbricata</i> | Ал. | — | — | То же | |
| | <i>Strongylocentrotus droebachiensis</i> | Ал. | 30 | 12 | » | |
| | Оболочники | <i>Styela plicata</i> | Ал. | 38 | 28 | » |

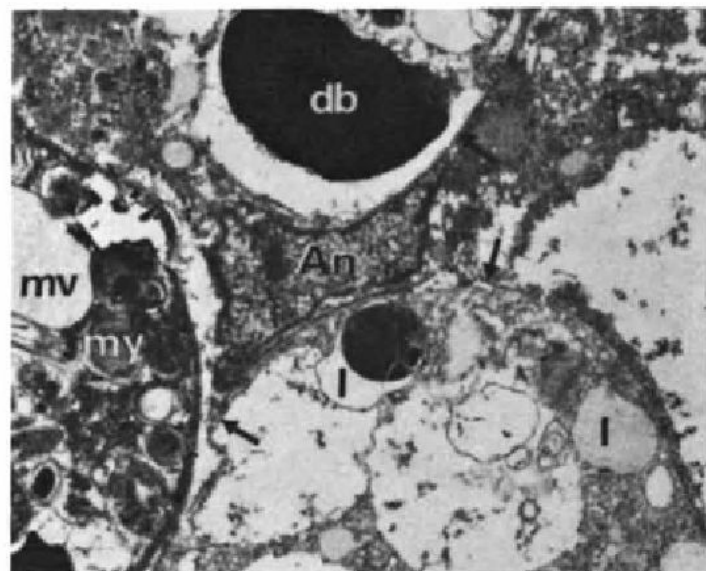
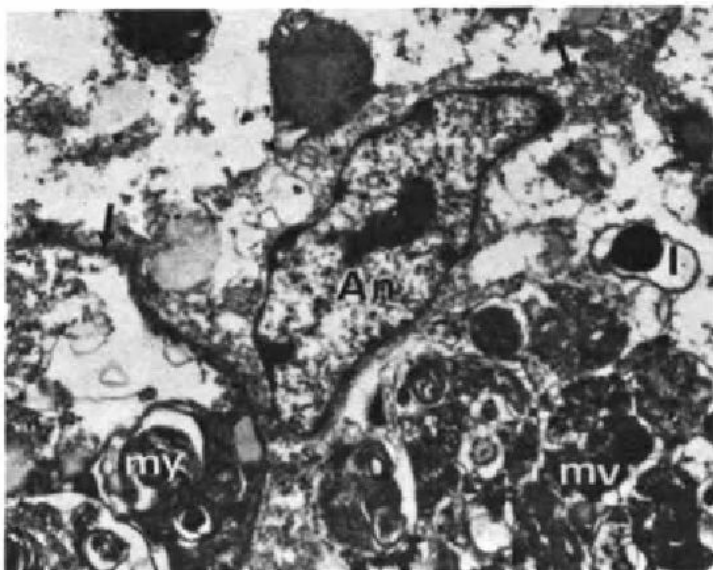
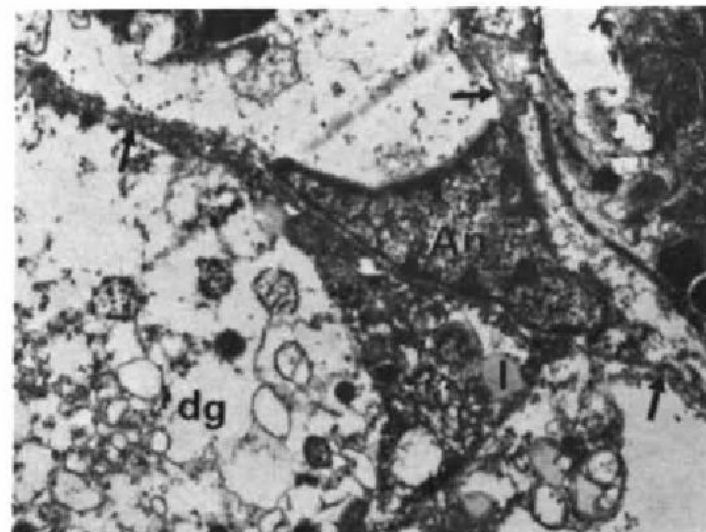
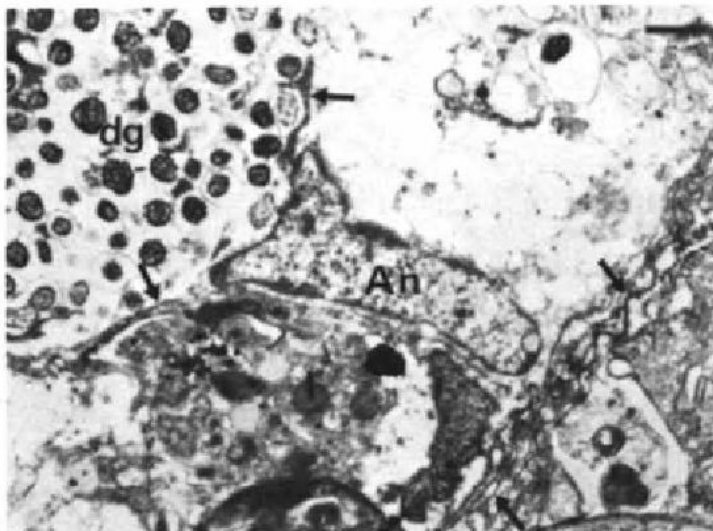
Table 1. Phyletic survey of chimera formation and somatic tissue compatibility*

| Group | Mechanism of chimera formation | Evidence of somatic tissue compatibility system [†] |
|------------------|--------------------------------|--|
| Protists: | | |
| Dictyostelids | Coaggregation, mutation | Failure to coaggregate, separation during migration or culmination |
| Myxomycetes | Plasmodium fusion | Failure of plasmoidia to fuse |
| Fungi: | | |
| Phycomycetes | Mutation | Failure of hyphal fusion |
| Ascomycetes | Hyphal fusion | |
| Basidiomycetes | Hyphal fusion | |
| Plants: | | |
| Rhodophyta | Sporeling | Root fusion |
| Gymnosperms | coalescence | |
| Angiosperms | | |
| Animals: | | |
| Porifera | Larval fusion | Failure of fusion, strain-specific reaggregation |
| Coelenterates | Planulae fusion | Failure to fuse |
| Annelids | | Graft rejection |
| Molluscs | | |
| Echinoderms | | |
| Arthropods | | Graft rejection |
| Chordates | | |
| Ascidians | Colony fusion | Failure to fuse |
| Vertebrates | Bovine twins, malignancy | Lymphocyte-mediated immune response |

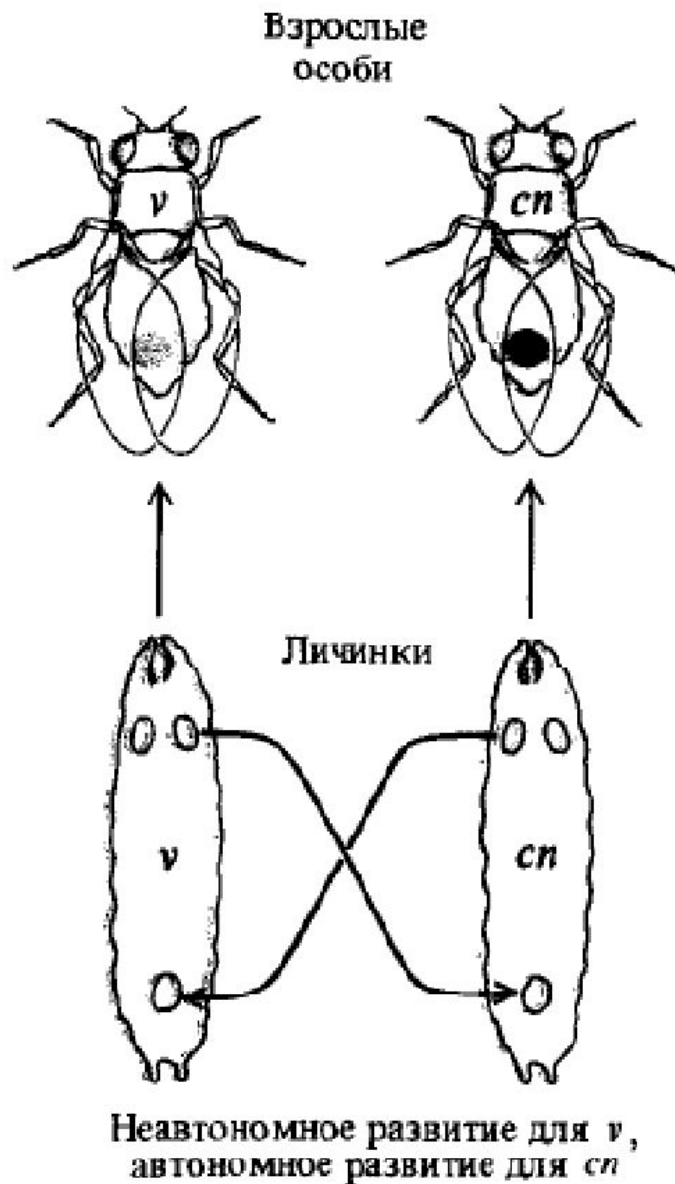
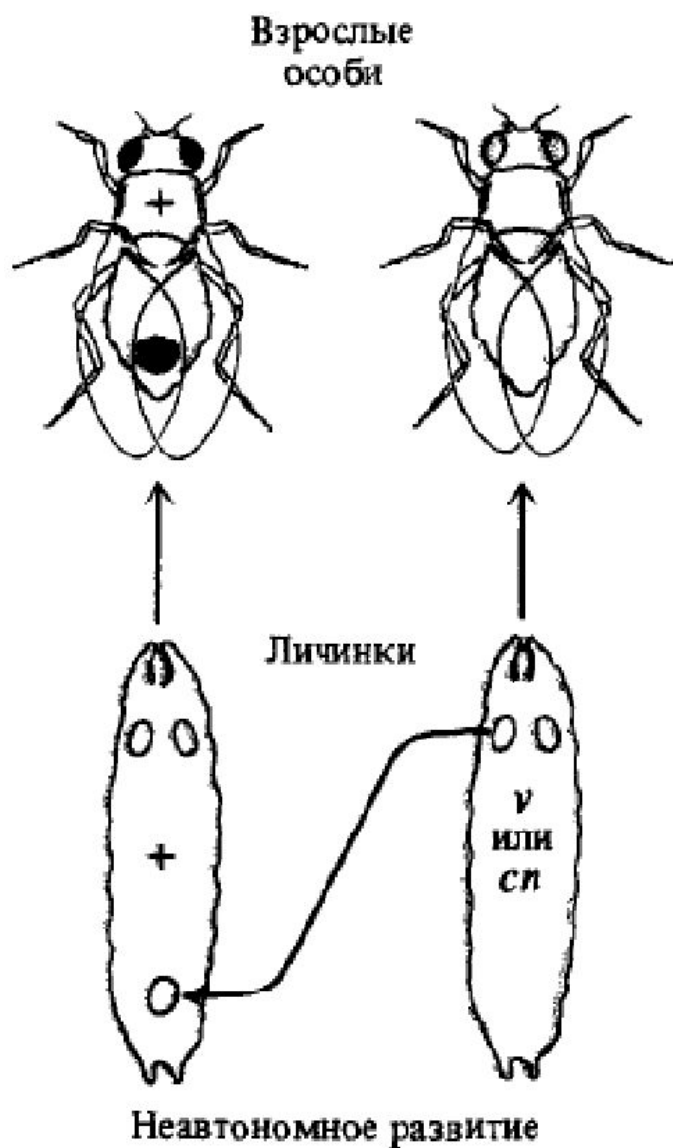
TABLE 1: Invertebrates exhibiting induction, specificity, and/or immunological memory in the nonpathogenic context of first and second challenges with transplants (n.a.: not analyzed).

| Species | Challenge | Specificity | Memory | References |
|--|-------------------------------|-------------|--------|--|
| Porifera | | | | |
| <i>C. diffusa</i> | Tissue (allograft) | + | + | Smith and Hildemann, 1986 [35] |
| <i>G. cydonium</i> | transplantation | + | n.a. | Müller et al., 1999 [36] |
| Cnidaria | | | | |
| <i>E. stricta</i> | Colonial contact/allograft, | + | n.a. | Theodor, 1970 [37] |
| <i>M. verrucosa</i> | xenograft | + | + | Hildemann et al., 1977 [38] |
| Nemertea | | | | |
| <i>L. ruber</i> | Tissue (allograft, xenograft) | + | + | Bierne and Langlet, 1974 [39]; |
| <i>L. lacteus</i> | transplantation | | | Langlet and Bierne, 1975 [40]; 1982 [41]; 1984 [42] |
| Annelida | | | | |
| Earthworms <i>L. terrestris</i> <i>E. fetida</i> | Tissue (allograft, xenograft) | + | + | Cooper, 1969 [43]; Cooper and Roch, 1986 [44] |
| Leeches <i>H. medicinalis</i> <i>G. complanata</i> | Tissue (allograft, xenograft) | + | + | Tettamanti et al., 2003 [45] |
| Mollusca | | | | |
| <i>I. fruhstorferi</i> | Tissue (allograft) | + | n.a. | Yamaguchi et al., 1999 [46] |
| Arthropoda | | | | |
| <i>P. americana</i> <i>B. orientalis</i> | Tissue (allograft, xenograft) | + | + | Hartmann and Karp, 1989 [47]; Karp and Meade, 1993 [48] |
| Echinodermata | | | | |
| <i>S. purpuratus</i> <i>L. pictus</i> | Tissue (allograft) | + | – | Coffaro and Hinegardner, 1977 [49] |
| <i>D. imbricata</i> | transplantation | + | + | Karp and Hildemann, 1976 [50] |
| Tunicata | | | | |
| <i>B. schlosseri</i> | Colonial contact/allograft | + | n.a. | Rinkevich et al., 1998 [51]; Scofield et al., 1982 [52]; |
| <i>S. plicata</i> | | + | + | Raftos et al., 1987 [53]; 1988 [54] |

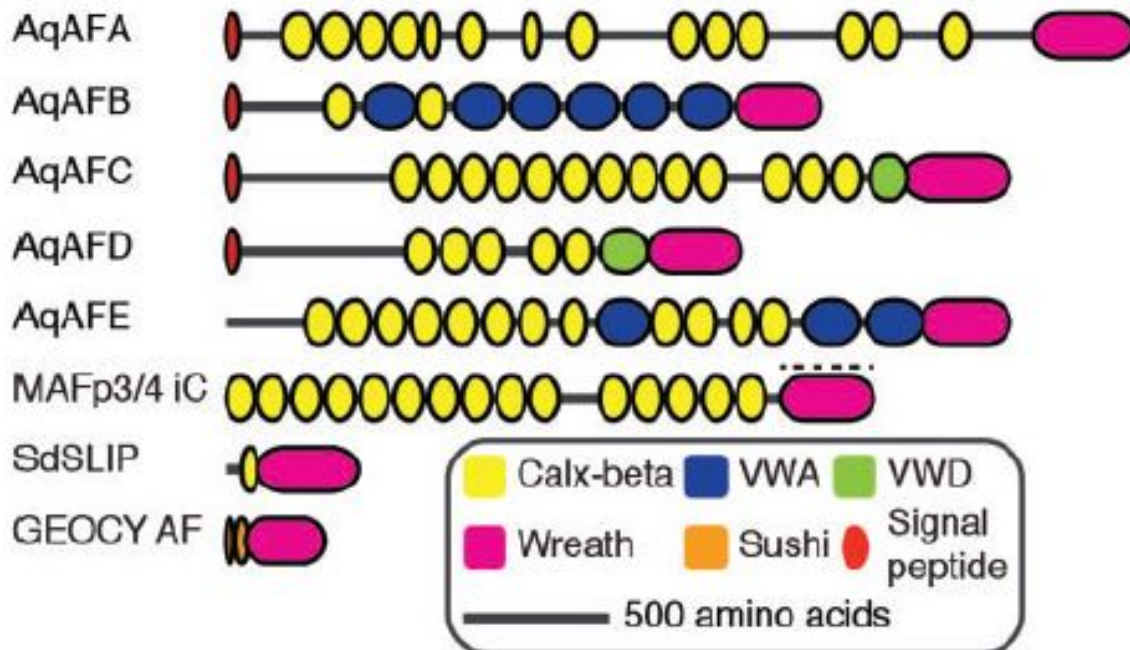
Амебоциты инфильтрировались в аллотрансплантант немертины



Пересадка имагинальных дисков дрозофилы



Факторы агрегации губок



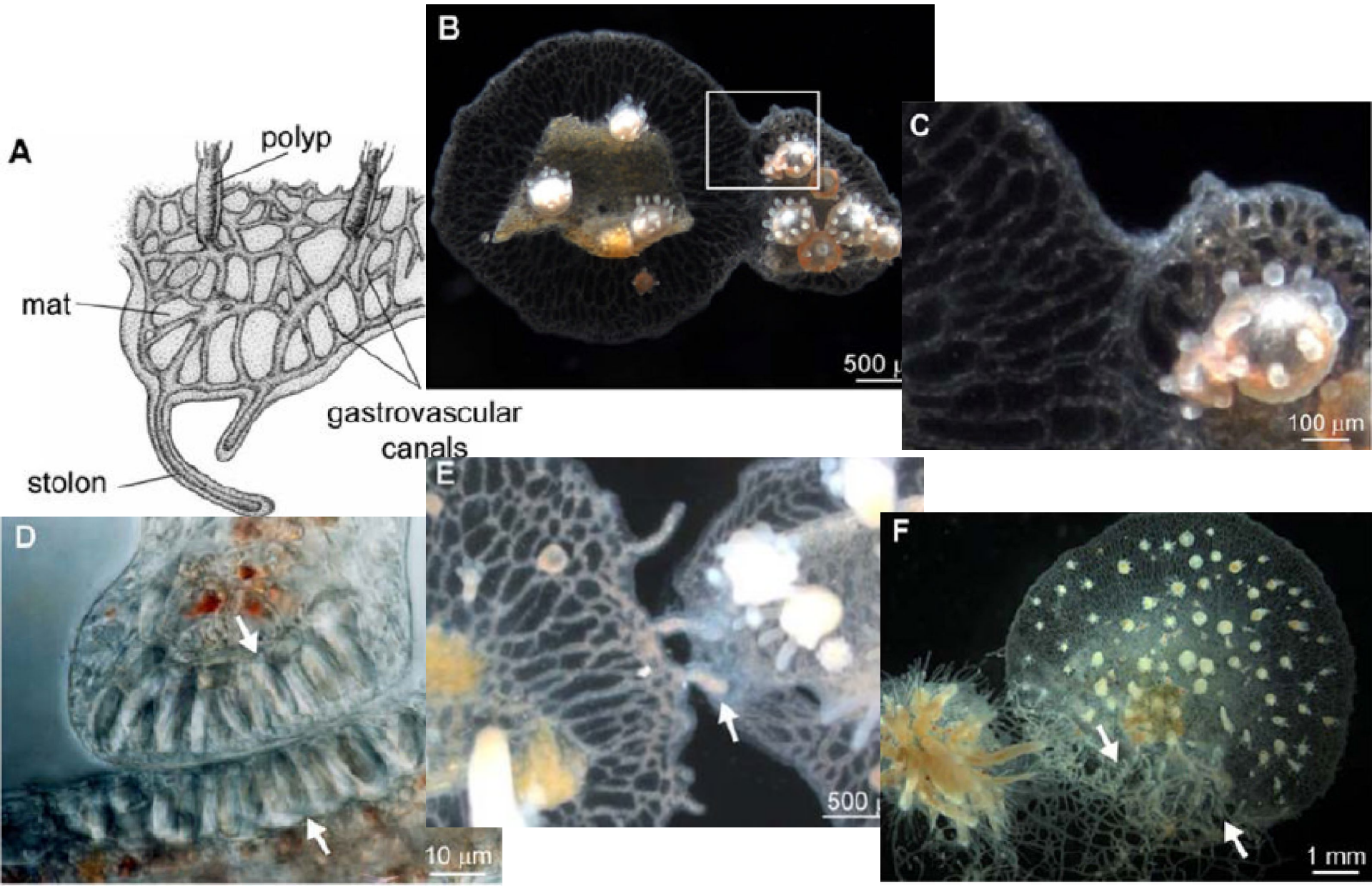
Amphimedon sp.



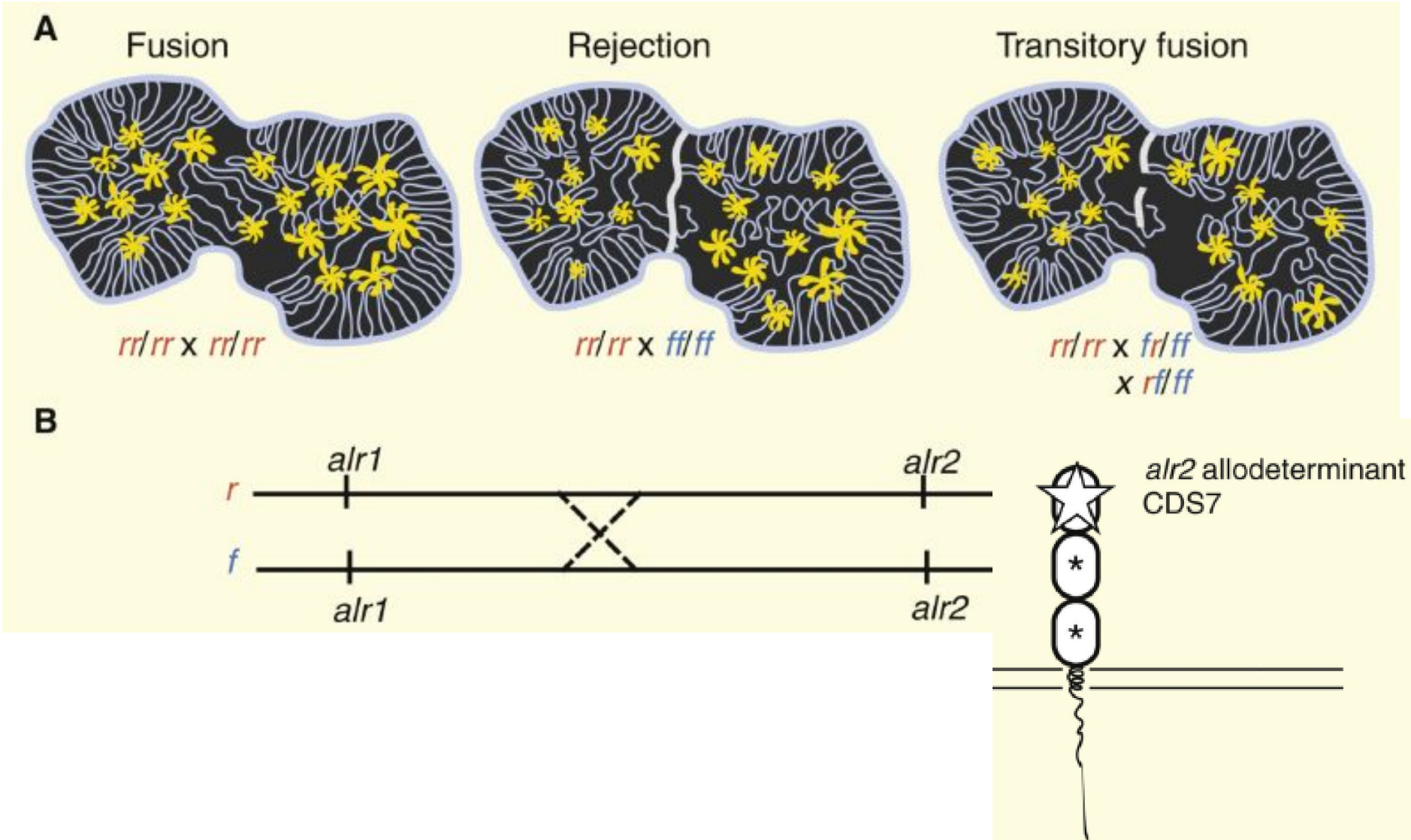
Clathria sp.

FIG. 1. Domain architecture of aggregation factor proteins. Aggregation factor proteins from *Amphimedon queenslandica* (AqAFA–AqAFE), *Clathria prolifera* (MAFp3/4 iC), *Suberites domuncula* (SdSLIP) and *Geodia cydonium* (GEOCY AF) are shown. Colored shapes represent predicted protein domains and sequence features. Models are approximately to scale. For *C. prolifera*, MAFp3 (indicated by dashed line) and MAFp4 are represented as a single contiguous sequence; the longest isoform (isoform C) is shown. iC, isoform C; VWA, von Willebrand type A domain; VWD, von Willebrand type D domain.

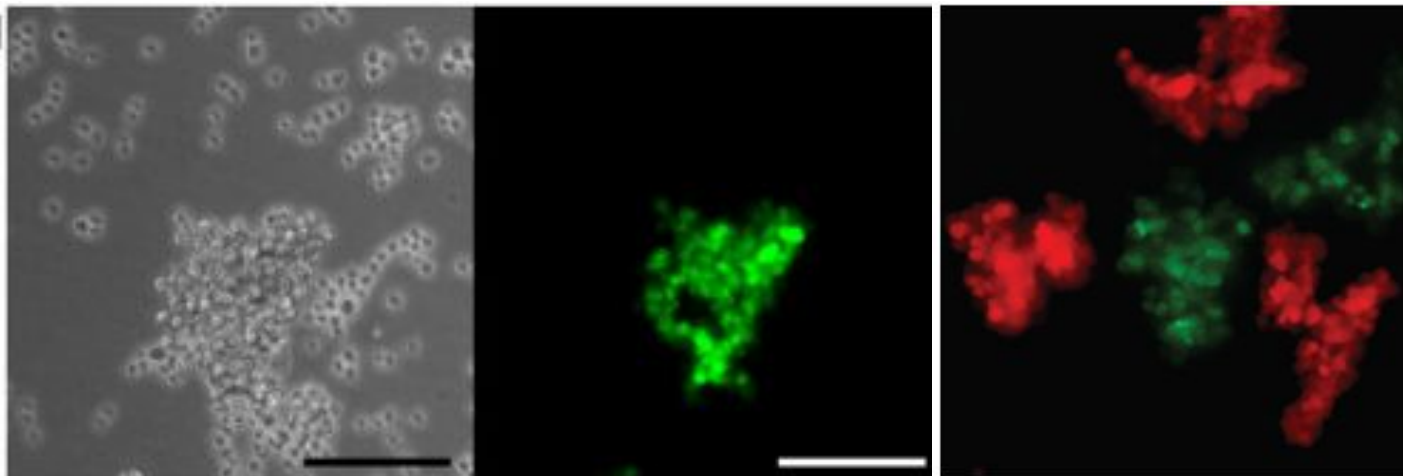
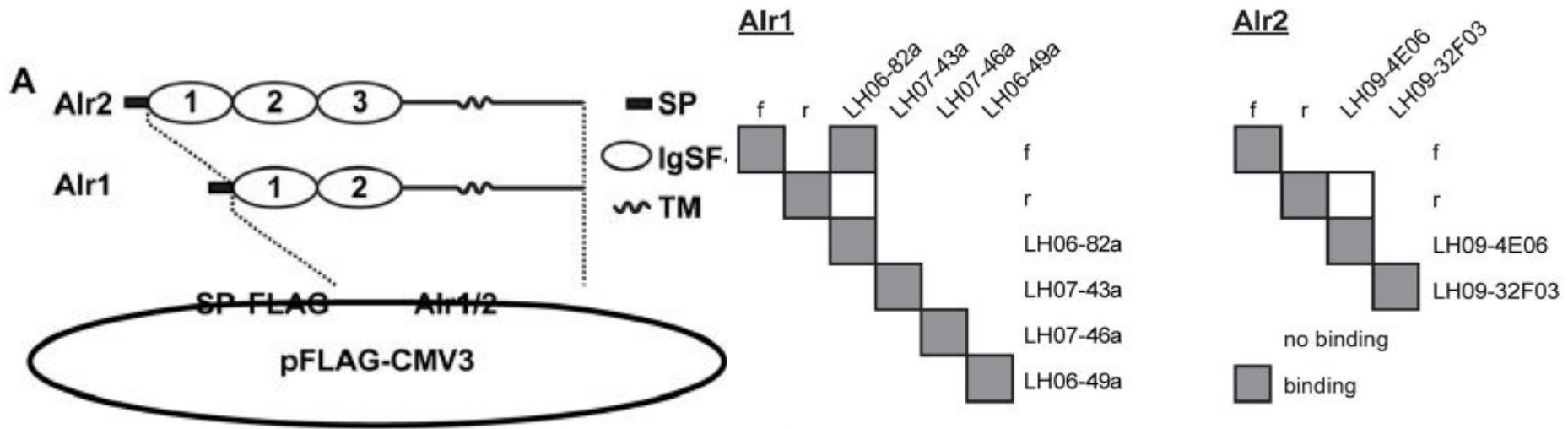
Слияние (верхний ряд) и отторжение (нижний) колоний у *Hydractinia*



Отторжение чужих колоний у гидроида *Hydractinia* можно объяснить системой всего двух локусов, *alr1* и *alr2*

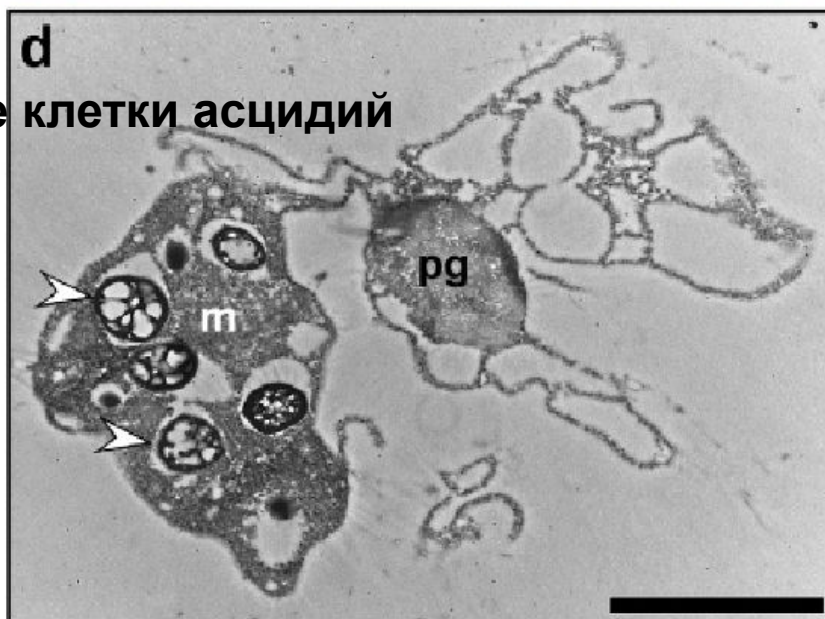
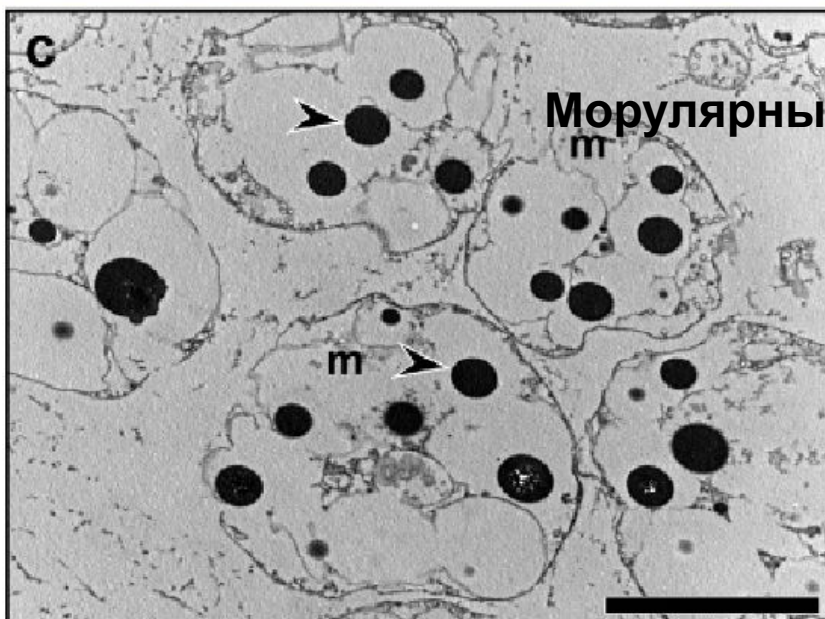
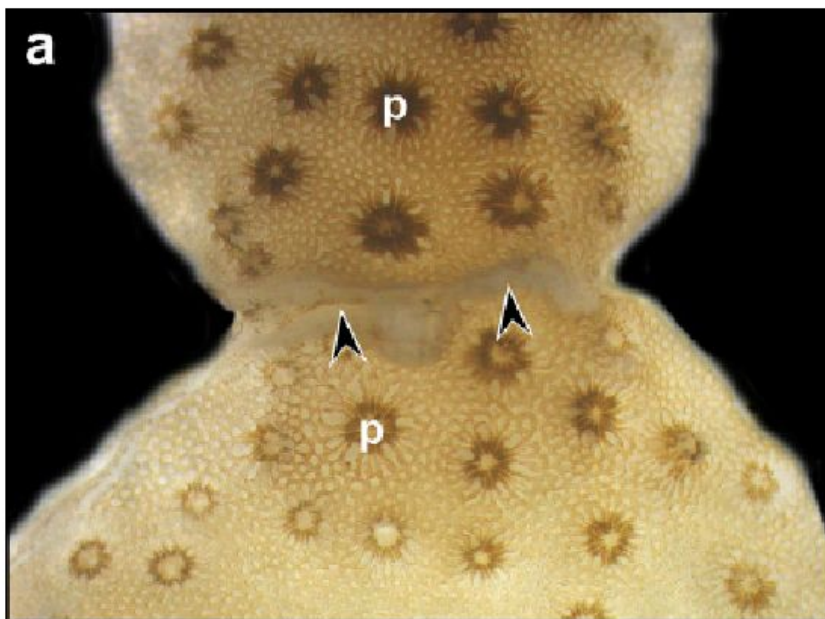


Гомофильное связывание изоформ Alr1 и Alr2 обеспечивает сегрегацию клеток

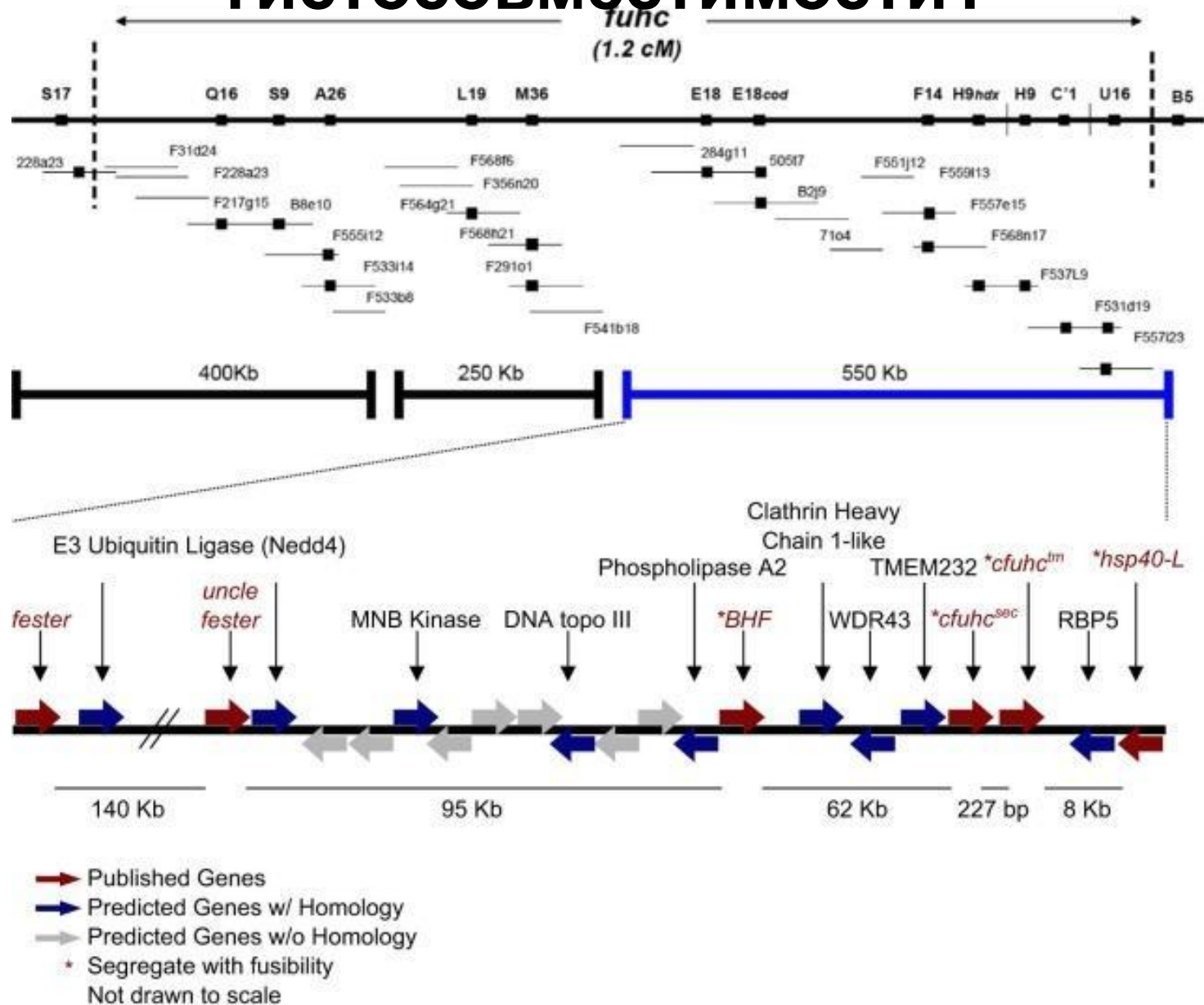


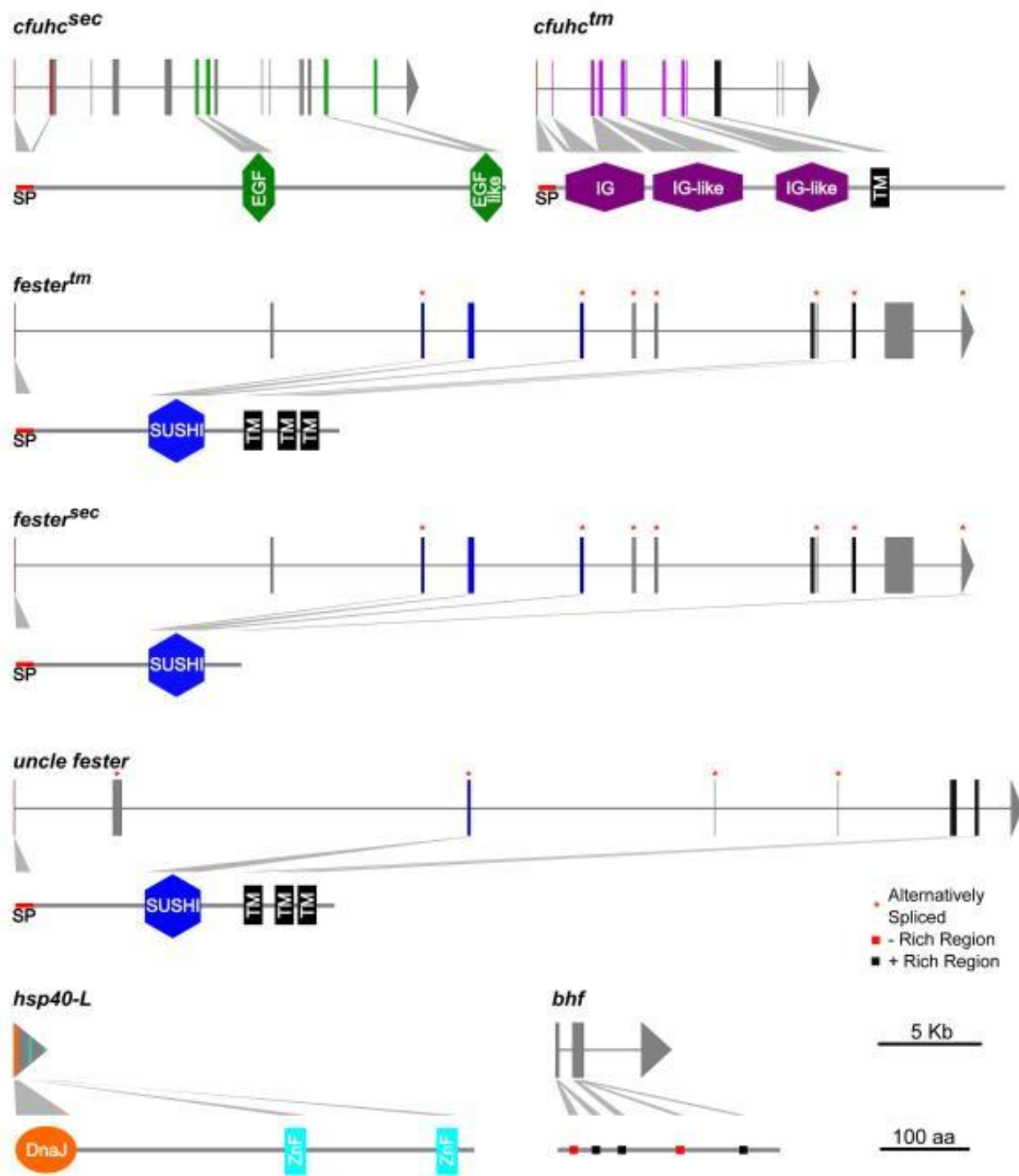
Агрегаты клеток СНО, трансфицированных флуоресцентным маркером и специфической изоформой Alr1

Полип *Stylophora* и асцидия *Botryllus*

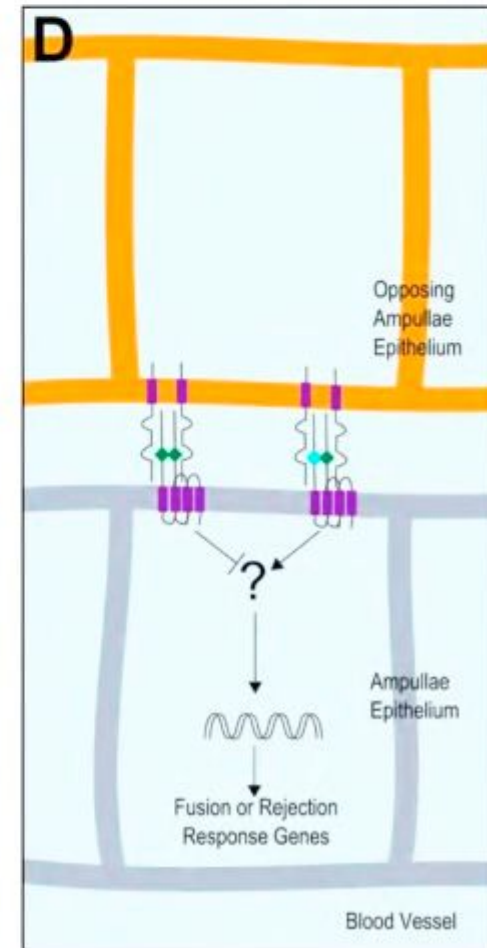
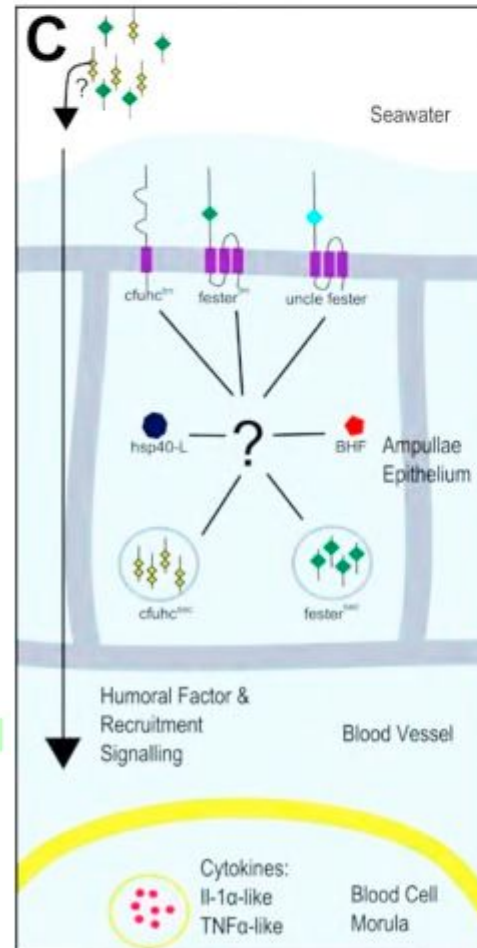
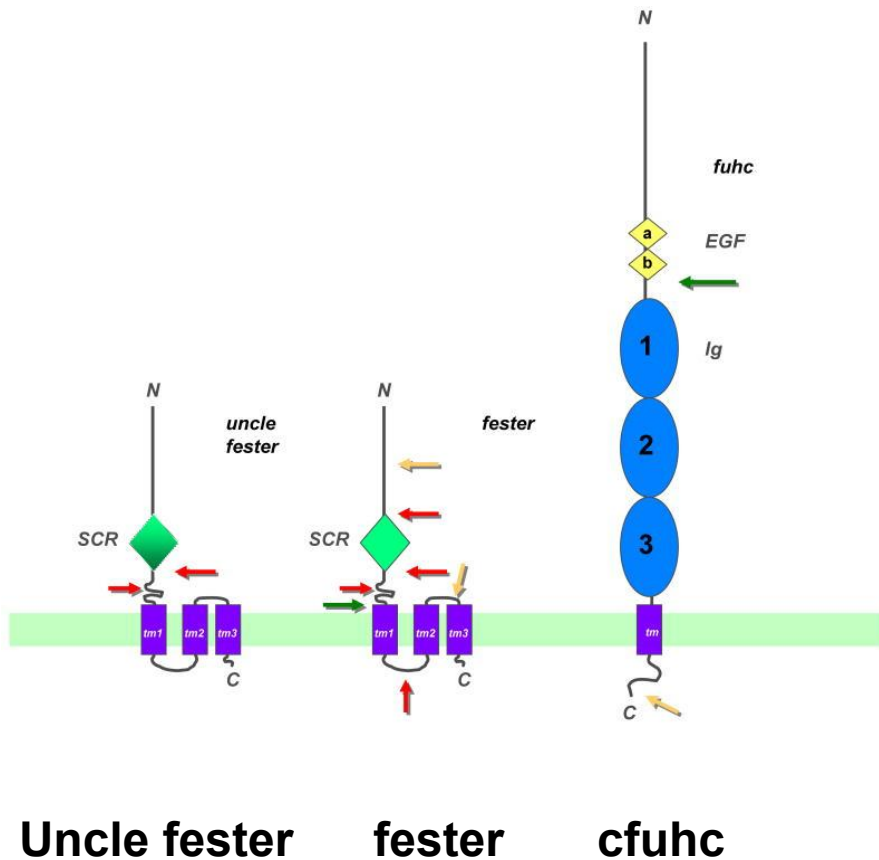


Локус *fuhc* асцидий содержит 6 генов гистосовместимости?

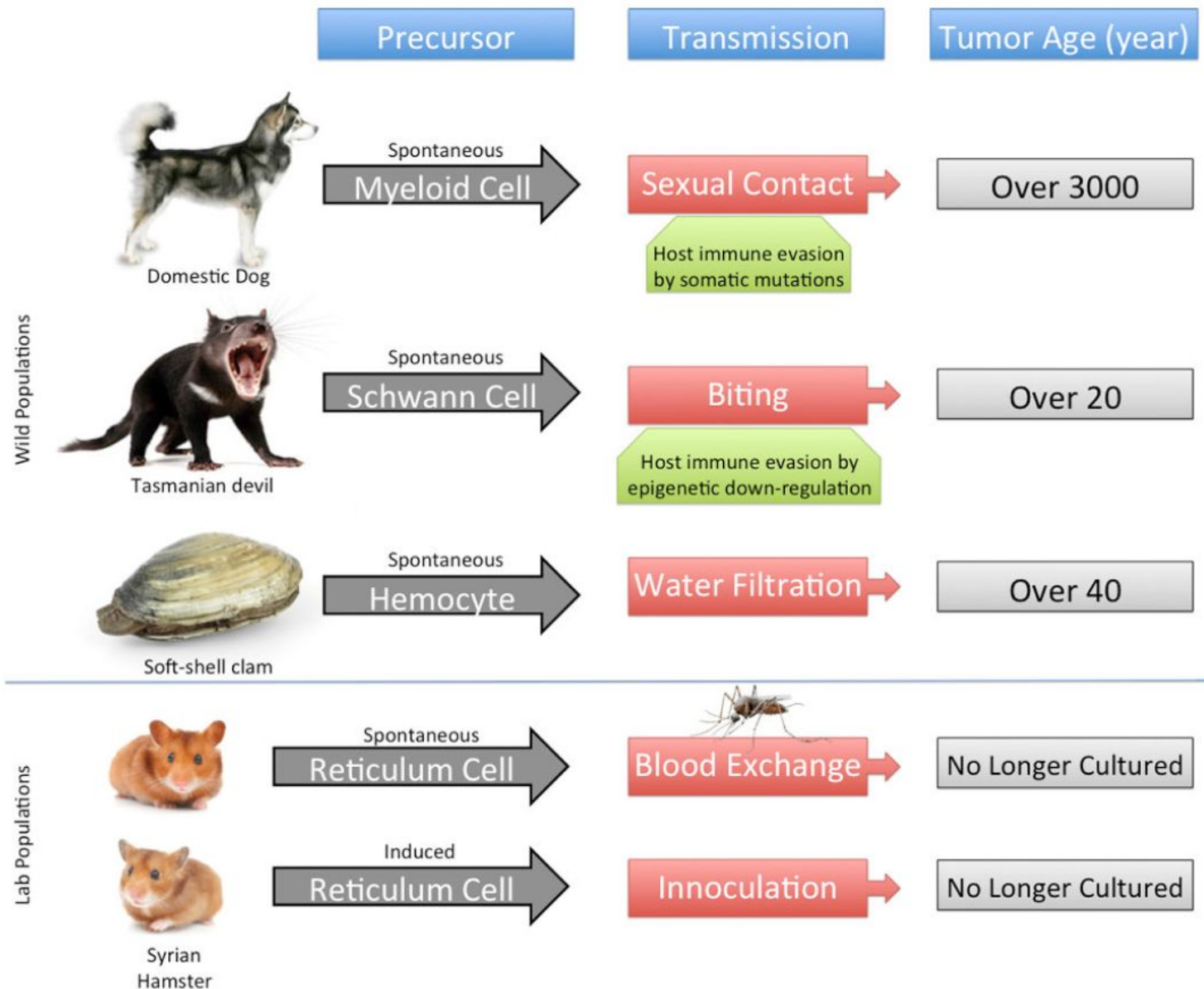




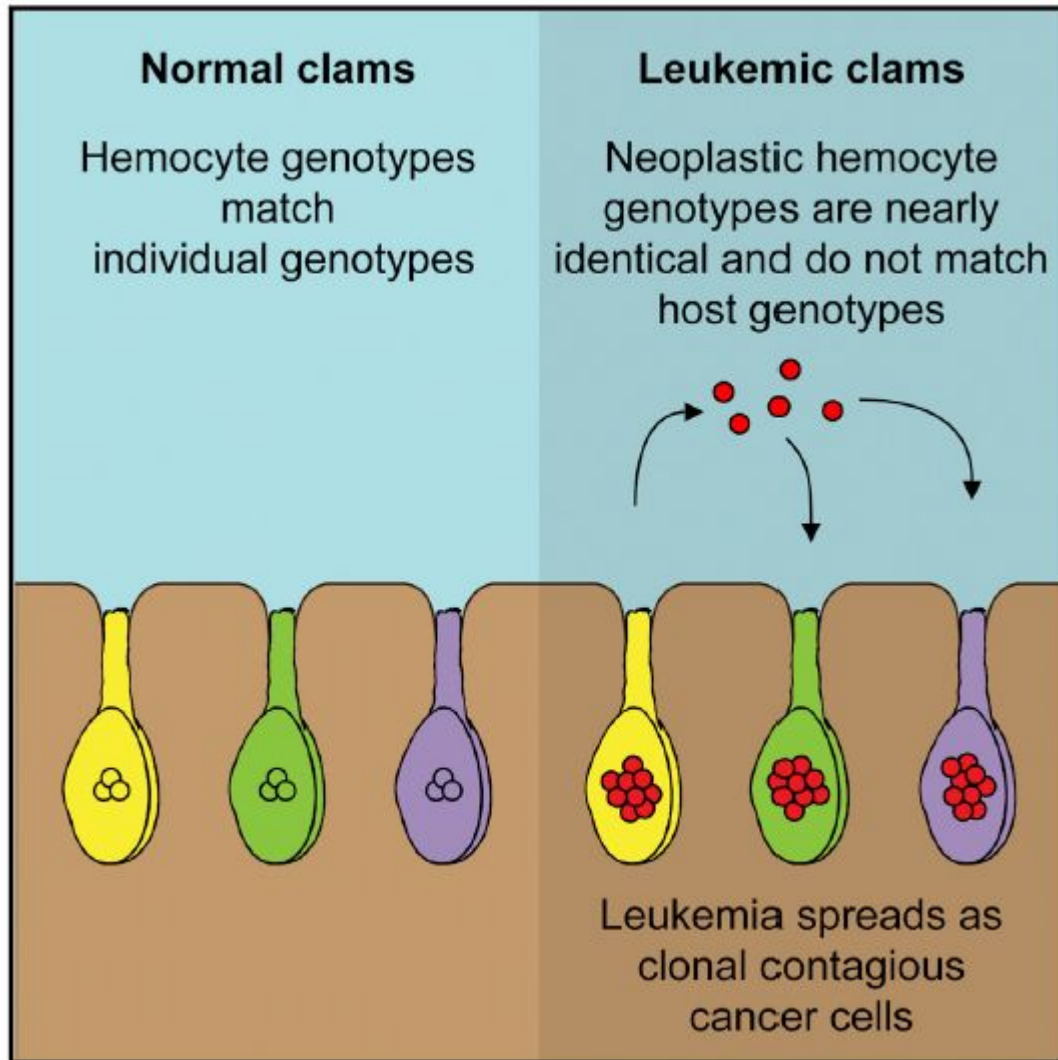
Адгезионные молекулы асцидий



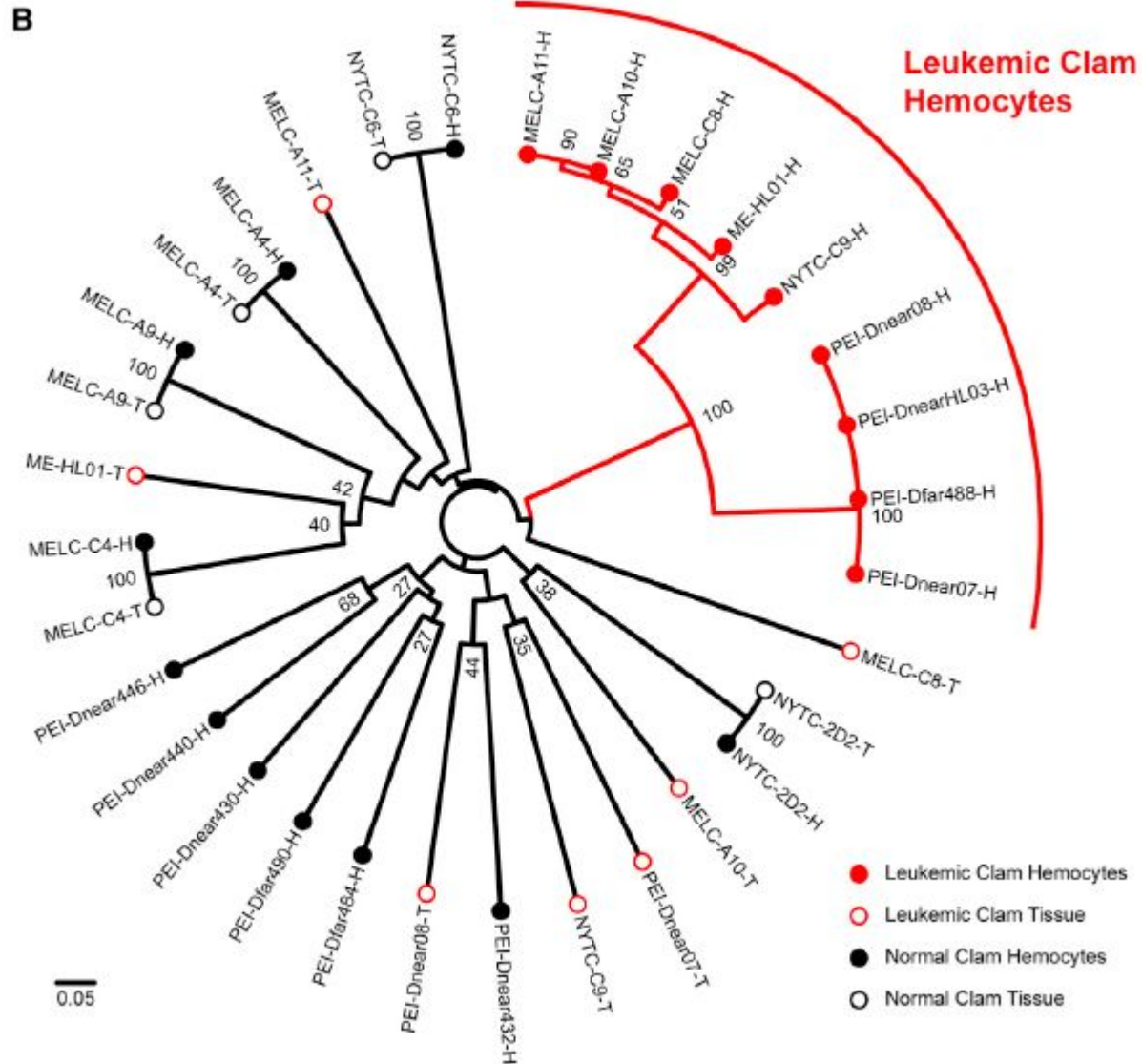
Трансмиссибельные опухоли



Лейкемия моллюсков *Mya arenaria*



Опухолевые клетки имеют множественные копии транспозона Steamer и демонстрируют общее происхождение

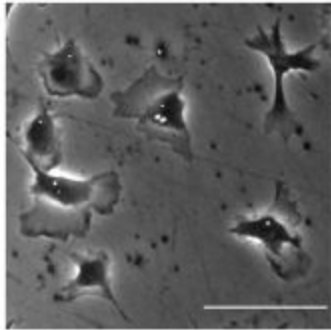


Гольфстрим разносит опухолевые клетки моллюсков по мировому океану?

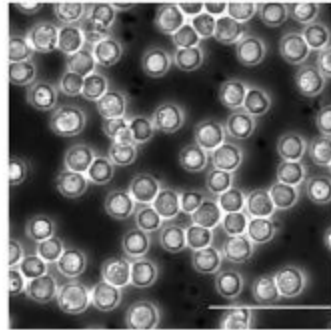
A



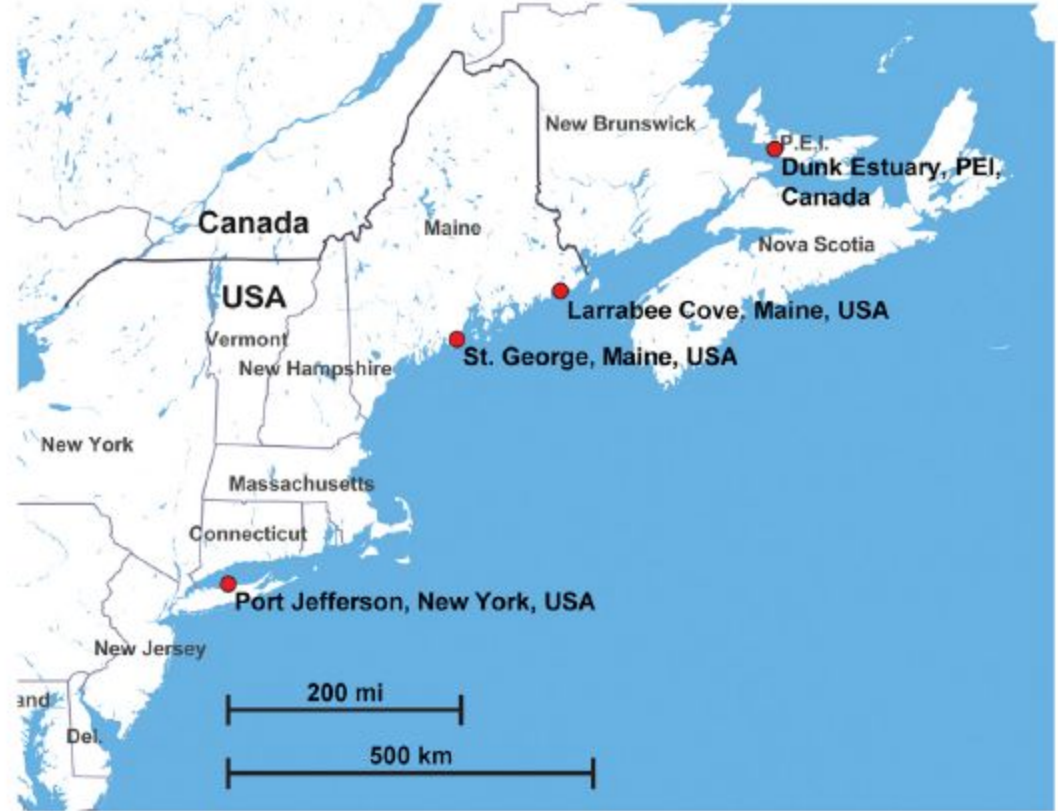
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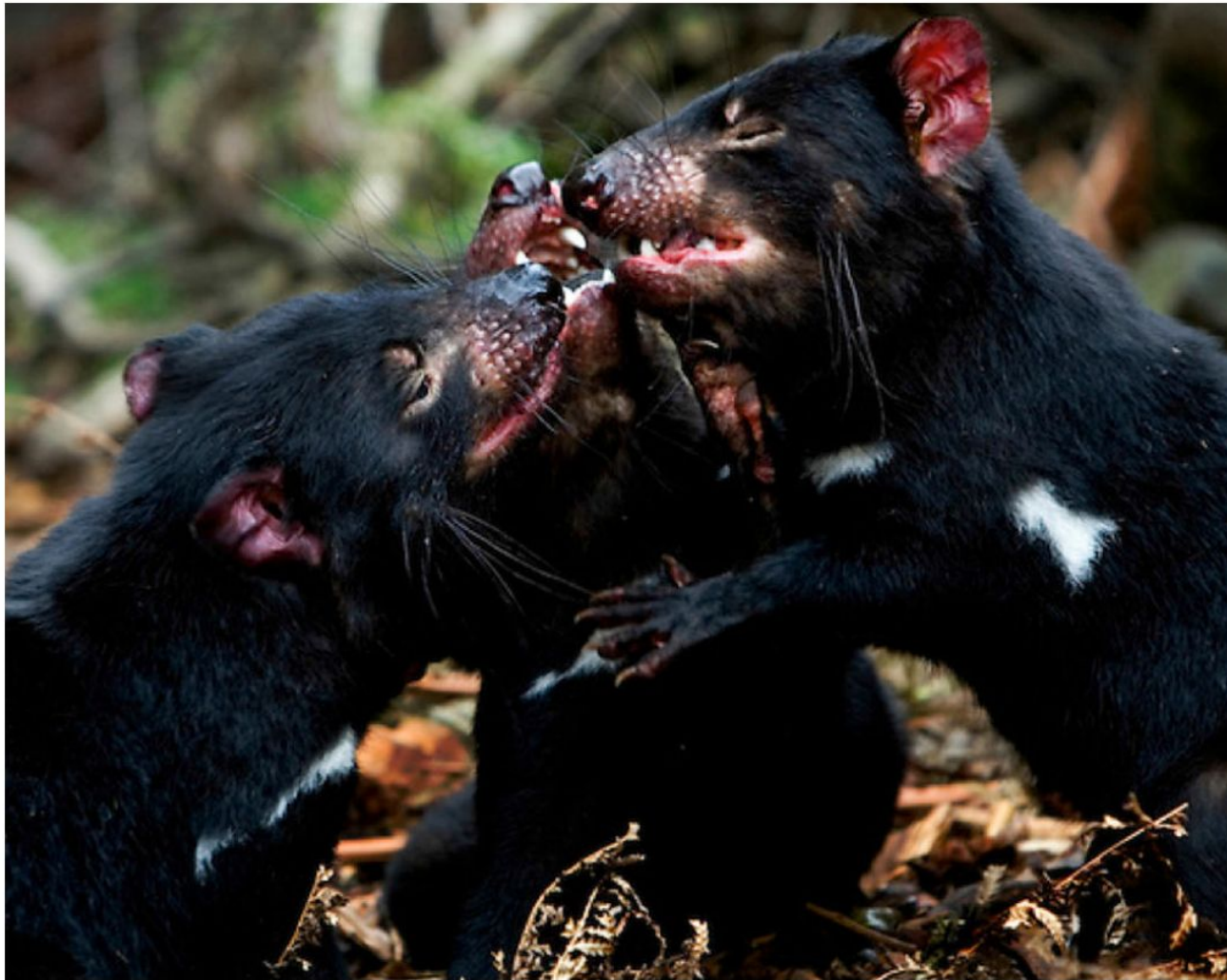
C



D



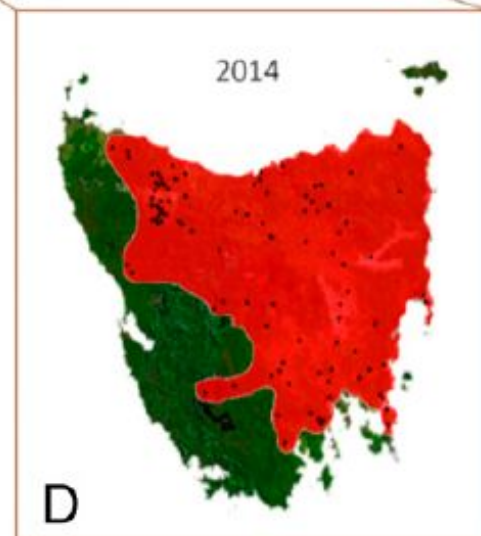
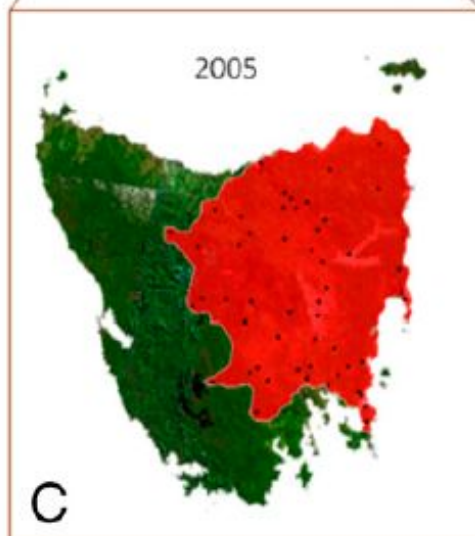
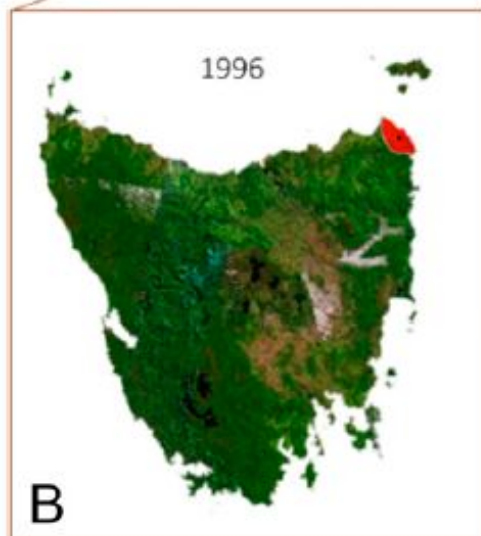
DFTD - Devil Facial Tumour Disease



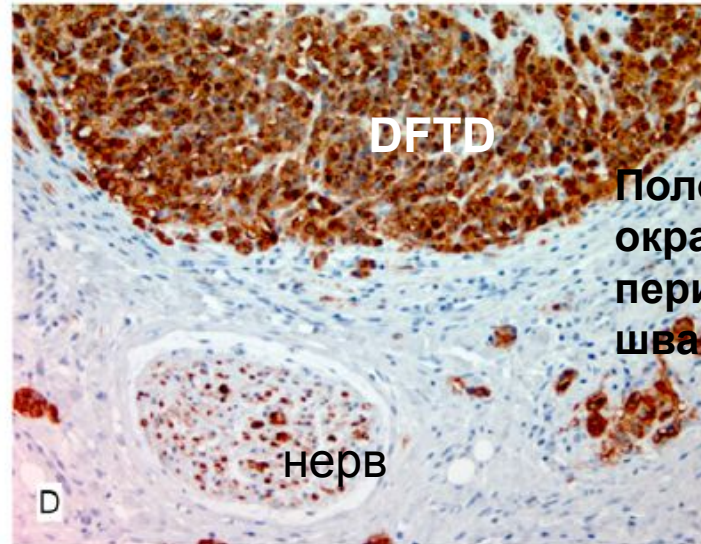
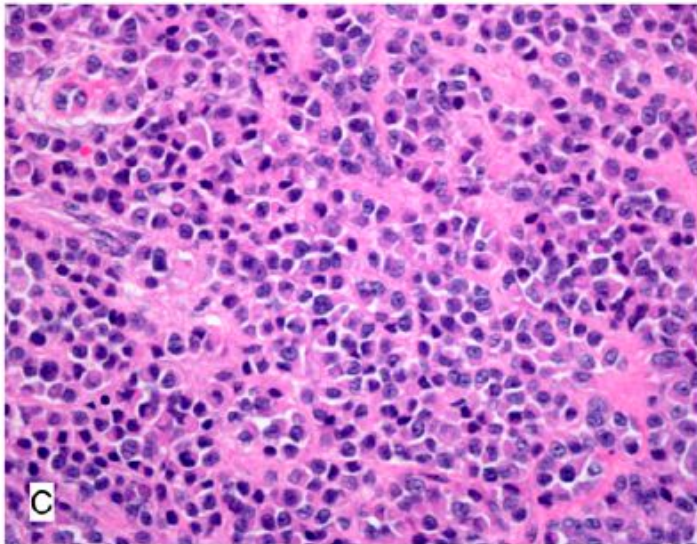
Высокоагрессивная опухоль, возникшая на основе Шванновской клетки, впервые описана в 1996 году, передается при укусах.

Распространение DFTD

A



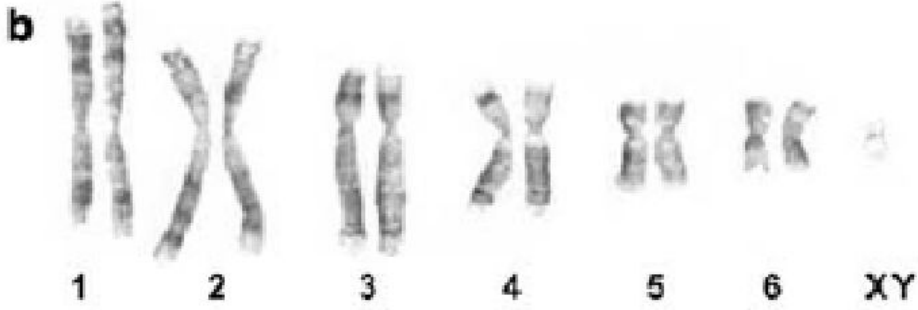
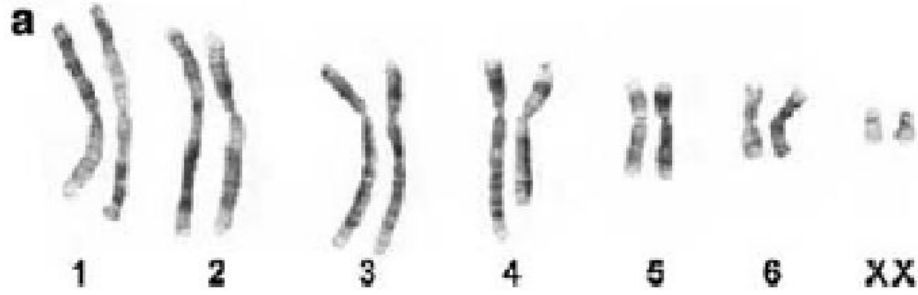
Происхождение DFTD доказывается с помощью иммуногистохимии



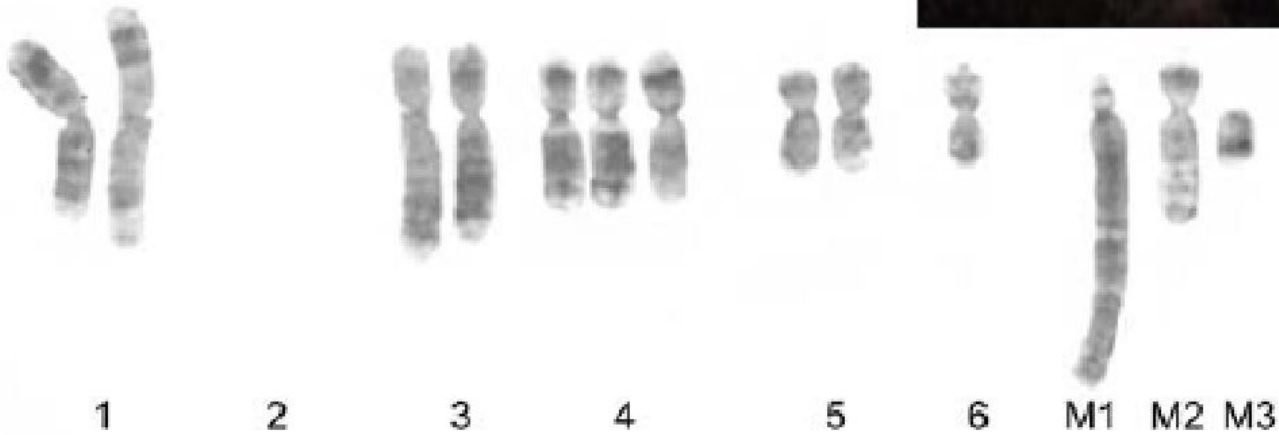
Положительное окрашивание на периаксин - белок шванновских клеток

Клональность DFTD

A



B



Таманские дьяволы действительно высоко изогенны по генам МНС

Table 3. MHC genotyping of Tasmanian devils used for skin graft experiments.

| Tasmanian devil ID | MHC I $\alpha 1$ sequence variants | MHC II $\beta 1$ sequence variants |
|--------------------|------------------------------------|------------------------------------|
| TD 190 | Sahal*27, 28, 32, 35, 49 | SahaDAB*01, 03, 05 |
| TD 199 | Sahal*27, 28, 32, 35, 49 | SahaDAB*01, 03, 05 |
| TD 187 | Sahal*28, 32, 35, 49, 57 | SahaDAB*01, 03, 05, 12 |
| TD 200 | Sahal*27, 28, 32, 35, 49 | SahaDAB*01, 03, 05, 11 |
| TD 188 | Sahal*27, 32, 35, 49 | SahaDAB*01, 03, 05, 15 |
| TD 189 | Sahal*27, 32, 35, 48 | SahaDAB*01, 03, 05, 11, 15 |
| TD 191 | Sahal*28, 32, 34, 48, 49 | SahaDAB*01, 03, 05, 13 |

TD 190 and TD 199 shared all MHC I and II alleles. The remaining devil pairs had two to three MHC I allelic mismatches and one to two MHC II allelic mismatches.
doi:10.1371/journal.pone.0022402.t003

Table 4. Amino acid difference count at peptide binding sites and MLR results within skin graft devil pairs.

| Tasmanian devil ID | Amino acid difference count at peptide binding sites at MHC I $\alpha 1$ | Amino acid difference count at peptide binding sites at MHC II $\beta 1$ | Mixed lymphocyte reaction (SI) |
|--------------------|--|--|--------------------------------|
| TD 190 and TD 199 | 0 | 0 | 1 |
| TD 187 and TD 200 | 0 | 0 | 1 |
| TD 188 and TD 189 | 2 | 0 | 17 |
| TD 190 and TD 191 | 0 | 0 | 5 |

TD 188 and TD 189 had two amino acid differences at peptide binding sites at MHC I $\alpha 1$ and had a strong MLR response. The other three pairs did not have amino acid difference count at peptide binding sites at MHC I $\alpha 1$ or MHC II $\beta 1$ and had low MLR responses.

doi:10.1371/journal.pone.0022402.t004

Разницы генотипов между больными и здоровыми дьяволами нет

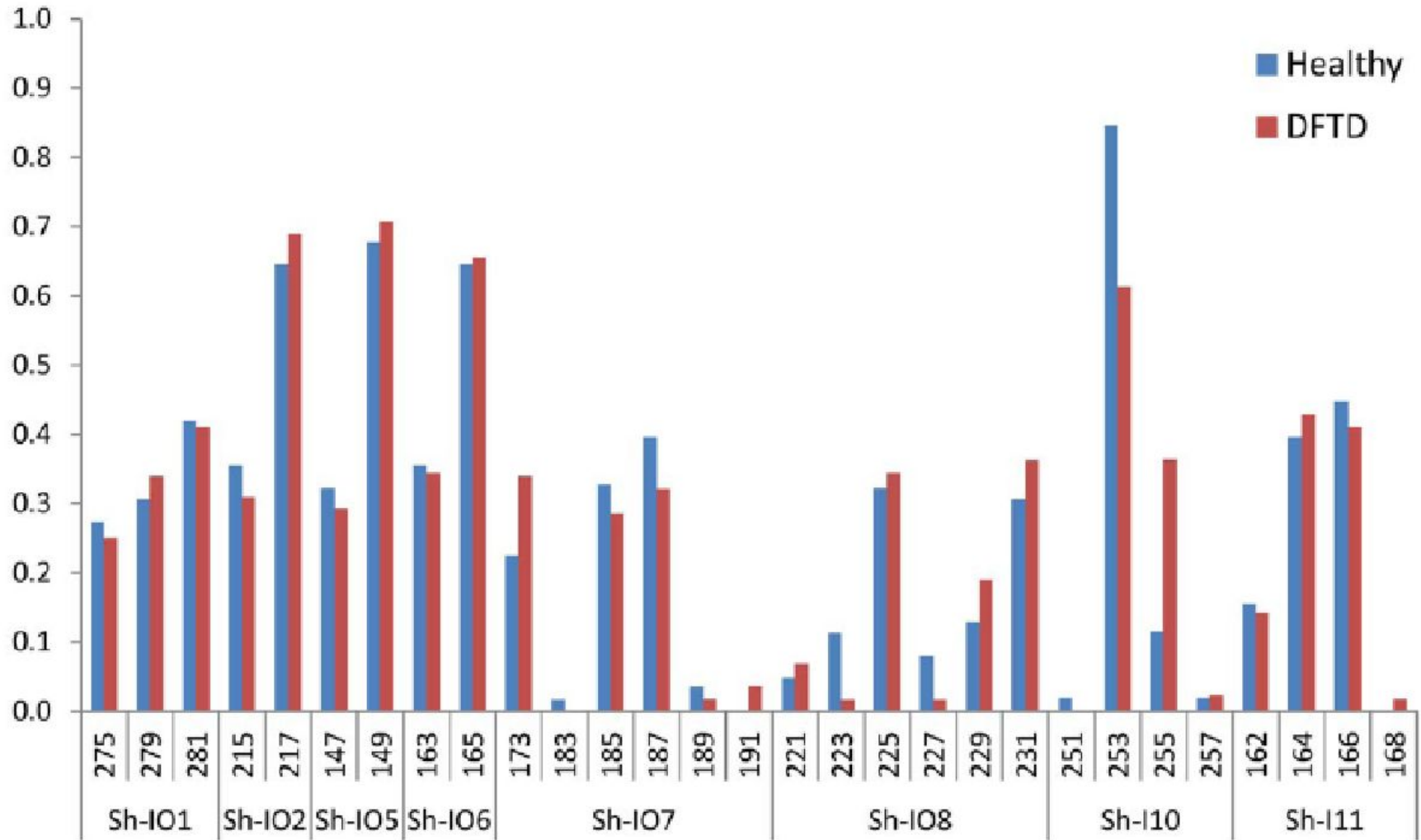


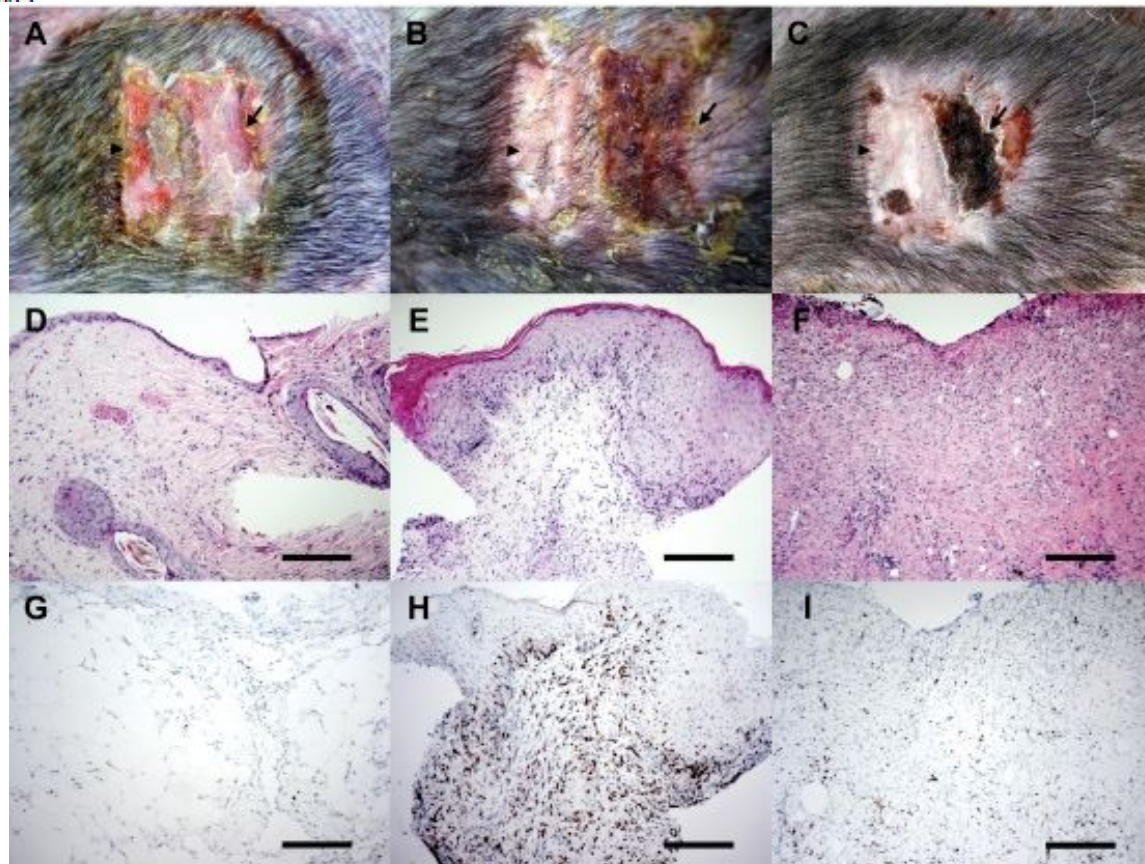
Figure 2. MHC-linked microsatellite loci allele frequencies showing little variation between healthy and DFTD infected devils.

Отторжение аллогraftов у тасманского дьявола

Table 2. Outcome of skin allografts.

| Tasmanian devil ID | Day 7 | Day 14 | Day 21 |
|--------------------|-------------------------------|-----------|--------------------------------|
| TD 190 | Grade II | Grade IV | Necrotic skin, biopsy not done |
| TD 199 | No evidence of rejection | Grade III | Grade IV |
| TD 187 | Unable to determine rejection | Grade IV | Necrotic skin, biopsy not done |
| TD 188 | No evidence of rejection | Grade III | Grade IV |
| TD 191 | No evidence of rejection | Grade III | Necrotic skin, biopsy not done |

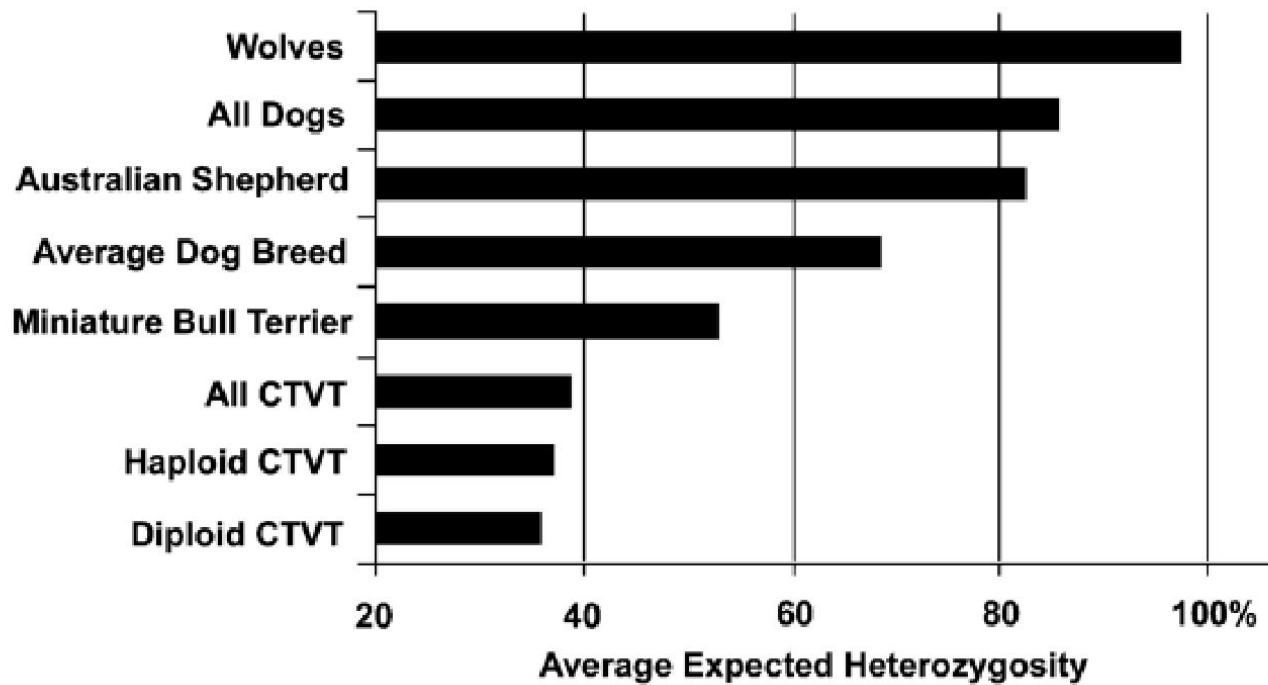
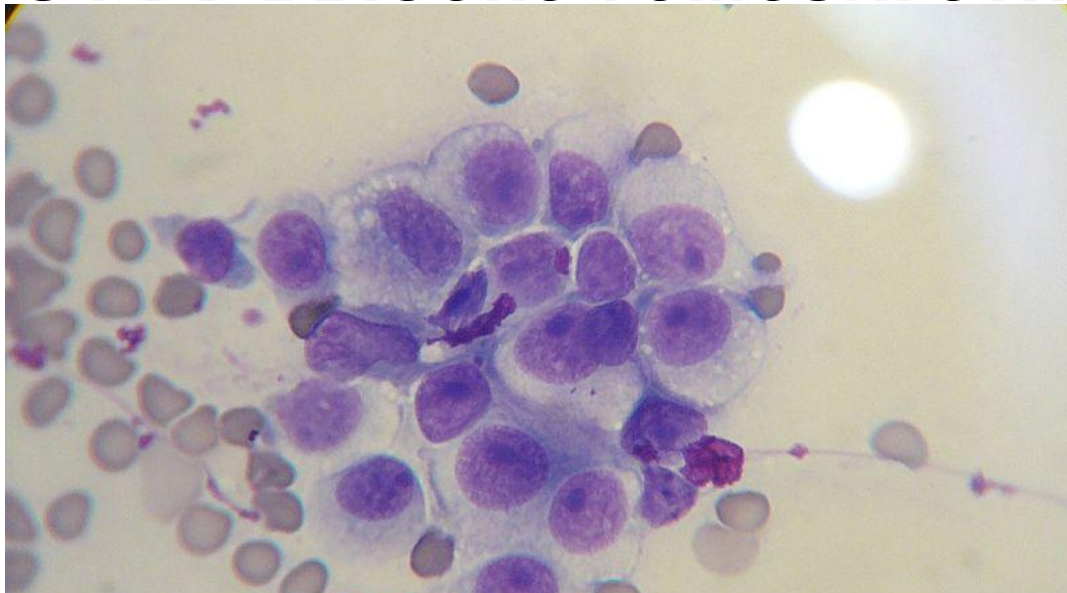
TD 190 was the only devil that showed an early rejection response at Day 7. All five devils had Grade III to Grade IV rejection at Day 14 and Day 21.



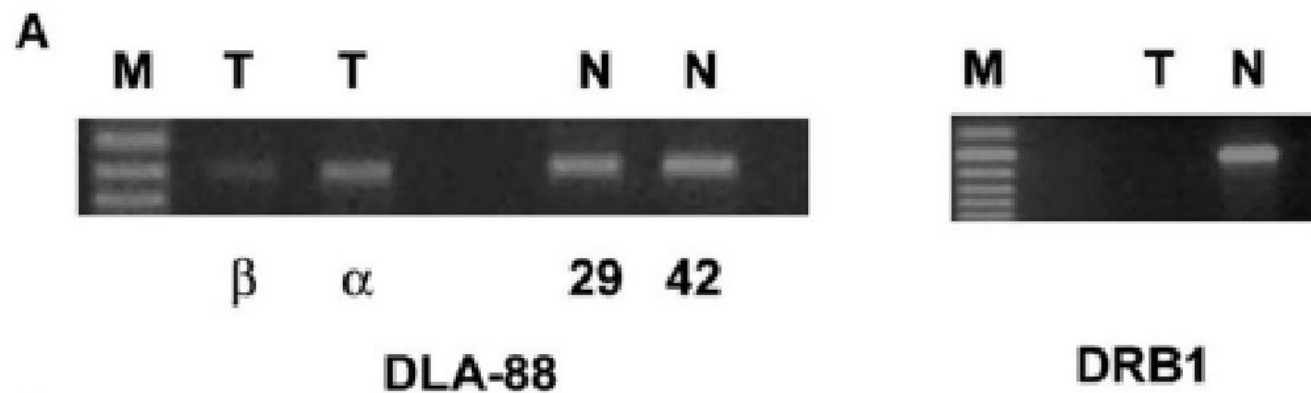
Венерически передаваемая опухоль собак - СТТТ



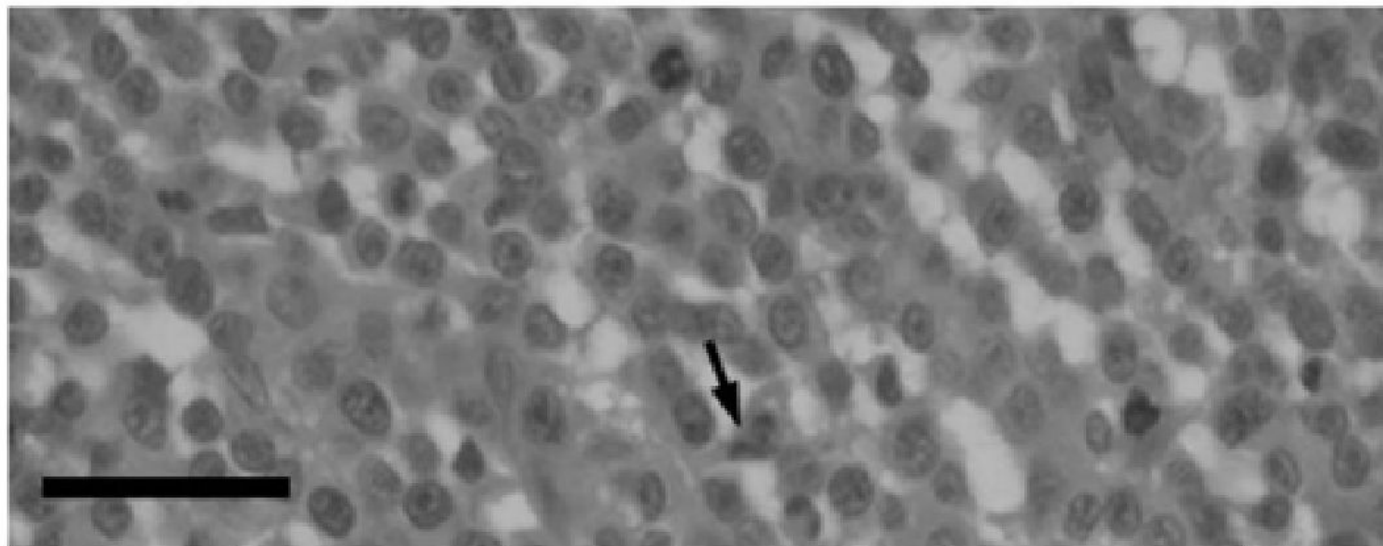
CTVT ВЫСОКО ГОМОЗИГОТНА



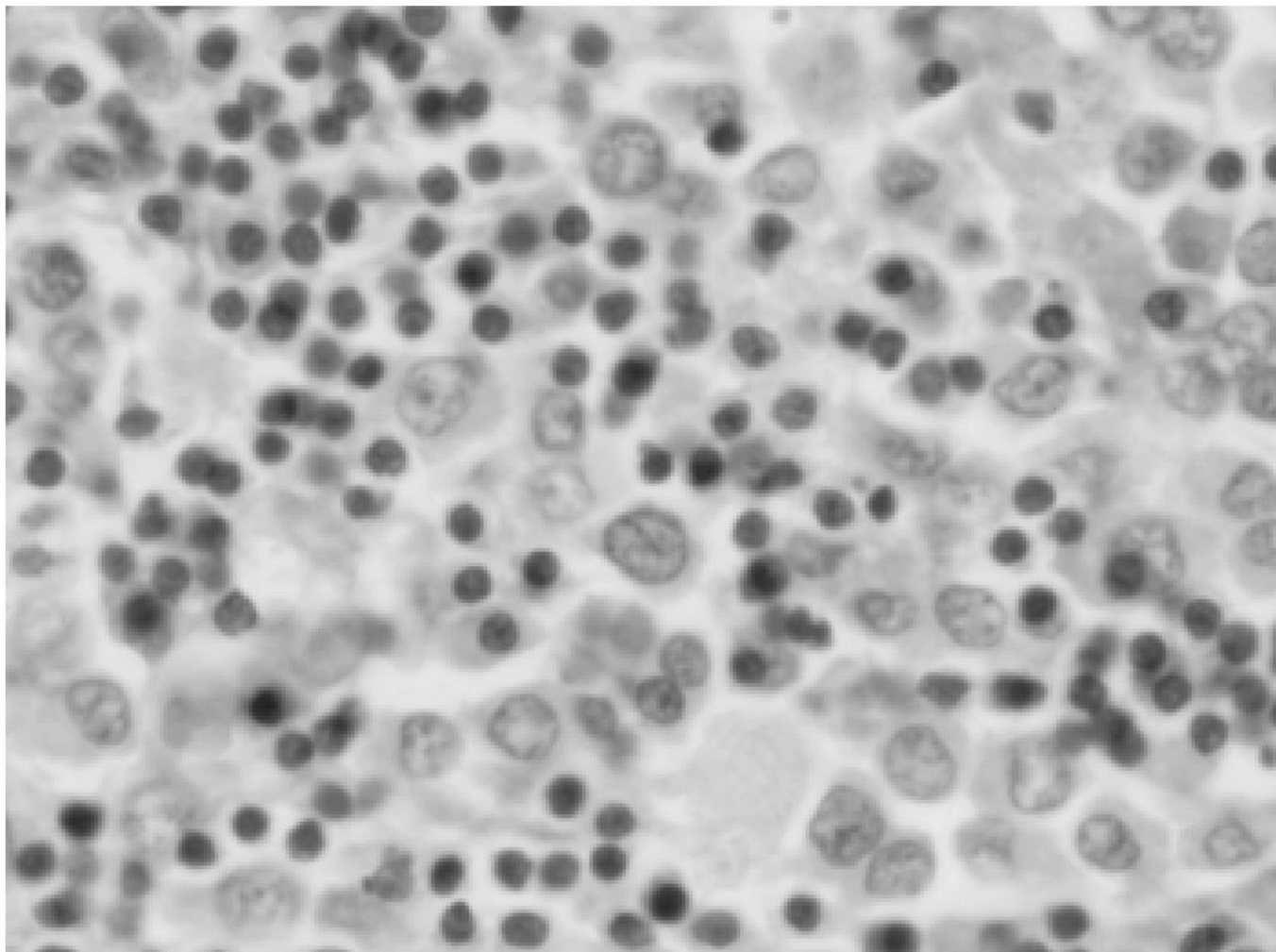
Стадия прогрессии СТТТ и сниженная экспрессия МНС



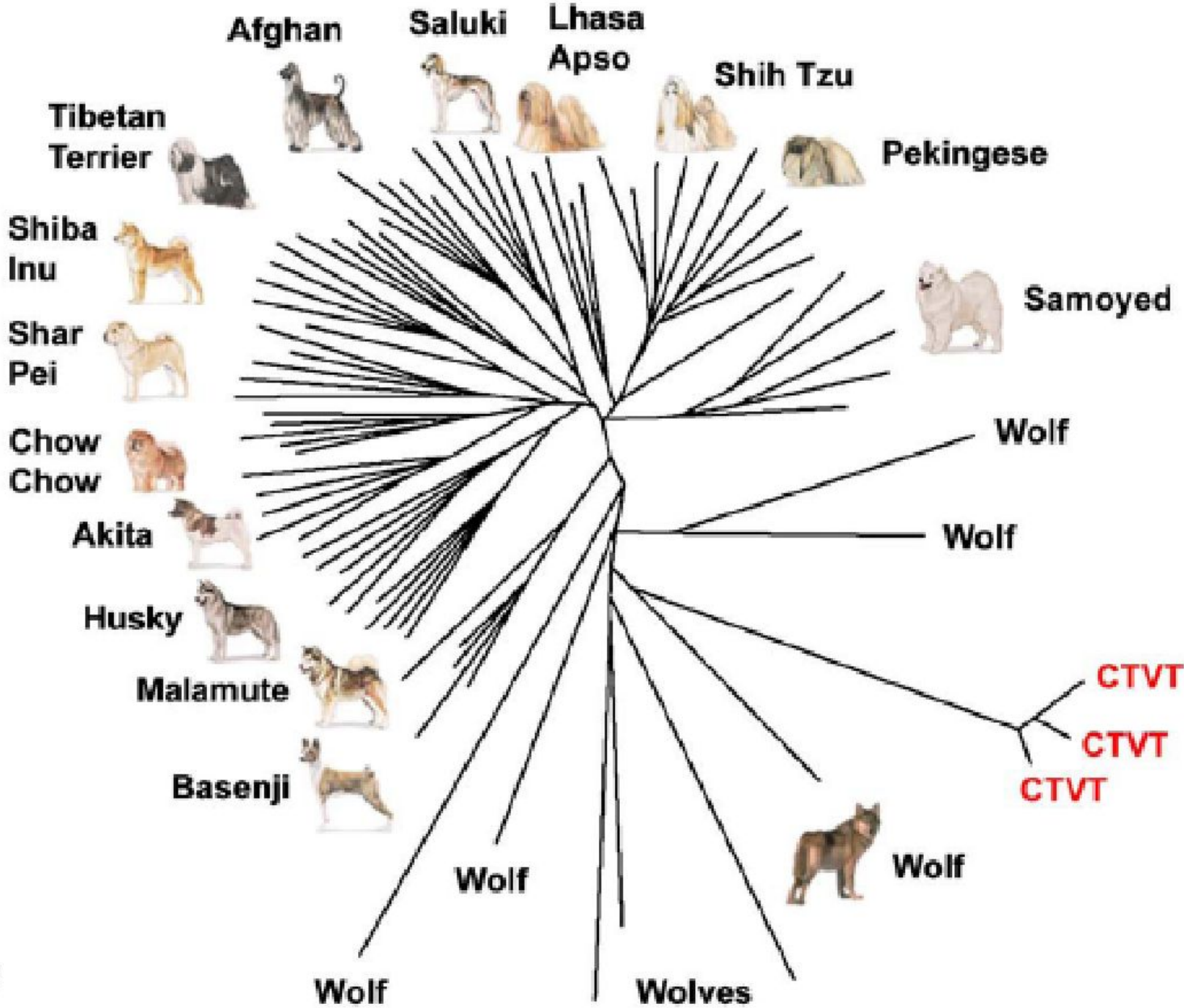
B



Стадия регрессии СТТ



Происхождение СТТТ и передача от волка собаке



B

Саркома Стикера гетерогенна по митохондриальной ДНК

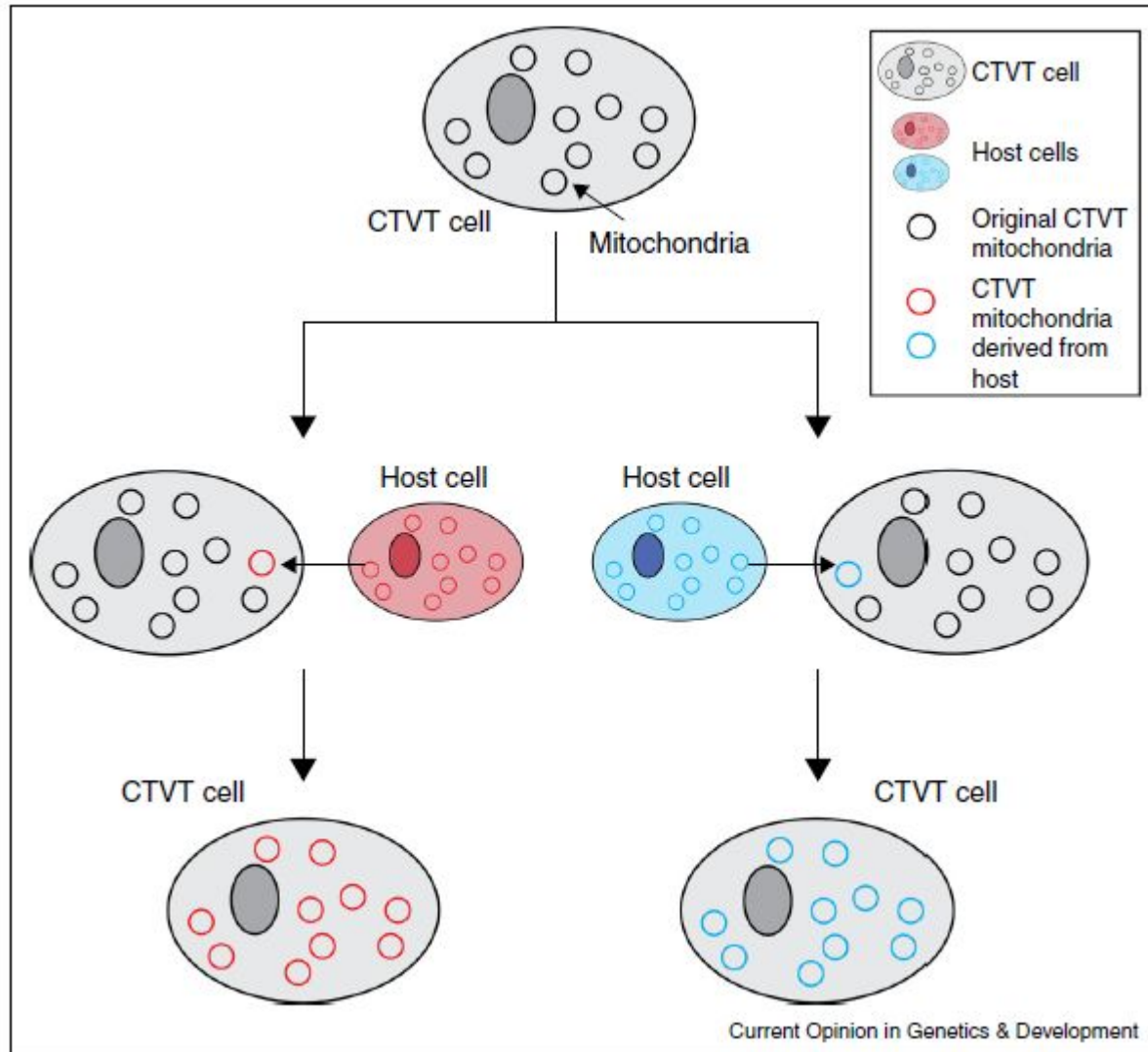
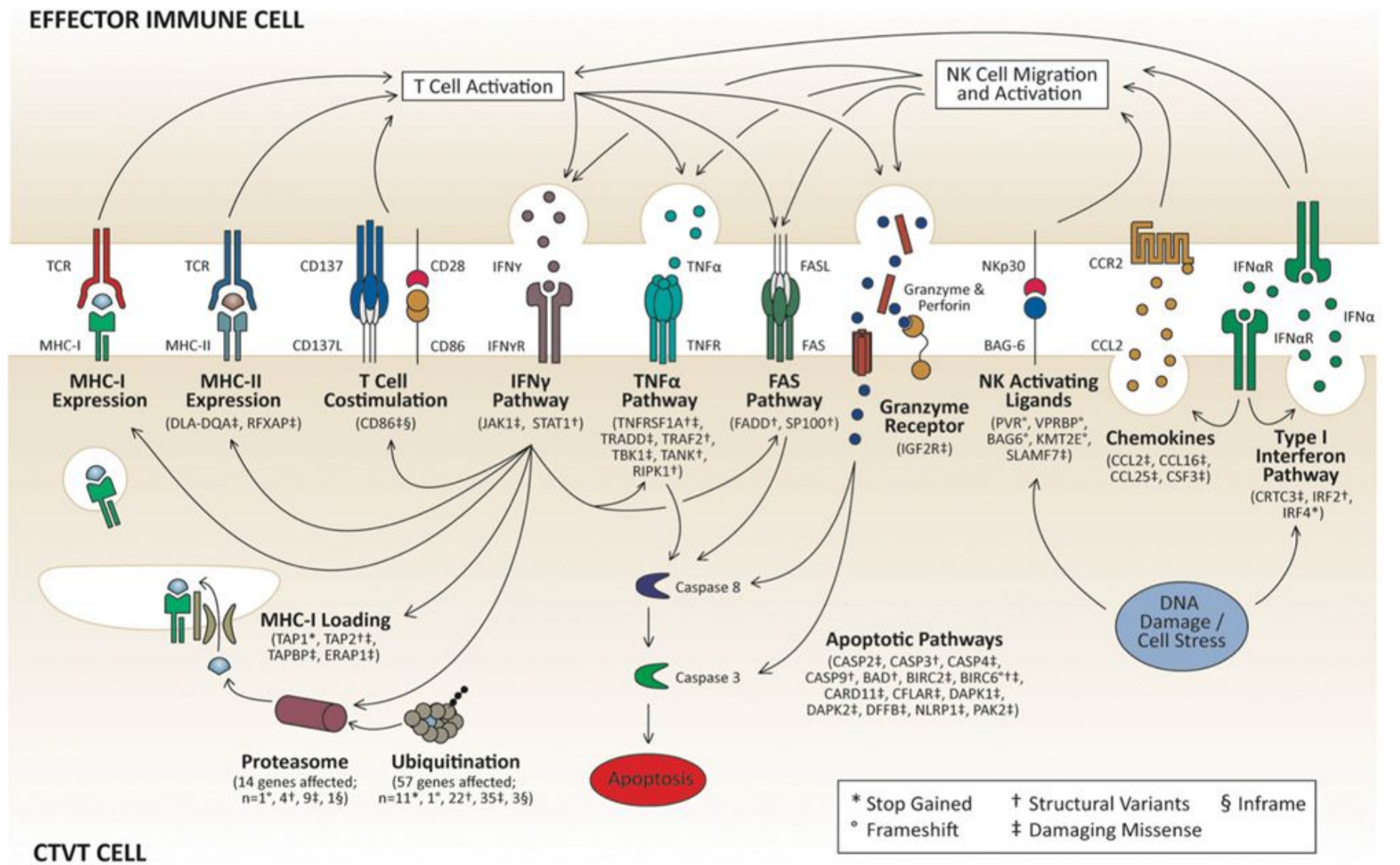


Table 2 Comparison of DFTD and CTVT

| | <i>DFTD</i> | <i>CTVT</i> |
|---------------------------|---|--|
| Host species | Tasmanian devil | Dog |
| Species of origin | Tasmanian devil | Wolf or dog |
| Distribution | Mainland Tasmania (excluding northwest) | Worldwide |
| Time of origin | 15–20 years ago | 7800–78 000 years ago |
| Body location | Face, oral cavity | External genitalia |
| Mode of transfer | Biting | Coitus |
| Histogenesis | Neuroendocrine | Myeloid |
| Metastasis | Common | Common in immune-compromised animals |
| Spontaneous regression | 0% | Common in experimentally inoculated CTVT, prevalence in naturally occurring CTVT unknown |
| Mortality | 100%, within 6–12 months after appearance of symptoms | Rare in experimentally inoculated CTVT, prevalence in naturally occurring untreated CTVT unknown |
| Treatment | None | Chemotherapy, radiation therapy |
| Effect on host population | Host population decline/possible imminent extinction | Probably little effect |

Многие гены СТТВ несут мутации потери функции



На самом деле, устойчивость DFTD вызвана эпигенетическим подавлением экспрессии TAP1, TAP2 и β_2 -микроглобулина, что ведет к снижению у нее количества МНС I

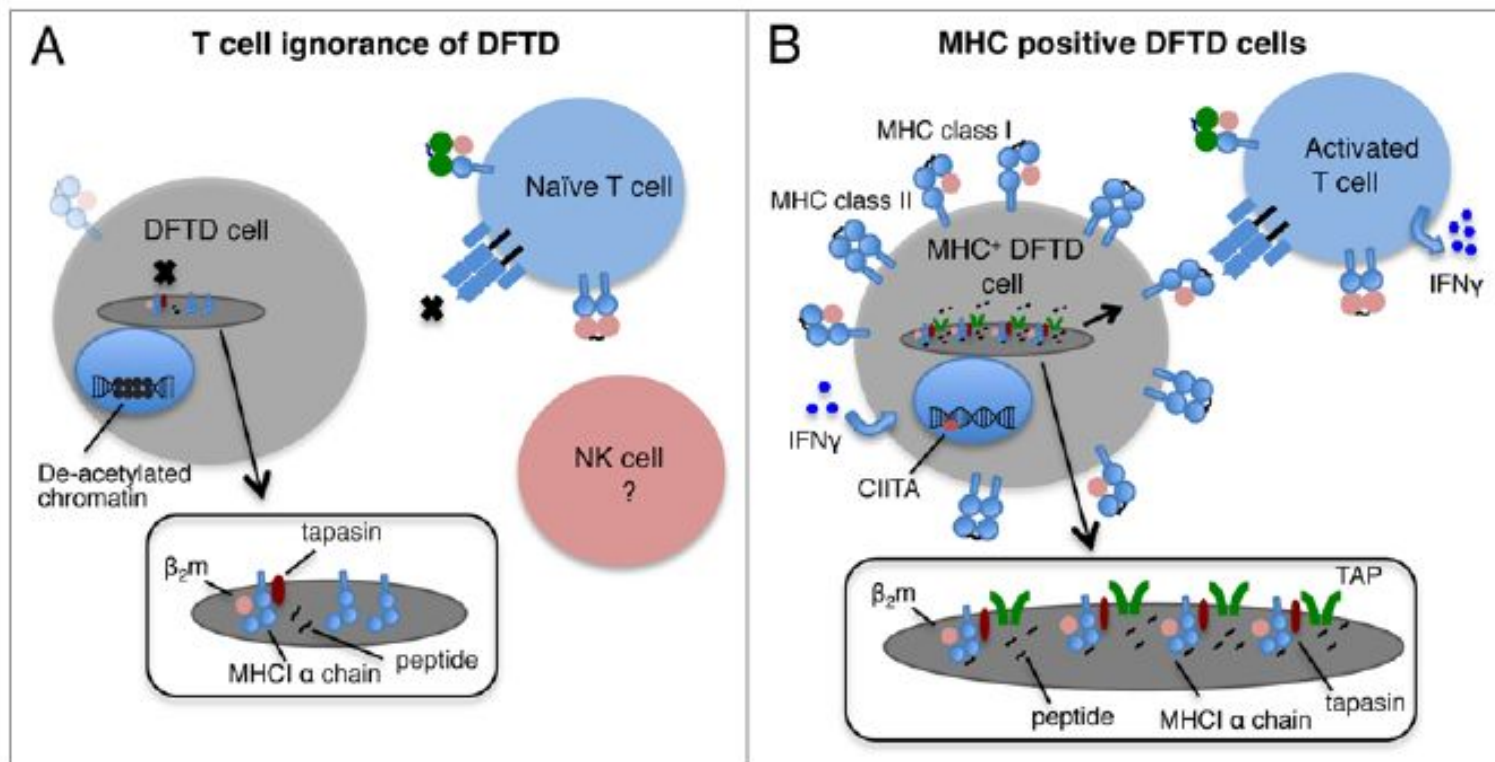
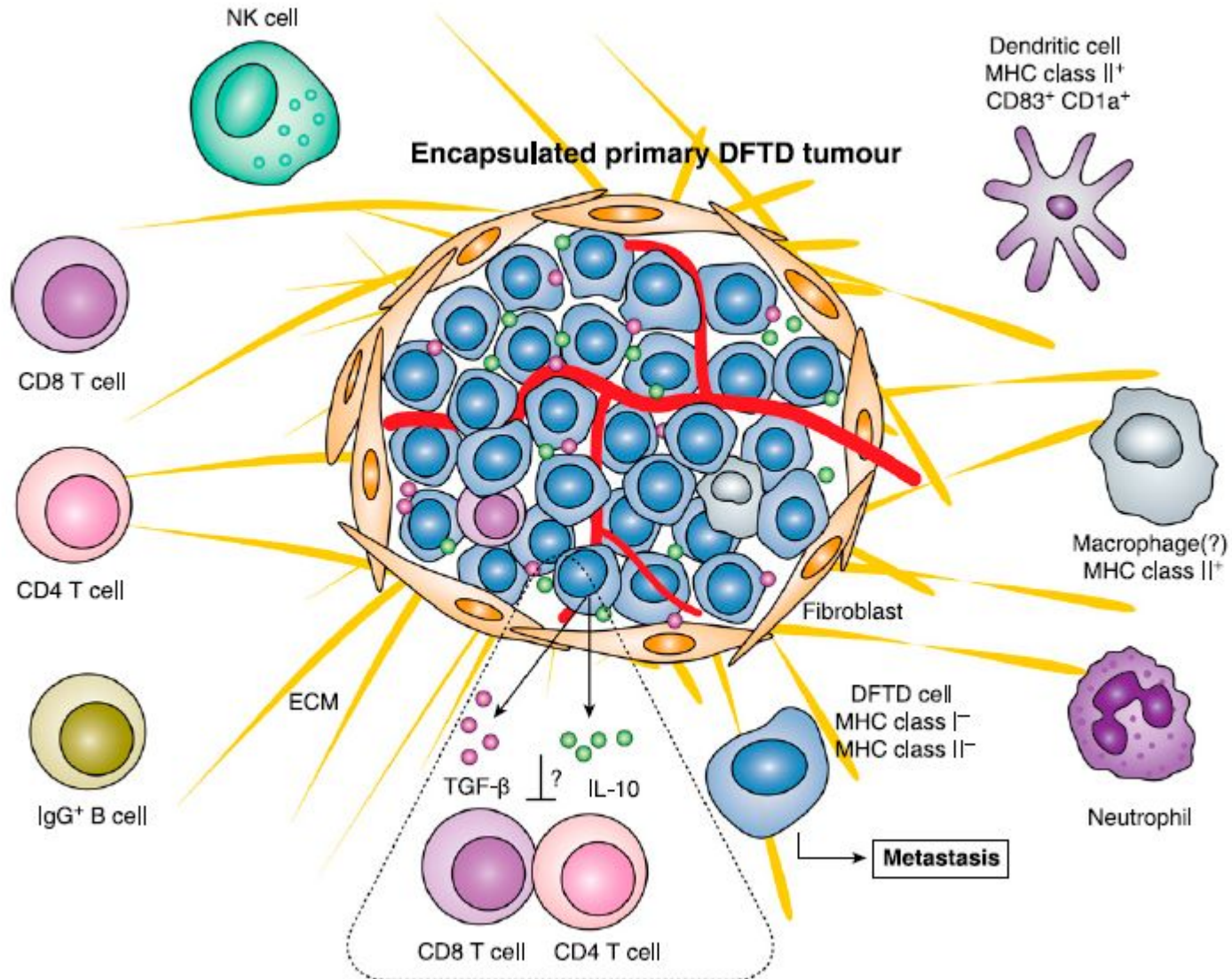
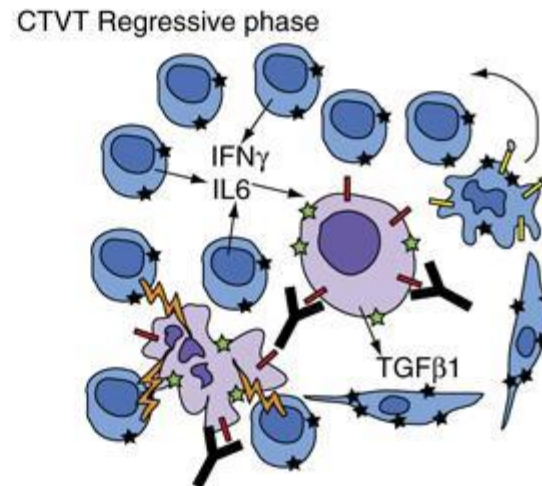
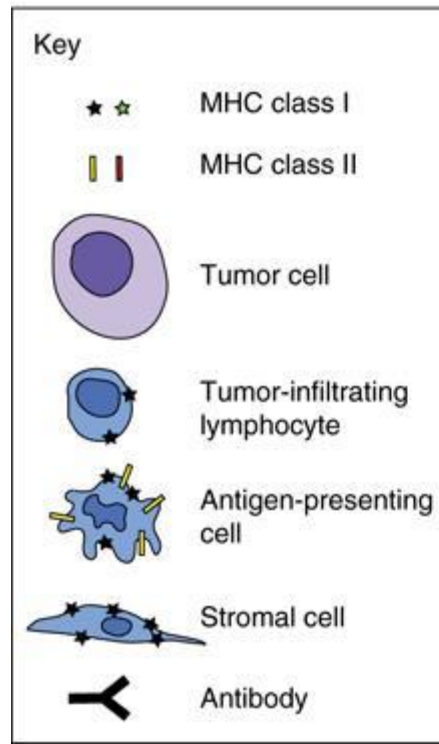
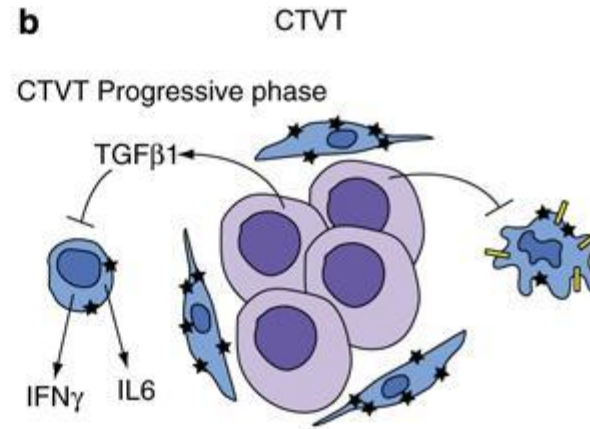
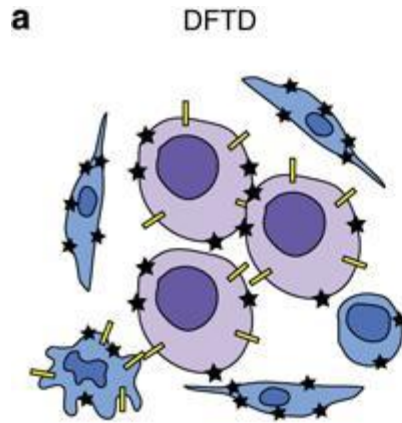


Figure 1. Mechanisms of immune evasion by DFTD cells. (A) Devil T lymphocytes fail to recognize devil facial tumor disease (DFTD) cells as the latter lack MHC molecules on their surface. This is mainly due to the deacetylation-dependent repression of transcription from β_2 -microglobulin (β_2m), transporter associated with antigen presentation (TAP) 1 and TAP2-coding genes. In this situation, MHC Class I heavy chains are produced but retained in the endoplasmic reticulum (ER). Low levels of MHC Class I molecules may be found on the surface of DFTD cells owing to the synthesis of trace amounts of β_2m and to peptides derived from ER-resident proteins. (B) DFTD cells can re-express MHC Class I molecules on their surface. Upon interferon γ (IFN γ) treatment of DFTD cells, β_2m , TAP1, TAP2, MHC Class II molecules and the transcription factor Class II transactivator (CIITA) are upregulated and MHC Class I molecules are expressed on the cell surface. Devils vaccinated with MHC Class I-expressing DFTD cells are expected to activate a protective T-cell response. Insets represent magnified view of the ER.

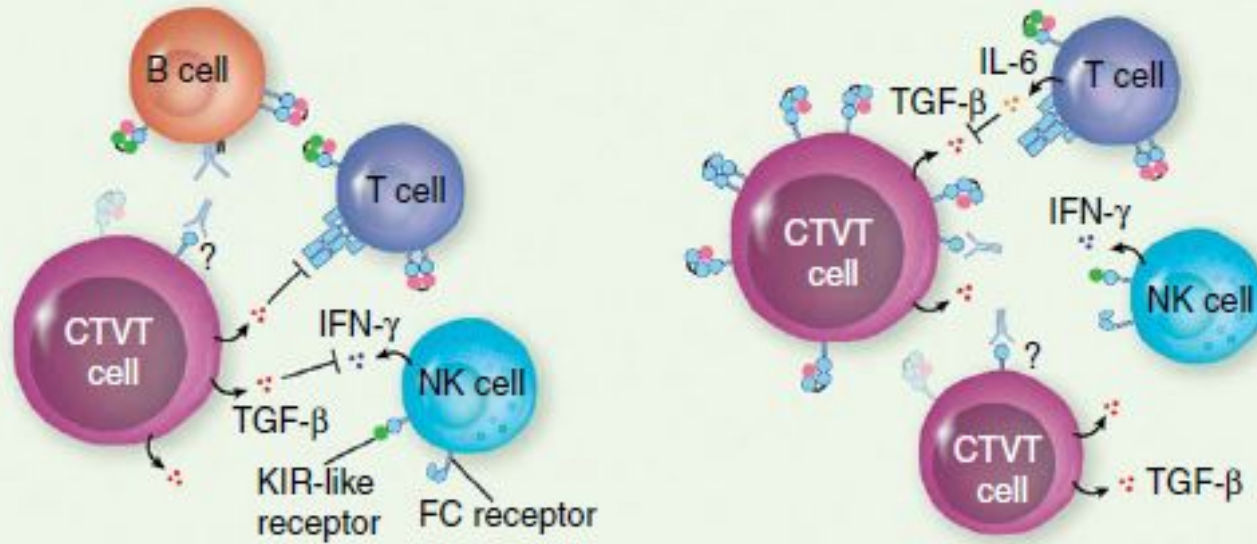
Толерогенное действие клеток DFTD



Сравнение DFTD и CTVT



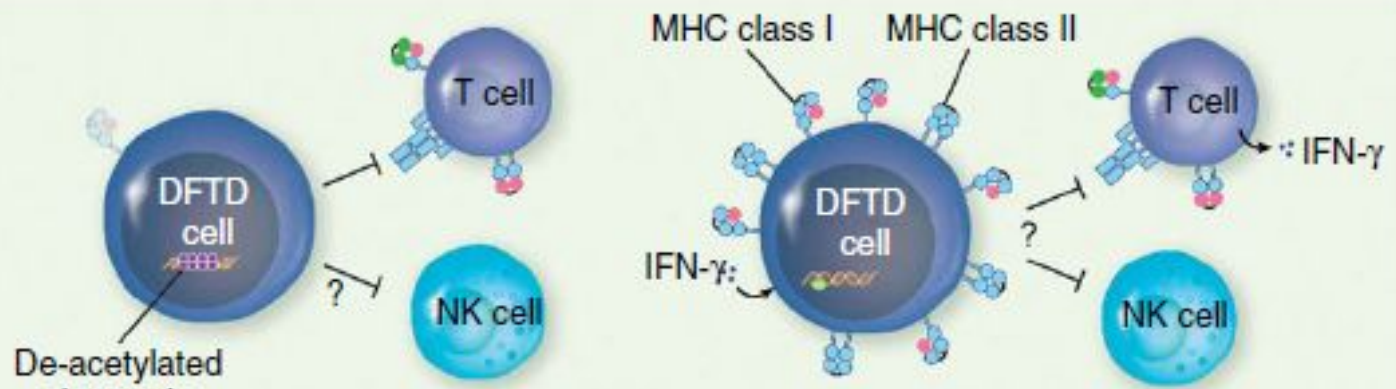
(a) CTVT progression



(i) Growth phase

(ii) Regression phase

(b) DFTD progression



(i) Before IFN- γ treatment

(ii) After IFN- γ treatment

Паразитизм карликовых самцов удильщиков – потенциальная иммунологическая модель?



Ткани самца и самки удильщиков срастаются

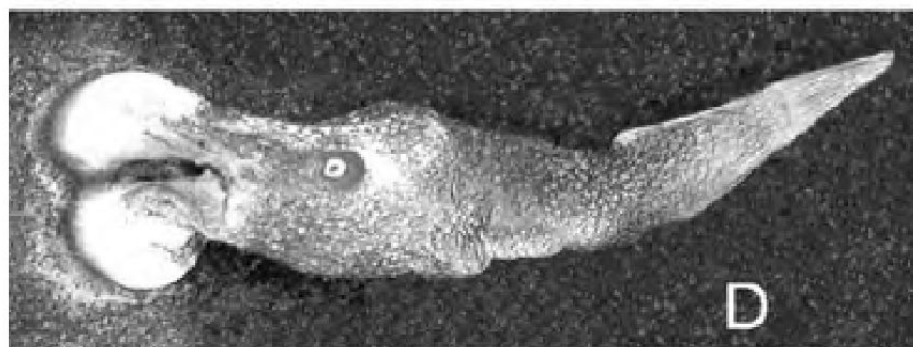


Fig. 8. Parasitic and temporarily attached males. **A** *Caulophryne polynema*, 15 mm, parasitically attached to a 137-mm female, MNHN 2001-140; **B** *Melanocetus murrayi*, 15 mm, temporarily attached (without tissue fusion) to a 73-mm female, BSKU 57842; **C** *Bertella idiomorpha*, 11 mm, parasitically attached to a 77-mm female, UW 48712; **D** *Cryptopsaras couesii*, 10 mm, parasitically attached to a 45-mm female, ARC 8707665; **E** *Neoceratias spinifer*, 18 mm, parasitically attached to a 108-mm female, SIO 70-336

- Следующая лекция 2 апреля – об иммунитете Круглоротых